

UVA Radiation Studying Effect on Morphological Structure, Biochemical and Antioxidant Properties of Yellow Mustard

Abu Bakr El-Bediwi^{1*}, A. Abdelrazek¹, RH. Ebrahim², Hamed M El-Shora³

¹Physics Department, Faculty of Science, Mansoura University, Egypt

²Higher Future Institute of Engineering and Technology, Mansoura, Egypt

³Botany Department, Faculty of Science, Mansoura University, Egypt

* **Corresponding author:** Abu Bakr El-Bediwi, Physics Department, Faculty of Science, Mansoura University, Egypt; Email: baker_elbediwi@yahoo.com

Received date: January 10, 2022, Manuscript No. M- tspa-22-51501; **Editor assigned:** January 13, 2022, PreQC No. P- tspa-22-51501; **Reviewed:** January 25, 2022, QC No. tspa-22-51501; **Revised:** February 4, 2022, Manuscript No. R- tspa-22-51501; **Published date:** February 7, 2022, DOI: 10.37532/2320-6756.2022.10(2).262

Abstract

Plants are living chemical factories for enormous array of the secondary metabolites but the production by plants of compounds useful as medicines or raw materials for manufacture of medicines is influenced by ultraviolet radiation. The aim of research is to investigate the effect of UVA on structure, biochemistry and physiology for yellow mustard. The results show internal structure and chemical composition of yellow mustard changed after exposure to UVA for different times and dissimilar distances. Glutathione and total phenolic in yellow mustard increased but total proline and tocopherol in it decreased after exposed by UVA. Total flavonoids in yellow mustard varied after exposure to UVA for different time and dissimilar distances.

Keywords: Phenolic; Glutathione; DPPH scavenging activity; Proline; Yellow mustard; UVA

Introduction

Solar radiation is a complex mixture of ultraviolet, visible light and infrared wavelengths. UVA radiation (315 nm-400 nm) is a component of solar radiation. Plants are living chemical factories for the biosynthesis of a huge array of the secondary metabolites. Plants are used medicinally in different countries and are a source of many powerful drugs. The World Health Organization stated that about 80% of the world's population depends mainly on traditional medicine that mainly includes the use of plant extracts. UV light is an important abiotic elicitor, and had use in phytochemical production in a variety of plant cultures in the past [1]. Exposure to UV light stress causes stimulation of defense mechanisms in plants, thus, producing commercially important secondary compounds [2]. Some reports discussed the effect of low levels of UV radiation on plant growth [3,4]. There are many beneficial uses of radiation that offer few risks when properly employed. The exposure to radiations can have stimulatory effects on specific morphological parameters. The mustard plant belongs to the Cruciferae (*Brassicaceae*) family, used in medicine is external as a liniment and relieving pain from bruises or a stiff neck and relieving colic and respiratory problems.

Citation: Abu Bakr El-Bediwi. UVA Radiation Studying Effect on Morphological Structure, Biochemical and Antioxidant Properties of Yellow Mustard, J Phys Astron.2022;10(2):262.

Growth behavior, secondary metabolites and vitamins of *Nigella Sativa* and *garden cress* changed after exposure by UVC [5,6]. UVs have adequate energy to break the chemical bonds causing photochemical reactions and inducing changes in plant metabolic enzyme, subsequently trigger the production of secondary metabolites [7-9]. Effect of UV is varied with duration and irradiation intensity. The objective of this work is to assess the effect of UVA radiation on structure, non-enzymatic and enzymatic antioxidants for yellow mustard.

Experimental Methods

Structure measurements

Internal structure and molecular structure of yellow mustard are studied by Shimadzu X-ray diffractometer, (Dx-30, Japan), scanning electron microscope (JEOL JSM-6510LV, Japan) and Nicolet™ iS™ 10 FT-IR Spectrometer from USA.

GSH determination

GSH is determined using UV/V spectrophotometer, Jenway, England.

DPPH determination

Concentration ranging from 0.4g/100g to 2g/100g are prepared with methanol from each sample (100 µl) extract and DPPH radical (100 µl, 2Mm) dissolved in methanol. The mixture is stirred and left to stand for 15 min in dark. Then the absorbance is measured at 517 nm against a blank. Percentage scavenging effect is calculated as $[(A_0 - A_1) / A_0] \times 100$ where A_0 is the absorbance of the control (without sample) and A_1 is the absorbance in the presence of the sample.

Total phenols determination

Total phenols are determined colorimetric by Folin-Ciocalteu reagent total phenolic content is calculated from the regression equation of the standard plot ($y = 3.005x - 993.56, r^2 = 0.9974$) and are expressed as mg Gallic acid equivalent/100g sample.

Flavonoid determination

Aluminum chloride colorimetric method is used to determine flavonoid content. 1 ml of sample extract is mixed with 3 ml of methanol, 0.2 ml of 10% aluminum chloride. 0.2 ml of 1 M potassium acetate and 5.6 ml of distilled water and remains at room temperature for 30 minutes. The absorbance is measured at 420 nm. Rutin is used as standard (1mg/ml). Flavonoid content is calculated from the regression equation of the standard plot [$y = 16.122x - 340.23, r^2 = 0.9777$] and are expressed as (mg Rutin equivalent/100g sample).

Proline determination

The proline concentration is determined after extraction with 3% (W/V) aqueous sulfosalicylic acid from a standard curve using D-Proline ($y = 36.738x + 1.2739, r^2 = 0.9777$) which give by used UV/V spectrophotometer, Jenway, England.

Results and Discussions

Internal structure

FIG.1 shows x-ray diffraction patterns of normal yellow mustard after exposure to UVA for different period times at dissimilar distances. There is a change in the main peak of yellow mustard, **TABLE 1**, as intensity, broadness, started base line and area under the peak after exposure to UVA at 5 cm and 20 cm for 1 hour and 4 hours. That is because the interaction of UVA with atoms or molecules break or modify bonds and or make rearrangement it. Also UV radiation stress is caused a change in bond structure. The interaction with atoms or molecules in the cell, particularly water, produce free radicals [10] which can damage or modify important components of plant cells, affected differentially the morphology, anatomy, biochemistry and physiology for plants [11]. Scanning electron micrographs show different features and qualities of the main cell and a round molecules as show in **FIG. 2**. These changed confirmed the x-ray and IR results.

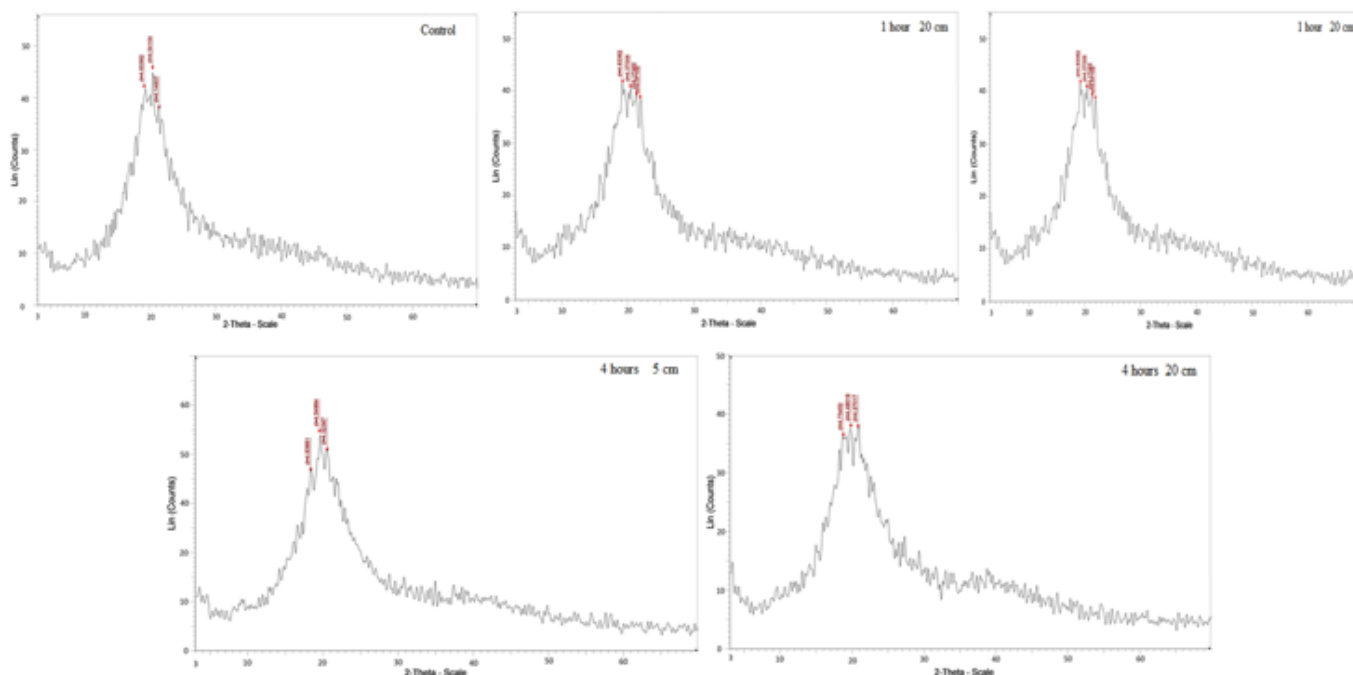


FIG.1. X-ray diffraction patterns of yellow mustard after exposure to UVA.

Exposure time (hour)	Area under the peak		Width of the peak	
	Exposure at 5 cm	Exposure at 20 cm	Exposure at 5 cm	Exposure at 20 cm
Zero (Control)	4.61			
S.No.	Exposure at 5 cm	Exposure at 20 cm	Exposure at 5 cm	Exposure at 20 cm
1 hour	6.1404938	4.5237738	7.3240309	6.7581327
4 hours	4.3627724	5.210452	6.2214994	6.4209859

TABLE 1. X-ray analysis of yellow mustard before and after exposed to UVA.

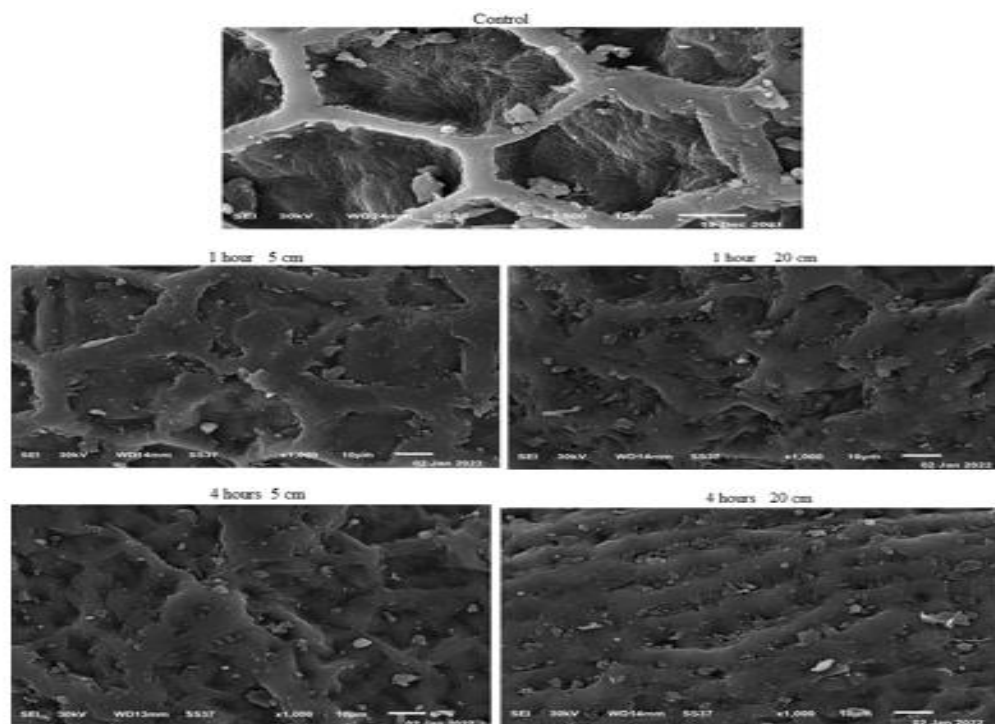


FIG.2. SEM of normal yellow mustard and after exposed to UVA.

Molecular Structure

FIG. 3 shows IR spectrum of yellow mustard, a plot of wave number (X-axis) vs. present transmittance (Y-axis). IR analysis of yellow mustard listed in TABLE 2 show transmittance the intensity increased after exposure to UVA at 5 cm for 1 hour and 4 hours but decreased at 20 cm for 1 hour and 4 hours. A significant change occurred in the main peak position, O-H, after exposure to UVA for 1 hour at 5 cm, but a little variation is occurred during other exposure times at 5 cm and 20 cm distance. That is mean, the absorption of UVA in cell, break or modify the position or degraded some molecular bond or switching off of the transcription-translation machinery during radiation exposure [12].

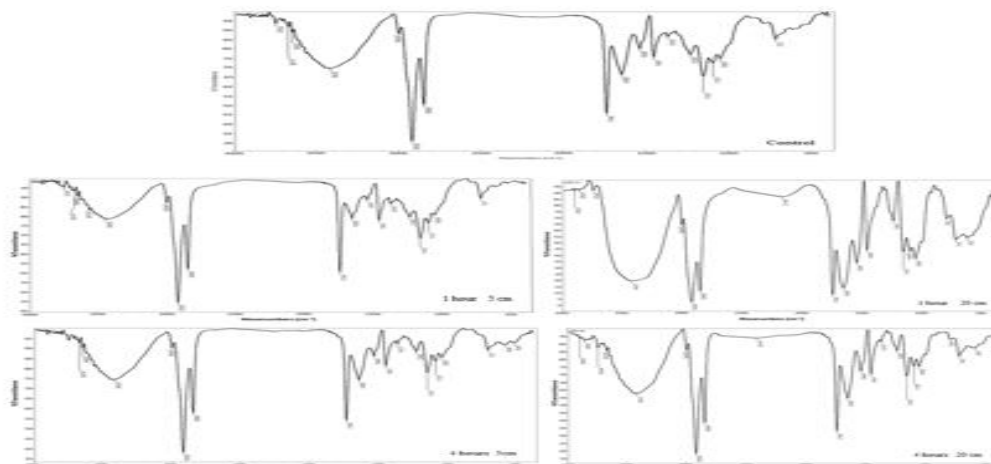


FIG.3. IR spectrum of yellow mustard after exposure to UVA.

Control (untreated sample)		
Position	Intensity %	Band
1746	45.6	C-O
2925	31.04	C-H
3422	69.4	O-H
1 hour at 5 cm		
1746	50.4	C-O
2925	34.36	C-H
3450	78.4	O-H
1 hour at 20 cm		
1746	8.32	C- O
2925	2.75	C-H
3421	19.22	O-H
4 hour at 5 cm		
1746	54.12	C- O
2925	38.11	C-H
3423	74.26	O-H
4 hour at 20 cm		
1746	32.9	C- O
2925	18.21	C-H
3421	57.37	O-H

TABLE 2. IR spectrum analysis of yellow mustard after exposure to UVA.

Glutathione Content

Glutathione content for yellow mustard is increased after exposure to UVA at 5 cm and 20 cm for different period of times as seen in **TABLE 3** and **FIG.4**. The results show it is increased by 17.44%, 19.3%, 7.8% and 4.58% and, by 14.68%, 19.3%, 0.45% and 5.5% at 5 cm and 20 cm distance far from UVA source for different period times. That is mean, it increased from 0.45% to 19.3% after exposure to UVA, because the glutathione, which one of the most important non-enzymatic antioxidants, is positively affected by radiation as it is a hermetic type of response under applied doses of radiation [13, 14].

Exposure time (hour)	GSH (mg/ 100g)	
Zero (Control)	72.66	
	Irradiated at 5 cm	Irradiated at 20 cm
1 hour	85.33	83.33
2 hours	86.66	86.66
3 hours	78.33	72.99
4 hours	75.99	76.66

TABLE 3. **Glutathione content of yellow mustard after exposed to UVA.**

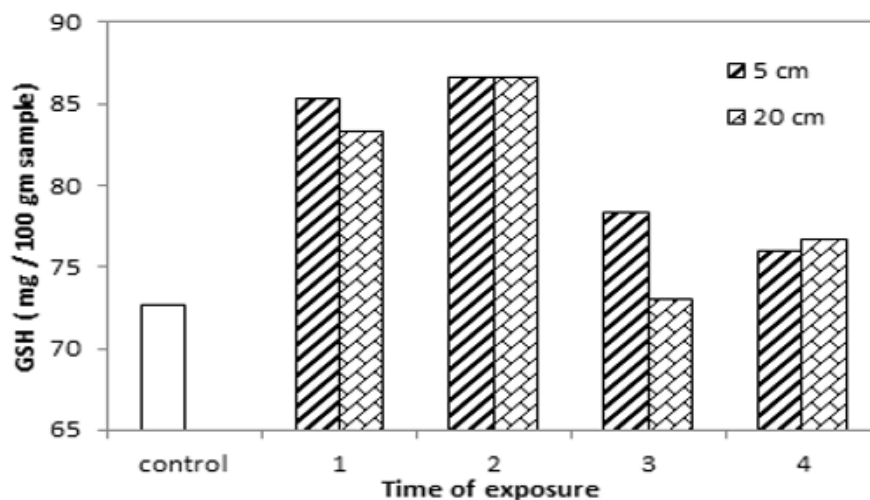


FIG.4. **Glutathione in yellow mustard after exposed to UVA.**

Phenolic Content

Total phenolic content for yellow mustard increased by 32.57%, 43.5%, 19.5% and 48.35% and by 18.17%, 8.31%, 59.71% to 1.82.5% after exposure to UVA at distances 5 cm and 20 cm for 1 hour, 2 hours, 3 hours and 4 hours as shown in **TABLE 4** and **FIG.5**. That is because UV radiation increases the accumulation of phenolic compounds along with antioxidant properties. Also phenolic compounds are plant secondary metabolites that hold an aromatic ring bearing at least one hydroxyl groups and IR analysis for yellow mustard show there is a change in the intensity and position of O-H band caused increased in phenolic. The pervious results also shown phenolic compounds is increased or induced after UV radiation exposure during different period times [15-19].

Exposure time (hour)	Total phenolic (mg/ 100g)	
Zero (Control)	409.48	
	Irradiated at 5 cm	Irradiated at 20 cm
1 hour	542.86	483.89
2 hours	587.57	443.50
3 hours	489.33	654.21
4 hours	607.44	416.92

TABLE 4. **Total phenolic content of yellow mustard exposed to UVA.**

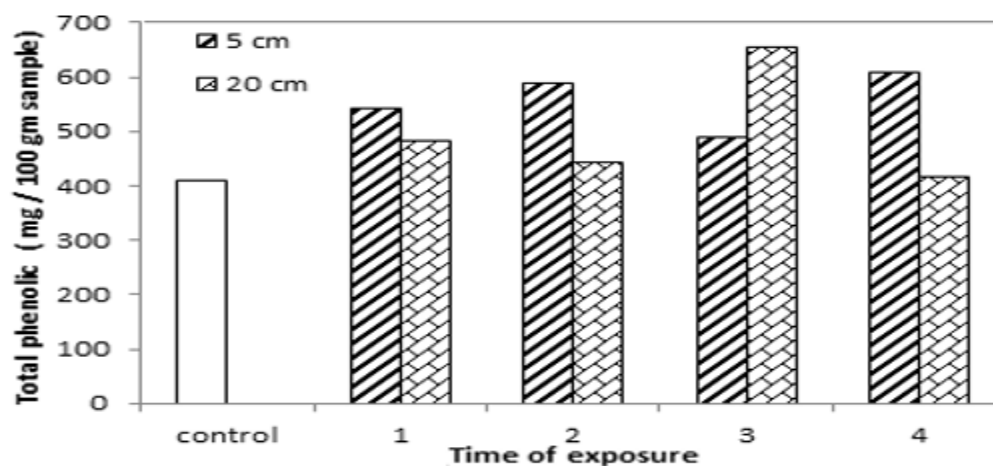


FIG.5. Total phenolic content in yellow mustard exposed to UVA.

Flavonoids Content

Flavonoid is a term that is a bit ambiguous literally it means flavone-like compound. **TABLE 5** and **FIG.6** show total flavonoids in yellow mustard decrease after exposure for 1 hour, 3 hours and 4 hours and 1 hour and 3 hours at 5 cm and 20 cm from UVA source but it increased after exposure for 2 hours at 5 cm distance and 2 hours and 4 hours at 20 cm. That is mean, flavonoids content in yellow mustard varied after exposed to UVA for different times and distances because flavonoids is plant secondary metabolites that hold an aromatic ring bearing at least one hydroxyl groups, the results, IR analysis, showed a change in hydroxyl group such as intensity and position.

Exposure time (hour)	Total flavonoids (mg/ 100g)	
Zero (Control)	297.6	
	Irradiated at 5 cm	Irradiated at 20 cm
1 hour	148.45	141.95
2 hours	395.71	446.75
3 hours	198.54	236.8
4 hours	183.07	422.64

TABLE 5. Total flavonoids content of yellow mustard exposed to UVA.

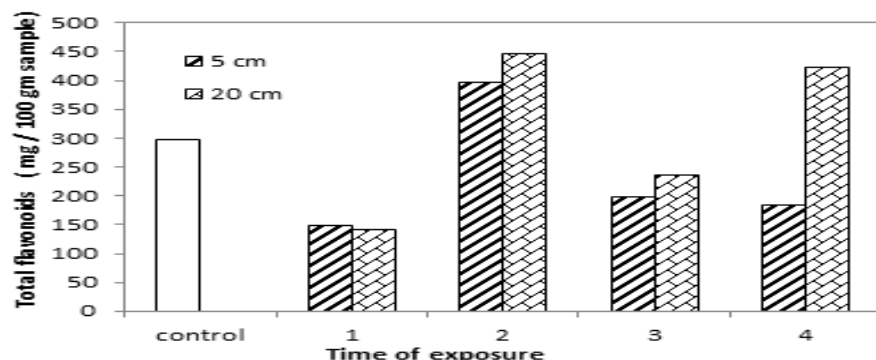


FIG.6. Total flavonoids content in yellow mustard exposed to UVA.

Total Proline Content

Proline plays important roles in protein synthesis and structure, metabolism and nutrition. Total proline content in yellow mustard decreased after exposure to UVA for different period times and distances as shown in **TABLE 6** and **FIG.7**. The chemical composition of yellow mustard such as protein changed after exposure to UVA. That is mean, total proline changed because proline is a substrate for protein synthesis. Also proline accumulation is response to biotic and abiotic stresses caused to the effect of UVA.

Exposure time (hour)	Total proline (mg/ 100g)	
Zero (Control)	925.84	
	Irradiated at 5 cm	Irradiated at 20 cm
1 hour	889.96	899.24
2 hours	883.56	862.40
3 hours	917.54	911.96
4 hours	914.41	888.98

TABLE 6. Total proline content of yellow mustard exposed to UVA at 5 and 20 cm for different period times.

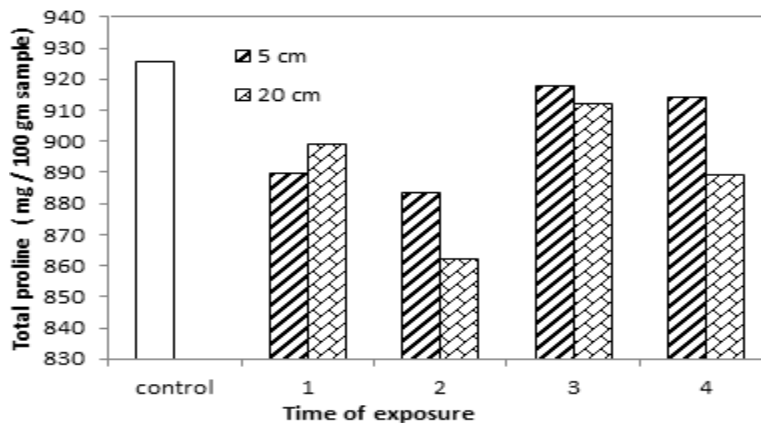


FIG.7. Total proline content in yellow mustard exposed to UVA.

DPPH Scavenging Activity

UVR carries higher energy than visible light and its effects on tissues include gene mutations, DNA damage, oxidative stress immunosuppression, and inflammatory responses. Therefore, some compounds increase and others may decrease due to abiotic stress as pathways for secondary metabolite production are interrelated. DPPH scavenging activity (%) for yellow mustard decreased after exposure to UVA for 1 hour, 2 hours, 3 hours and 4 hours at 5 cm and 20 cm as shown in **TABLE 7** and **FIG.8**.

Exposure time (hour)	DPPH scavenging activity (%) 5 cm		
	0.40%	1%	2%
Zero (Control)	53.9	64.89	85.82
1 hour	30.56	58.33	67.01
2 hours	26.67	39.56	62.85
3 hours	27.8	39.03	47.43
4 hours	26.04	40.6	60.76
Exposure time (hour)	DPPH scavenging activity (%) 20 cm		
	0.40%	1%	2%
Zero (Control)	53.9	64.89	85.82
1 hour	23.14	31.25	57.29
2 hours	29.51	51.39	68.75
3 hours	29.17	35.07	64.24
4 hours	24.31	34.03	63.54

TABLE 7. DPPH scavenging activity (%) of yellow mustard exposed to UVA at 5 cm and 20 cm for different period times.

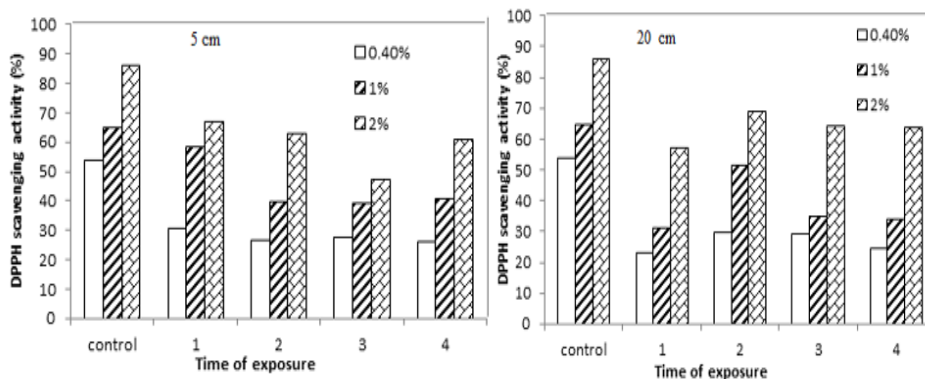


FIG.8. DPPH scavenging activity of yellow mustard exposed to UVA.

Tocopherol Content

TABLE 8 and **FIG.9** show, tocopherol content in yellow mustard decreased by 10.1%, 6.01%, 47.5% and 42.6% after exposure to UVA at 5 cm for 1 hour, 2 hours, 3 hours and 4 hours. Also it decreased gradually after exposure for 1 hour, 2 hours, 3 hours and 4 hours at 20 cm.

Exposure time (hour)	Tocopherol (mg/kg)	
Zero (Control)	2029.19	
	Irradiated at 5 cm	Irradiated at 20 cm
1 hour	1824.13	1965.43
2 hours	1907.79	1726.63
3 hours	1065.43	1335.00
4 hours	1165.43	1234.16

TABLE 8. Tocopherol of yellow mustard exposed to UVA.

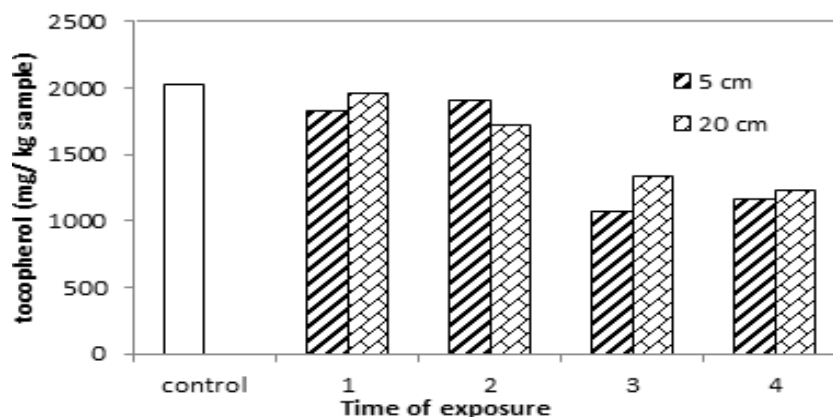


FIG.9. Tocopherol content in yellow mustard exposed to UVA.

Conclusion

Variation in radiation resulted in, significant differences in biomass accumulation, protein and total non-structural carbohydrates contents in plants. Exposure to UVA improve/or disprove some antioxidants for yellow mustard. Also UVA caused a change in yellow mustard internal structure.

REFERENCES

1. Xuan TD, Khanh TD, Khang DT, et.al. Changes in chemical composition, total phenolics and antioxidant activity of *Alpinia zerumbet* leaves exposed to UV. Int. Lett. Nat. Sci. 2016;55 25-34.
2. Yin X, Singer SD, Qiao H, et.al. Insights into the mechanisms underlying ultraviolet-C induced resveratrol metabolism in grapevine (*V. amurensis* Rupr).“Tonghua-3”. Front. Plant Sci. 2016;7:503.
3. Hideg E, Jansen MA, Strid A. Melatonin and reactive oxygen and nitrogen species: a model for the plant redox network. Trends Plant Sci. 2013;18:107-115.

4. Bornman JF, Barnes PW, Robinson SA, et.al. Solar ultraviolet radiation and ozone depletion-driven climate change: Effects on terrestrial ecosystems. Photochem Photobiol Sci. 2015;14(1):88-107.
5. El-Bediwi AB, Hasanin S, Abdelrazek A, El-Shora HM. Effect of Ultraviolet on Morphological and Secondary Metabolites Content of Garden Cress. 2018;4:187-194.
6. Zhang WJ, Björn LO. The effect of ultraviolet radiation on the accumulation of medicinal compounds in plants. Fitoterapia. 2009;80(4):207-218.
7. Hectors K, Van Oevelen S, Geuns J, et.al. Dynamic changes in plant secondary metabolites during UV acclimation in Arabidopsis thaliana. Physiologia Plantarum. 2014;152(2):219-230.
8. Ghasemi S, Kumleh HH, Kordrostami M. Changes in the expression of some genes involved in the biosynthesis of secondary metabolites in Cuminum cyminum L. under UV stress. Protoplasma. 2019;256(1):279-290.
9. Kovacs E, Keresztes A. Effect of gamma and UV-B/C radiation on plant cells. Micron. 2002;33(2):199-210.
10. Ashraf MU, Cheema AA, Rashid MU, et.al. Effect of gamma rays on M-1 generation in basmati rice. Pak J Bot. 2004;35:791-796.
11. Sen Raychaudhuri S, Deng XW. The role of superoxide dismutase in combating oxidative stress in higher plants. Bot Rev. 2000;66(1):89-98.
12. Stajner D, Popovic BM, Taski K. Effects of γ -irradiation on antioxidant activity in soybean seeds. Cent Eur J Biol. 2009;4(3):381-386.
13. Chakravarty B, Sen S. Enhancement of regeneration potential and variability by γ -irradiation in cultured cells of Scilla indica. Biol plant. 2001;44(2):189-193.
14. Surjadinata BB, DA Jacobo-Velázquez, L Cisneros-Zevallos. Biophysical methods used to generate tolerance to drought stress in seeds and plants: a review. Molecules. 2017; 22:1-13.
15. Teoh LS, Lasekan O, Adzahan NM, et.al. The effect of ultraviolet treatment on enzymatic activity and total phenolic content of minimally processed potato slices. J Food Sci Tech. 2016;53(7):3035-3042.
16. Ullah MA, Tungmunnithum D, Garros L, et.al. Effect of ultraviolet-C radiation and melatonin stress on biosynthesis of antioxidant and antidiabetic metabolites produced in in vitro callus cultures of Lepidium sativum. L Int J Mol Sci. 2019;20(7):1787.
17. Zagoskina NV, Alyavina AK, Gladysenko TO, et.al. Ultraviolet rays promote development of photosystem II photochemical activity and accumulation of phenolic compounds in the tea callus culture (Camellia sinensis). Russ J Plant Physiol. 2005;52(6):731-739.
18. Zagoskina NV, Dubravina GA, Alyavina AK, et.al. Effect of ultraviolet (UV-B) radiation on the formation and localization of phenolic compounds in tea plant callus cultures. Russ J Plant Physiol. 2003;50(2):270-275.