

Umbilical cord preservation- A Review

Shukla P*

Department of Bioengineering, BIT Mesra, Ranchi, India, Jharkhand, India

***Corresponding author:** Shukla P, Department of Bioengineering, BIT Mesra, Ranchi, Jharkhand, India, Tel: +919471317757; E-mail: shuklaprity28@gmail.com

Received: March 13, 2017; Accepted: March 14, 2017; Published: March 21, 2017

Abstract

Umbilical cord (UC) is the main connection between mother and the growing fetus. It acts as the career of nutrients helps in exchange of gases etc. The length of the UC is approximately 20 inches. UC is of main interest for researchers in the recent times. The umbilical cord blood (UCB) has been fruitful source of Stem Cells. UCB helps in the reformation of different blood components. The umbilical cord provides the growing embryo with oxygen, nutrient-rich blood from the placenta. In the same manner, the fetal heart pumps deoxygenated, nutrient-depleted blood through the umbilical arteries back to the placenta.

Keywords: Umbilical cord; Umbilical cord blood; Stem cells

Introduction

"The umbilical cord blood (UCB) has been used progressively as a reservoir of stem cells for hematogenic reconstitution as an alternate to bone marrow or peripheral blood progenitor cells. Related and unrelated transplants of UCB are used for kids suffering with any malignant or non-malignant disease. Transplantation of umbilical cord blood has been used with successful results for the treatment of diseases like leukaemia, lymphoma, myelodysplasia, aplastic anemia (AA), hemoglobinopathies, metabolic storage diseases and immunodeficiencies.

There is still research going on in many parts of the world to know more uses of the UCB. The main emphasis is on the treatment of fatal diseases without the use of harmful medicines. Such technique provides an opportunity to repair the system naturally. The research works focus mainly on finding the use of UCB to cure damages caused due to heart disease, brain trauma, diabetes mellitus, and spine injury.

UCB is that the blood that remains inside the umbilical canal after the baby is born and cord being cut. In past, the umbilical cord, its blood and placenta was considered as a clinical/hospital waste, and were ultimately disposed of. Since the discovery that cord blood may be a reservoir of hemopoietic stem cells and since the primary transplant in 1988, this valuable source of stem cells is no longer thought of as a clinical waste. Public and family banks were created to process and cryopreserve this blood for future use.

Cord blood contains all the traditional components of blood - red blood cells, white blood cells, platelets and plasma. It is also made in hemopoietic (blood-forming) stem cells, similar to those found in bone marrow. The Haematopoietic (progenitor) stem cells area unit set in the liver and spleen of each embryo. Just before labour, migration of these cells via

blood stream begins from the liver to the bones for bone marrow formation. The umbilical cord can be removed either after the birth of the baby and expulsion of the placenta (placenta is outside the uterus attached with the baby) or after the birth of the baby and before the expulsion of the placenta (baby is born but the placenta remains inside the uterus).

Cord blood will be used for transplantation as another to bone marrow cells. Compared with adult peripheral blood or bone marrow, cord blood contains a larger proportion of extremely proliferative hemopoietic root cells. Most cord blood transplants have been done to treat diseases of the blood and system. It has also been accustomed restore the useful deficiencies of many genetic metabolic diseases. To date, more than seventy totally different diseases are treated with wire blood transplants.

The number of transplantations mistreatment duct cells is rising each year. Since 2005 allogeneic umbilical wire transplants in kids number the transplantations of bone marrow cells.

The positive outcomes from cord blood analysis and applications have led to new experimental applications for the treatment of different serious conditions. Scientists are investigation the chance that stem cells in wire blood could also be ready to replace cells of different tissues like nerve or heart cells.

Researchers and scientists accept that future therapies can be supported stem cells from numerous sources together with wire blood. Research is in progress in various centres for therapies of presently incurable diseases such as sclerosis and Alzheimers'.

Components of Umbilical Cord Blood:

The cellular component of UCB is primarily comprised of lymphocytes and monocytes. It has a comparable B-lymphocyte population and a lower absolute number of T-lymphocytes (CD3+) however a better CD4+/CD8+ quantitative relation compared to APB. UCB also has higher numbers of NK cells whereas lower numbers of CD56+ cytotoxic T-lymphocytes. UCB's relative immaturity compared to adult cell sources is more classified as showing a higher proportion of immature T-lymphocytes (CB45RA+) and reduced numbers of mature memory T-lymphocytes (CD45RO+). UCB cells also turn out fewer absolute levels of cytokines than adult cell sources. Furthermore, of the mRNA that is expressed in UCB, the anti-inflammatory cytokines interferon- γ (INF- γ), interleukin (IL)-4 and IL-10 aradditional well-endowed than for the unhealthy protein IL-2. This lack of mature immune function is attributed to UCB's low incidence of GvHD and microorganism transmission. Such cellular constitution could enable for less tight donor–recipient matching necessities, hence leading to shorter waiting amount for treatment. Rocha found that GvHD incidence was significantly lower in kids receiving UCB transplants compared to BM recipients once the supply was from associate degree HLA-identical relation. Rocha also incontestable a lower GvHD incidence in unrelated HLA-mismatched UCB recipients compared to HLA-identical BM recipients.

The enthusiasm over UCB began when it absolutely was found to contain an outsized population of haematogenic stem/progenitor cells compared to adult sources. These easily procured, low immunogenic sources of multipotential cells are thought to have the potential to become any form of cell within the body below specific conditions. Not only will the MNF contain roughly one hundred and twenty fifth CD34+ cells, a marker designated for its role in early haemopoiesis, but these cells appeared to be additional immature than those found in BM. In general, the level of maturity of a cell is identified by the cell's presence of or lack of a mixture of cell surface antigens. For instance, the CD34+ population in UCB can be outlined as

additional primitive than those found in BM as a result of a better proportion $(4\times)$ of them ar negative for CD38, a marker for pre-lymphoid cells. Another subset of CD34+ cells found in comparatively high numbers in UCB ar the additional primitive CD133+ cells. CD133+ cells have been identified in foetal brain and during this space ar thought-about to be neural stem cells (NSC) (67,70). However, it is not yet notable whether or not the CD133+ cells found in UCB ar phenotypically and functionally a twin of the NSC found in foetal brain.

Non-hematopoietic stem cell (a type of mesenchymal stem cell) has also been found in the UCB, but lesser in number when compared to Bone Marrow Cells (BM/BMC).

The mesenchymal stem cell (MSC) produces much large amount of osteoblast, adipocytes, astrocytes, neurons, chondroblasts and hematopoietic cells. Yang et al. characterized the master's degree as having positive markers for CD13, CD29, CD44, and CD90 and negative markers for CD14, CD31, CD34, CD45, CD51/61, CD64, CD106, and HLA-DR, while Robinson et al. defined the master's degree as positive for CD73, CD90, CD105, and CD166 and negative for CD31, CD34, CD45, CD80, and HLA-DR. Universal agreement on a phenotype of these cells has to initial be reached before their true abundance in UCB will be notable. Regardless, it is of a general consensus that there are way fewer master's degrees in UCB compared to BM.

Types of stem cells:

1. Embryonic stem cells:

These include cells that are found within the embryo, the foetus or the umbilical cord blood depending upon when they are harvested, embryonic stem cells can give rise to just about any cell in the human body.

2. Adult stem cells:

These are found in infants, children and adults; found in already developed tissues such as heart, brain and kidney. They usually give birth to cells within their resident organs.

3. Induced pluripotent stem cells (IPSC):

These stem cells are mature/adult, differentiated cells that have been experimentally "reprogrammed" into a stem cell-like

Process of umbilical cord preservation

- > Normal process of umbilical cord preservation is as follows:
- > The umbilical cord is acquired before or after the expulsion of the placenta.
- The cord is cut and its blood is collected
- Segments of the cord is cleaned, cut and put in container with antibiotics
- > The cord is then transferred to laboratory for further use.

Why are Umbilical cord stem cells beneficial? (http://www.cryoviva.in/banking-benefits/why-store-umbilical-cord/)

a. Rich Source of Stem Cells:

When compared to bone marrow, cord blood contains 10 times more number of stem cells, when both taken in equal proportion.

b. Regenerative Source:

The cord blood cells being younger have higher proliferating property as compared to stem cells found in bone marrow, and hence have a regenerative capacity.

c. Availability:

There are approximately 25000 individuals who are diagnosed with diseases that can be cured with stem cell therapy. Sometimes these sufferers do no find a perfect donor or have to pay a huge amount for the other treatment. In case of severe diseases as anemia, where the patient has to suffer blood deficiency and infections, for such sufferers cord blood banking is a

boon. Cord blood banking can be used for self and for other patients as well, since it is promptly available for transplant. This helps in saving many lives.

d. Pain:

Cord blood transplantation is a painless and a rapid process. It is so because the stem cells are separated from the umbilical cord beforehand. The whole procedure requires general anesthesia. When compared to extraction from bone marrow, it is a painful procedure because it is to be removed from hind end of the pelvic bone through a series of injections.

List of companies for preserving Umbilical Cord:

- 1. Cord Blood Registry California, USA,
- 2. ViaCord Massachusetts, USA
- 3. Cryo-Cell Florida, USA
- 4. China Cord Blood Corporation Beijing, China
- 5. Cryo-Save Netherlands
- 6. New York Cord Blood Program New York City,
- 7. CordVida São Paulo, Brazil,
- 8. Americord New York City,
- 9. CryoHoldco Latin America
- 10. Vita34 Leipzig, Germany,
- 11.LifeCell across india
- 12. Cordlife Sciences Kolkata India Pvt. Limited India
- 13. BabyCell Lonavala in Maharashtra india

REFERENCES

- 1. Jovandaric MZ. The Effect of Abruptio Placentae on Perinatal Outcome of Pregnancy. J Clin Case Rep. 2016;6:775.
- Cho TH, Park KM. Use of Acupuncture Point Injection with Placental Extract for Treatment of Complex Regional Pain Syndrome. J Pain Relief. 2016;5:246.
- 3. Mahendru R, Bansal S. Placenta Accreta: Then and Now. J Preg Child Health. 2016;3:e124.
- Framarino-dei-Malatesta M, D'Amelio R, Piccioni MG, Martoccia A, Casorelli A, et al. Conservative Management of Placenta Accreta by Systemic Metho-trexate: Report of Two Cases and Review of the Literature. J Clin Case Rep. 2016;6:706.
- 5. Elshennawy TMA. Effect of Gestational Diabetes on Gross Morphology, Histology and Histochemistry of Human Placenta. Endocrinol Metab Syndr. 2016;5:227.
- Haridas N, Venkatesan V, Bhonde R. Establishment of Impaired Angiogenesis Using Human Placental Mesenchymal Stem Cells under Micronutrient Deficiency. Transl Med. 2015;6:162.
- 7. Oliveira PN, Caldas R, Rodrigues C, Kok MF, Rasteiro C, et al. Congenital Acute Lymphoblastic Leukemia with Placental Involvement: Case Report x. J Preg Child Health. 2015;2:204.
- Ojeda NB. Necrotizing Enterocolitis in Rat Offspring Exposed to Placental Insufficiency: Role of Aldosterone, Oxidative Stress and Leptin. J Preg Child Health. 2015;2:196.

- Sakaguchi Y, Hamaguchi S, Otani T, Furukawa N, Yamaguchi S. Anesthetic Management of Cesarean Section in a Grown-Up Congenital Heart Patient with Placenta Previa and Giant Placental Tumor: A Case Report. J Anesth Clin Res. 2015;6:586.
- Fali L, Yadav Y, Dushyant M, et al. Materno-fetal Outcomes in Patients with Abnormally Invasive Placenta: A 14 Year Experience. J Women's Health Care. 2015;4:286.
- 11. Zeng G, Li Q, Ju H, et al. Therapeutical Comparison of Mesenchymal Stem Cells from Different Tissues of Human Placenta in the Treatment of Mice with Acute Hepatic Failure. J Stem Cell Res Ther. 2015;5:316.
- 12. Nogueira R, Pinto-Ribeiro F, Pereira SM, et al. Macroscopic and Histopathological Study of the Placenta An Essential Resource in Litigation Processes. J Clin Res Bioeth. 2015;6:247.
- Nakamura T, Iwase A, Ishida C, et al. A Placental Site Trophoblastic Tumor Complicated with Arteriovenous Malformation: A Case Report. J Clin Case Rep. 2015;5:596.
- Fiossi-Kpadonou É, Kpadonou GT, Azon-Kouanou A, et al. Placenta Processing: Sociocultural Considerations and Impact on the Future of Child in Benin. J Child Adolesc Behav. 2015;3:222.
- 15. Jepsen E, Behling E, Schwarting R, et al. Extranodal Natural Killer/T-Cell Lymphoma of the Placenta: A Case Report of a Previously Undescribed Entity and Review of the Literature. J Clin Case Rep. 2015;5:540.
- Chen T, Wang X, Wu Q, et al. Comparison of Bacterial Composition in Blood and Placentas Using Conventional and Molecular Methods. Clin Microbiol. 2015;4:200.
- 17. Treesh SA, Khair NS. Histological Changes of the Human Placenta in Pregnancies Complicated with Diabetes. J Cytol Histo.l2015; 6:307.
- Mamour G, Diarra NGM, lamine CM, et al. Risk of Abruption Placenta in Women with Preeclampsia Undergoing Labour Induction with Misoprostol. J Women's Health Care. 2015;4:224.
- Omer HA, Kutb MA, Kaatabi HA. Histopathological Changes in Placenta of Rat Induced by Levtricetam. Int J Neurorehabilitation. 2014;1:134.
- Bosco C, Eugenia D. Effects of Maternal Alcoholism on Placental Function and Lung Fetal Development. J Cell Sci Ther. 2014;5:178.
- Ringdén O, Solders M, Erkers T, et al. Successful Reversal of Acute Lung Injury using Placenta-Derived Decidual Stromal Cells. J Stem Cell Res Ther. 2014;4:244.
- 22. Singh N, Kumari P. Placenta Accreta: A Mini Review. J Preg Child Health. 2014;1:e104.
- Elie N, Telesphore ME, John A. A Case Report of a Successful Conservative Management of Placenta Increta. J Preg Child Health. 2014;1:113.
- Hasan S, Imtiaz F, Ali A, et al. Feto-Maternal Outcome of Placenta Praevia after Previous Cesarean Section in a Tertiary Care Hospital. Epidemiology (Sunnyvale). 2014;4:162.
- 25. Roman C, Dafashy T, Hegde S, et al. Epigenetic Modifications of Preeclamptic Placenta-A Systematic Review. Gynecol Obstet(Sunnyvale). 2014;4:233.
- 26. Berthelot-Ricou A, Villot A, Bernard J, et al. New Multidisciplinary Approach of Conservative Surgical Management of Placenta Percreta Antenatally Diagnosed. Surgery Curr Res. 2014;4:195.
- 27. Al-Bayati MA, Ahmad MA, Khamas W. The Potential Effect of L-arginine on Mice Placenta. Adv Pharmacoepidemiol Drug Saf. 2014;3:150.

- 28. Nabulsi BK, Kadi M, AlAbadi H, et al. Pregnant Woman with Fulminant Disseminated TB to the Omentum and Placenta. Gynecol Obstet (Sunnyvale). 2014;4:225.
- Chen J, Qian Li, Rialdi A, et al. Influences of Maternal Stress during Pregnancy on the Epi/genome: Comparison of Placenta and Umbilical Cord Blood. J Depress Anxiety. 2014;3:152.
- 30. Moodley S. Transplacental Infection of HIV-1 and the Associated Risk Factors In Utero. J Clin Cell Immunol. 2014;5:195.
- Akinwuntan Akinwunmi L, Oshinowo Omololu S. Conservative Management of Morbidly-Adherent Placenta Following Vaginal Deliveries: A Case Series. Gynecol Obstet (Sunnyvale). 2014;4:207.
- 32. Matsuda Y, Ogawa M, Konno J. Prognosis of the Babies Born from Placental Abruption Difference between Intrauterine Fetal Death and Live-Born Infants. Gynecol Obstet (Sunnyvale). 2013;4:191.
- Robert Johnston C, Morgan Swank L, Vineet Shrivastava K. Management of Placenta Accreta Complicated by Pulmonary Embolus and Heparin Induced Thrombocytopenia. Gynecol Obstet. 2013;3:188.
- Yoshikawa C, Takano F, Ishigaki Y, et al. Effect of Porcine Placental Extract on Collagen Production in Human Skin Fibroblasts *In Vitro*. Gynecol Obstet. 2013;3:186.
- 35. de la Monte S. Prenatal Alcohol Exposure Mediates Adverse Effects on Placental and Fetal development by Impairing Trophoblast Differentiation and Vascular Transformation. J Clin Exp Pathol. 2013;3:139.
- Undogan F, Gilligan J, Ooi JH, et al. Dual Mechanisms of Ethanol-Impaired Placentation: Experimental Model. J Clin Exp Pathol. 2013;3:142.
- 37. Novotny S, Wallace K, Herse F, et al. CD4+ T Cells Play a Critical Role in Mediating Hypertension in Response to Placental Ischemia. J Hypertens. 2013;2:116.
- Androutsopoulos G, Decavalas G. Perioperative Internal Iliac Artery Balloon Occlusion in Patients with Abnormal Placental Invasion. J Community Med Health Educ. 2013;3:e117.
- Gunatillake T, Chui A, Said JM. The Role of Placental Glycosaminoglycans in the Prevention of Pre-Eclampsia. J Glycobiol. 2013;2:105.
- Corocleanu M. Embryonic-Like Stem Cells Derived from Postpartum Placenta Delivered After Spotaneous Labor Emerging as Universal Prophylactic Cancer Vaccine. J Vaccines Vaccin. 2012;3:163.
- Genovese F, Marilli I, Carbonaro A, et al. Management and Time of Delivery in Asymptomatic Complete Placenta Previa: A Case Report and Review of Literature. Gynecol Obstet. 2012;2:130.
- Alanis MC, Steadman EM, Manevich Y, et al. Maternal Obesity and Placental Oxidative Stress in the First Trimester. J Obes Wt Loss Ther. 2012;2:143.
- 43. Wallace SC. Root Coverage Grafting Comparing Placental Derived Membrane to Acellular Dermis Matrix: A Case Series. Dentistry. 2012;2:137.
- 44. Abreu LA, Madruga B, Gouvea J, et al. Anesthesia for a Cesarean Section in a Patient with a Congenital Heart Disease and Complete Placenta Previa. J Anesth Clin Res. 2012;3:212.
- 45. Adigun TA, Eyelade O. Choice of Anaesthetic Technique for Delivery of Pregnancy Complicated by Placenta Previa in Ibadan. J Anesth Clin Res. 2012;3:205.
- Reiter JL, Lee MJ. Can Loss of Imprinting in the Placenta Serve as a Biosensor of the Perinatal Environment? J Mol Biomark Diagn. 2012;3:e109.

- 47. Pafumi C, Leanza V, Carbonaro A, et al. Focal Placenta Accreta and Spontaneous Uterus Rupture in the Post-Partum. J Women's Health Care. 2012;1:105.
- Butrym JA, Rybka J, Jurczyszyn A, et al. Adult Acute Biphenotypic Leukemias: Polish Single Centre Experience. J Mol Biomark Diagn. 2015;6:209.
- 49. Hanna JRA. Expression of CD95 in Acute Lymphocytic Leukemia (ALL) in Egyptian Children before and after Treatment. J Blood Disord Transfus. 2015;6:250.
- 50. Galanzha EI. Blood and Lymph Circulating Cells: Well-Known Systems, Well-Forgotten Interdependence. J Blood Lymph. 2011;1:e104.
- 51. Song Y, Bixby Y, Roulston D, et al. The Challenge of t (6;9) and FLT3-Positive Acute Myelogenous Leukemia in a Young Adult. J Leuk (Los Angel). 2014;2:167.
- 52. Schiller GJ, Muchmore E. How to Teach the Topic of Acute Myelogenous Leukemia: Recommendations for Achieving Curricular Milestones. J Leuk (Los Angel). 2014;2:148.
- 53. Cerny J, Ramanathan M, Rosmarin AG, et al. What should be the Therapy for CD25 Positive Acute Myelogenous Leukemia?. J Hematol Thrombo Dis. 2014;2:e111.
- 54. Taylor AG, Snyder AE, Anderson JG, et al. Gentle Massage Improves Disease- and Treatment-Related Symptoms in Patients with Acute Myelogenous Leukemia. J Clin Trials. 2014;4:161.
- 55. Achkar WAL, Moassass F, Ikhtiar A, et al. Cytogenetic Evolution in a Patient with Chronic developing a Secondary Acute Myelogenous Leukemia Subtype M5 Resistant to Imatinib Mesylate Therapy. J Leuk (Los Angel). 2013;1:118.
- 56. Kiran K, Narender Kumar, Man Updesh Singh S, et al. An Uncommon Morphology of Acute Undifferentiated Leukemia: Report of a Rare Case. J Bone Marrow Res. 2013;1:103.
- 57. Tulara NK. Adult T Cell Leukemia/Lymphoma in a 56 years Old Indian Male with History of Miliary Koch's on Anti-Tubercular Therapy. Oncol Cancer Case Rep. 2016;2:110.
- Orensen SV, Peng S, Dorman E, et al. The Cost-Effectiveness of Ibrutinib in Treatment of Relapsed or Refractory Chronic Lymphocytic Leukemia. Health Econ Outcome Res Open Access. 2016;2:121.
- 59. CBharath V, Hsia CC. Chronic Lymphocytic Leukemia with Leptomeningeal Involvement. Chemo Open Access. 2016;5:I102.
- 60. Frezzato F, Trimarco V, Visentin A, et al. Targeting Bruton's Tyrosine Kinase in Chronic Lymphocytic Leukemia at the Crossroad between Intrinsic and Extrinsic Pro-survival Signals. J Leuk. 2016;4:207.
- 61. Robak T. BCL-2 inhibitors for Chronic Lymphocytic Leukemia. J Leuk. 2015;3:e114.
- 62. Pathak P. The Changing Therapeutic Landscape of Chronic Lymphocytic Leukemia. J Blood Lymph. 2015;5:e120.
- 63. Knauf W, Re D. Chronic Lymphocytic Leukemia: Raising Expectations in the Treatment of Elderly Patients. J Leuk. 2015;3:181.
- 64. Yanagiya S, Sato K, Tsukada N, et al. B-cell Chronic Lymphocytic Leukemia/Small Lymphocytic Lymphoma, without the Development of Richter's Syndrome, with Neoplastic Cells Lacking CD20 Antigen Expression after Rituximab Treatment. J Leuk. 2015;3:179.
- 65. Zhou J, Tewari A, Nassiri M, et al. Changes in Antigen Expression in a Follow-up of Chronic Lymphocytic Leukemia/Small Lymphocytic Lymphoma. J Leuk. 2015;3:177.

- 66. Robak T. Staging and Prognostic Factors in Chronic Lymphocytic Leukemia: Current Status. J Leuk (Los Angel) 2014;2:e111.
- 67. Robak T. Approval for Novel Drugs in Chronic Lymphocytic Leukemia. J Develop Drugs. 2014;3:e138.
- Jordaan G, Liao W, Coriaty N, et al. Identification of Histone Epigenetic Modifications with Chromatin Immunoprecipitation PCR Array in Chronic Lymphocytic Leukemia Specimens. J Cancer Sci Ther. 2014;6:325-332.
- 69. Jeyakumar G, Ferrajoli A. New Therapeutic Advances in Chronic Lymphocytic Leukemia. J Leuk (Los Angel). 2014;2:144.
- 70. Jonsson V, Awan H, Johannesen TB, et al. Chronic Lymphocytic Leukemia, Advantages of Monoclones?. J Leuk (Los Angel). 2014;2:142.
- Ghannam DME, Taalab MM, Ghazy HF, et al. CD26 Expression in Mature B-cell Neoplasms and its Prognostic Impact on B-Cell Chronic Lymphocytic Leukemia. J Blood Disord Transfus. 2014;5:222.
- 72. Gaman AM,Gaman MA. Immune Thrombocytopenia in Chronic Lymphocytic Leukemia. J Blood Disord Transfus. 2014;5:198.
- 73. Matschke J, Eisele L, Sellmann L, et al. Abnormal Free Light Chain Ratios are Significantly Associated with Clinical Progression in Chronic Lymphocytic Leukemia. J Leuk (Los Angel). 2014;2:127.
- 74. Kreuzer KA. Chronic Lymphocytic Leukemia below the Radar J Leuk (Los Angel). 2014;1:e104.
- 75. Jaglowski SM, Geyer S, Heerema NA, et al. Barriers to Proceeding to Reduced-Intensity Allogeneic Stem Cell Transplant in Chronic Lymphocytic Leukemia. J Leuk (Los Angel). 2013;1:121.
- 76. Shukla A, Chaturvedi NK, Ahrens AK, et al. Stromal Tumor Microenvironment in Chronic Lymphocytic Leukemia: Regulation of Leukemic Progression. J Leuk (Los Angel). 2013;1:113.
- 77. Robak T. Inhibitors of B-Cell Receptor Signaling for the Treatment of Chronic Lymphocytic Leukemia. J Leuk (Los Angel). 2013;1:e101.
- 78. Sora F, Chiusolo P, Laurenti L, et al. Ponatinib before and after Allogeneic Stem Cell Transplantation for Ph+ Acute Lymphoblastic Leukemia or Lymphoid Blast Crisis of Chronic Myelogenous Leukemia: A Single Center Experience. J Bone Res. 2016;4:169.
- 79. Pennisi A, Jewell S, Gralewski J, et al. Acute Lymphoblastic Leukemia evolving from atypical Chronic Myelogenous Leukemia: Case Report and Review of the Literature. J Leuk. 2015;S1: S1-007.
- 80. Marfe G, Stefano CD. Cancer Stem Cells in Chronic Myelogenous Leukemia. J Leuk (Los Angel). 2014;2:159.
- Ahmed MM, Said ZS, Montaser SAW. Chronic Myelogenous Leukemia: Cytogenetic and Biochemical Consequences and Applications for Diagnosis and Judgment. J Cytol Histol. 2014;S4:015.
- 82. El Naggar AA, Shama A, Zaki NE, et al. Bilateral Visual Loss in a Patient with Chronic Myelogenous Leukemia after Initiation of Imatinib Therapy. J Leuk (Los Angel). 2013;1:119.
- 83. Imtiyaz Ahmad AB, Mir R, Zuberi M, et al. () Inactivation of RIZ1 Gene by Promoter Hypermethylation is Associated with Disease Progression and Resistance to Imatinib in Indian Chronic Myelogenous Leukemia Patients, First Study from India. J Cancer Sci Ther. 2013;5:045-051.
- Chen WY. Cancer Acquired Resistance: A New Lesson from Chronic Myelogenous Leukemia. J Bone Marrow Res. 2013;1:e101.

- Yin CC. Detection and Molecular Monitoring of Minimal Residual Disease in Chronic Myelogenous Leukemia. J Clin Exp Pathol. 2012;2:e110.
- 86. Kuroda J, Taniwaki M. Principles and Current Topics Concerning Management of Tyrosine Kinase Inhibitor Therapy for Chronic Myelogenous Leukemia. Translational Medic. 2011;S2:001.
- Abduljabbar A, Alanazi O, Fathadin A, et al. Poromatosis Following Ewing Sarcoma. J Clin Exp Dermatol Res. 2016;7:366.
- Madoz-Gúrpide J, Herrero-Martín D, Gómez-López G, et al. Proteomic Profiling of Ewing Sarcoma Reveals a Role for TRAF6 in Proliferation and Ribonucleoproteins/RNA Processing. J Proteomics Bioinform. 2016;9:166-175.
- 89. Elm'hadi C, Khmamouche MR, Toreis M, et al. An Atypical Etiology of Mediastinal Lymphadenopathy: Extraskeletal Ewing Sarcoma. J Integr Oncol. 2016;S1:007.
- 90. Tacyildiz N, Tanyildiz G, Soydal C, et al. Assessment of Sorafenib and AntiVEGF Combination Therapy Response which Added to Neoadjuvant Therapy in two Pediatric Metastatic Ewing Sarcoma Patients by Fluorine-18 Fluorodeoxyglucose Positron Emission Tomography (18F-PET) Method: It may Determine the Prognosis. J Nucl Med Radiat Ther. 2015;6:212.
- Fluck F, Falcão LM, Paixão R, et al. Extraskeletal Ewing Sarcoma in a Young Patient During Pregnancy. J Clin Case Rep. 2015;5:485.
- 92. Nam JK. Primary Ewing Sarcoma of the Kidney with Inferior Vena Cava Invasion. Med Surg Urol. 2015;4:146.
- 93. Kikuta K, Tsunehiro Y, Yoshida A, et al. Proteome Expression Database of Ewing sarcoma: a segment of the Genome Medicine Database of Japan Proteomics. J Proteomics Bioinform. 2009;2:500-504.
- Gunawardena D, Kuruppumullage SDL. A Rare Case of Light Chain Myeloma with Amyloidosis. J Blood Lymph. 2016;6:151.
- 95. Nishimura K, Tanaka S, Takahashi Y, et al. Huge Localized Amyloidosis of the Sinonasal Cavity: A Rare Case Report. J Clin Case Rep. 2016;6: 797.
- 96. Dogan EE, Şahin F, Saydam G. Amyloidosis: Laboratory and Clinical Perspectives. Med chem (Los Angeles). 2016;6:270-279.
- Terzi S, Calabrò T, Brodano GB, Gasbarrini A, Boriani S. Systemic Amyloidosis with Predominant Spine Involvement: A Case Report. Orthop Muscular Syst. 2015;4:202.
- Arous S, Bensahi I, Noureddine M, Habbal R. A Typical Case of a Multiple Myeloma Revealed by Cardiac Amyloidosis. Angio. 2015;1 3:163.
- 99. YY, Su L, Lin M, Li J, Ding Ml, et al. Systemic Amyloidosis in a Patient with Type 2 Diabetes Mellitus as a Uncommon Cause of Non-Diabetic Renal Disease. J Diabetes Metab. 2015;6:528.
- 100.García de Veas Silva JL, Guitarte CB, Valladares PM, Millán RD, Rojas Noboa JC. A Challenging Case of IgD Kappa Multiple Myeloma Associated With Primary Amyloidosis: Importance of Serum Free Light Chains in Monitoring Treatment Response and Disease Relapse. J Leuk (Los Angel). 2014;2:164.
- 101.Jaggia A, Wechalekar A, Khullar D, Kumaran V. Primary Amyloidosis (A Rare Disorder): Clearing Myths and Fallacies. J Nephrol Ther. 2014;4:187.
- 102.Rocha A, Bravo F, Beirão I, Vizcaíno J, Oliveira JC, et al. Urinary Biomarkers for Kidney Disease in ATTR Amyloidosis. J Nephrol Ther. 2014;4:181.

- 103.Liu M, Song Z, Liu J, Liu R, Liu H, et al. 18F-FDG-PET-CT with Little Value in Different Diagnosis between Pulmonary Malignancy and Amyloidosis. J Hematol Thrombo Dis. 2014;2:156.
- 104.AL-Saedi M, Batwa F, Absi A, Dahlan Y, Satti M, et al. Liver Amyloidosis Complicated with Liver Failure: A Case Report and Review of the Literature. J Gastroint Dig Syst. 2012;S3:002.