



Trade Science Inc.

Organic CHEMISTRY

*An Indian Journal**Full Paper*

OCAIJ, 6(2), 2010 [160-165]

Synthesis of 6-chloro-8-substituted-9[H]-purine derivatives and bioactivity studies

M.Suresh, P.Lavanya, K.Vasu, C.Venkata Rao*

Department of Chemistry, Sri Venkateswara University, Tirupati - 517 502, (INDIA)

E-mail : cvrsvu@gmail.com

Received: 14th February, 2010 ; Accepted: 24th February, 2010

ABSTRACT

Selective preparation of 6-chloro-8-substituted-9[H]-purines has been achieved, by treating 6-halo-4,5-diaminopyrimidine and substituted aromatic acid with pyridine reagent to get 75–90% yields by refluxing at short reaction time 1.5 to 2.0 hours. Their chemical structures were characterized using IR, ¹H NMR and Mass spectral studies. All the above compounds were screened for anti-microbial activity, anti-oxidant activity and their bioassay showed them to possess significant antimicrobial activity and anti-oxidant activity. © 2010 Trade Science Inc. - INDIA

KEYWORDS

Purine derivatives;
6-chlorodiaminopyrimidine;
Aromatic acids;
Pyridine and acetonitrile;
Anti-microbial activity;
Anti-oxidant activity.

INTRODUCTION

Purines derivatives have been the subject of considerable interest from many biological^[1-5] point of interest and consequently the synthesis of these compounds have gained importance. The condensation of pyrimidines^[6,7] and substituted aromatic acids in the presence of pyridine under reflux condition yields 6-chloro-8-substituted-9[H]-purines derivatives (**5**) in good yield. The reaction is very fast and 90% conversion was observed in 2 hours time of reaction. We have been encouraged by these results and examined with several aromatic acids under the optimized conditions (TABLE 1). Here the acetonitrile is much better solvent than all other tested solvents such as MeOH, DCM, THF in small amount of pyridine to get high yields 75-90%.

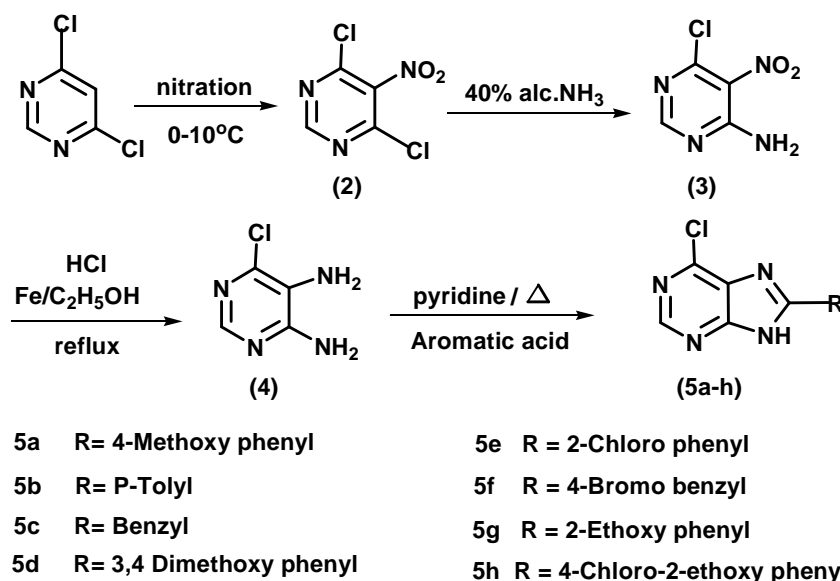
4,6-dichloro-5-nitropyrimidine (**2**)^[8-11] was prepared from the reaction of 4,6-dichloropyrimidine with nitration to obtained 90% yield due to the mild conditions in strong nitric acid was used. Compound (**2**) was

further converted to compound (**3**)^[12,13] by reaction with 40% ethanolic ammonia^[14] in good yield. Further compound (**3**) under reduction^[15-18] with Fe/HCl in ethanol to give 6-chloro-4,5-diaminopyrimidine (**4**)^[19-22].

Here we have developed an efficient synthesis of several new 6-chloro-8-substituted-9[H]-purine derivatives through the condensation between 6-chloro-4,5-diaminopyrimidines and substituted benzoic acids and phenyl acetic acids using pyridine as the catalyst. The attractive features of this process are low cost reagents, good reaction yields and short reaction time. Therefore it is an effective method for the synthesis of 8-substituted-9[H]-purine derivatives TABLE 1.

EXPERIMENTAL PROCEDURE

The ¹H-NMR spectra were recorded at 500 MHz with a Bruker Avance DPX 300 instrument. Mass spectra were recorded under ESI-Mass with a LC-Trap-SL instrument and presented as m/z (% rel int.). Elemental analyses (C, N, H) results were found to



Scheme 1

be in good agreement with the calculated values. Melting points were determined with Capillaries Thomas Hoover melting point apparatus and are uncorrected. TLC monitored all reactions and purity of the synthesized compounds.

General procedure

6-Chloro-8-substituted-9[H]-purines: A mixture of 6-chloro-4,5 diaminopyrimidines (1mmol), substituted benzoic acids and phenyl acetic acids (1.1mmol), acetonitrile (5ml) and catalytic amount of pyridine were stirred under reflux for about two hours. The resulting mixture was cooled and acidified and recrystallized with ethanol.

6-Chloro-8-(4-methoxy phenyl)-9[H]-purine (5a)

0.76g (85% yield), Solid, mp.: 242-244°C (ethanol). ¹H NMR spectrum (DMSO-d₆, 200 MHz): δ 3.8 (3H, s); 7.0 (2H, d); 8.25 (2H, d); 8.6 (1H, s). ESI-Mass spectrum, m/z (I, %): 261[M+H]⁺ (100). Found, %: C 55.26; H 3.44; N 21.48; Cl 13.61; O 6.13. C₁₂H₉ClN₄O. Calculated, %: C 55.28; H 3.46; N 21.50; Cl 13.63.

6-Chloro-8-p-tolyl-9[H]-purine (5b)

0.71g (83.5% yield), Solid, mp.: 257-258°C (ethanol). ¹H NMR spectrum (DMSO-d₆, 200 MHz): δ 2.45 (3H, s); 7.30 (2H, d); 8.2 (2H, d); 8.6 (1H, s). ESI-Mass spectrum, m/z (I, %): 244.9[M+H]⁺ (100).

Found, %: C 58.87; H 3.71; N 22.89; Cl 14.50. C₁₂H₉ClN₄. Calculated, %: C 58.89; H 3.70; N 22.90; Cl 14.52.

6-Chloro-8-benzyl-9[H]-purine (5c)

0.7g (81% yield), Solid, mp.: 160-163°C (ethanol). ¹H NMR spectrum (DMSO-d₆, 200 MHz): δ 4.22 (2H, s); 7.33 (5H, m); 8.6 (1H, s); 13.4 (1H, s). ESI-Mass spectrum, m/z (I, %): 244.9[M+H]⁺ (100). Found, %: C 58.87; H 3.66; N 22.88; Cl 14.50. C₁₂H₉ClN₄. Calculated, %: C 58.89; H 3.68; N 22.90; Cl 14.51.

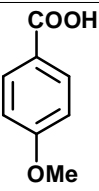
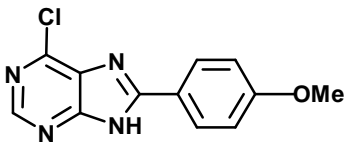
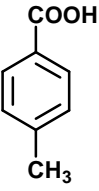
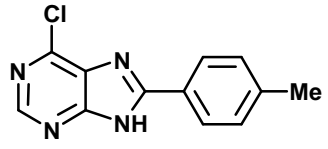
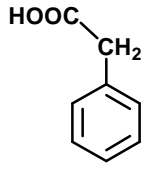
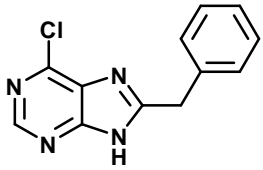
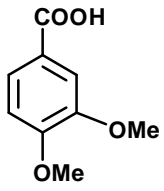
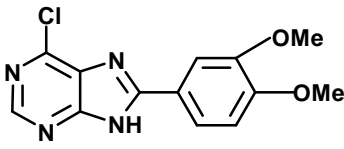
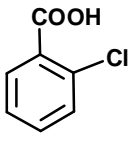
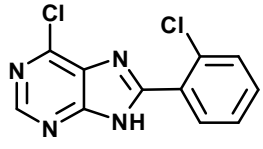
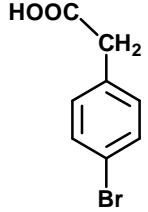
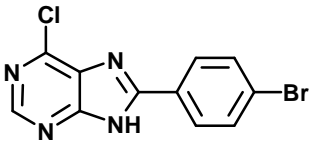
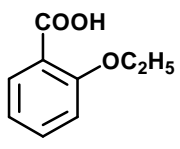
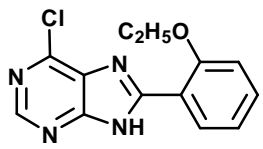
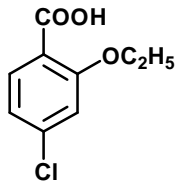
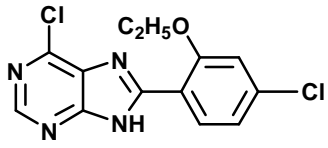
6-Chloro-8-(3,4-dimethoxyphenyl)-9[H]-purine (5d)

0.81g (90% yield), Solid, mp.: 250-252°C (ethanol). ¹H NMR spectrum (DMSO-d₆, 200 MHz): δ 3.90 (3H, s); 3.95 (3H, s); 7.08 (1H, d); 7.85 (2H, m); 8.6 (1H, s). ESI-Mass spectrum, m/z (I, %): 291[M+H]⁺ (100). Found, %: C 53.69; H 3.74; N 19.25; Cl 12.23. C₁₃H₁₁ClN₄O₂. Calculated, %: C 53.70; H 3.78; N 19.27; Cl 12.22.

6-Chloro-8-(2-chlorophenyl)-9[H]-purine (5e)

0.64g (80% yield), Solid, mp.: 204-205°C (ethanol). ¹H NMR (DMSO-d₆, 200 MHz): δ 7.4 (3H, m); 7.95 (1H, d); 8.7 (1H, s); 13.9 (1H, s/m). ESI-Mass spectrum, m/z (I, %): 291[M+H]⁺ (100). Found, %: C 49.80; H 2.24; N 21.11; Cl 26.80. C₁₁H₆Cl₂N₄. Calculated, %: C 49.81; H 2.26; N 21.13; Cl 26.79.

TABLE 1 : Synthesis of 6-chloro 8-substituted-9[H]-purines

Entry	R	Product	Time (h)	Yield (%)
5a			2.0	85%
5b			1.5	85%
5c			1.5	85%
5d			1.5	90%
5e			2.0	80%
5f			2.0	75%
5g			1.5	85%
5h			2.0	83%

6-Chloro-8-(4-bromobenzyl)-9[H]-purine (5f)

0.53g (75% yield), Solid, mp.: 181-182°C (ethanol). ¹H NMR spectrum (DMSO-d₆, 200 MHz): δ 4.22 (2H, s); 7.24 (2H, d); 7.42 (2H, d); 8.6 (1H, s); 13.6 (1H, s/m). ESI-Mass spectrum, m/z (I, %): 322.8[M+H]⁺ (100) & 324.9[(M+2)+H]⁺ (100).

Found, %: C 45.95; H 2.53; N 17.84; Cl 11.30; Br 22.30. C₁₂H₈BrClN₄. Calculated, %: C 45.93; H 2.55; N 17.86; Cl 11.32; Br 22.32.

6-Chloro-8-(2-ethoxyphenyl)-9[H]-purine (5g)

0.56g (78% yield), Solid, mp.: 170-172°C (etha-

TABLE 2 : Antibacterial activity* of the target compounds (5a-h)

Compound	Concentration (µg)	Zone of inhibition (mm)			
		Gram-positive bacteria		Gram-negative bacteria	
		<i>S.aureus</i>	<i>B.subtilis</i>	<i>P.vulgaris</i>	<i>K.pneumoniae</i>
5a	100	30	31	31	35
	200	35	38	40	39
5b	100	20	22	17	19
	200	26	25	22	21
5c	100	25	28	20	25
	200	30	32	23	28
5d	100	32	30	26	26
	200	35	35	28	30
5e	100	12	11	14	13
	200	15	13	17	16
5f	100	26	26	20	21
	200	32	30	24	24
5g	100	27	26	31	34
	200	32	30	36	40
5h	100	11	12	11	11
	200	14	16	14	13
Chloramphenicol	100	35	38	40	42
	200	39	41	44	45

*c = 100µg / ml. * c = 200µg / ml

nol). ¹H NMR spectrum (DMSO-d₆, 200 MHz): δ 4.22 (2H, q); 1.48 (3H, t); 7.18 (2H, m); 7.45 (1H, m); 8.25 (1H, d); 8.48 (1H, s); 12.8 broad (1H, s). ESI-Mass spectrum, m/z (I, %): 275[M+H]⁺ (100). Found, %: C 56.819; H 4.02; N 20.38; Cl 12.91. C₁₃H₁₁ClN₄O. Calculated, %: C 56.83; H 4.00; N 20.40; Cl 12.93.

6-Chloro-8-(4-chloro-2-ethoxyphenyl)-9[H]-purine (5h)

0.60g (83% yield), Solid, mp.: 157-158°C (ethanol). ¹H NMR spectrum (DMSO-d₆, 200 MHz): δ 4.22 (2H, q); 1.48 (3H, t); 7.18 (2H, d); 7.45 (1H, s); 8.35 (1H, s); 12.8 broad (1H, s). ESI-Mass spectrum, m/z (I, %): 309[M+H]⁺ (100). Found, %: C 50.48; H 3.24; N 18.17; Cl 22.90. C₁₃H₁₁ClN₄O. Calculated, %: C 50.51; H 3.26; N 18.12; Cl 22.94.

Antimicrobial testing

The compound (5a-h) was tested for in vitro antimicrobial activity at two different concentrations 100 and 200µg per disc. The antibacterial activity was screened against *Staphylococcus aureus*, *Bacillus*

subtilis (Gram-positive bacteria) and *Proteus vulgaris*, *Klebsiella pneumoniae* (Gram-negative bacteria) on nutrient agar plates at 37°C for 24 hrs using chloramphenicol as reference during. The compounds were also evaluated for their antifungal activity against *Aspergillus niger* and *Penicillium chrysogenum* using fluconazole as standard drug. Fungi cultures were grown on potato dextrose agar (PDA) medium at 25°C. The spore suspension was adjusted to 10⁶ pores ml⁻¹ at an mg ml⁻¹ concentration by the Vincent and Vincent method.

Antioxidant testing

The compounds (5a-h) is tested for antioxidant property by nitric oxide and DPPH methods.

Assay for Nitric Oxide (NO) Scavenging Activity Sodium nitroprusside (5µM) in phosphate buffer pH 7.4 was incubated with 100µM concentration of test compounds dissolved in a suitable solvent (methanol) and tubes were incubated at 25°C for 120 min. Control experiment was conducted with equal amount of solvent in an identical manner. At intervals 0.5ml of incubation solution was taken and diluted with 0.5ml

Full Paper

TABLE 3 : Antifungal activity* of the target compounds (5a-h)

Compound	Concentration	Zone of Inhibition (mm)	
	(µg/ml)	<i>A.niger</i>	<i>P.chrysogonium</i>
5a	100	24	20
	200	27	26
5b	100	14	14
	200	19	17
5c	100	26	26
	200	30	29
5d	100	31	18
	200	38	19
5e	100	15	25
	200	18	28
5f	100	17	30
	200	22	32
5g	100	30	26
	200	33	28
5h	100	28	16
	200	32	18
Fluconazole	100	38	41
	200	42	44

*c = 100µg /ml, *c = 200µg / ml

of griess reagent (1% sulfanilamide, 0.1% N-naphthyle-thylenediamine dihydrochloride and 2% o-phosphoric acid dissolved in distilled water). The absorbance of the chromophore formed during diazotization of nitrite with sulfanilamide and subsequent N-naphthyle-thylenediamine dihydrochloride was read at 546nm.

Reduction of 1,1-diphenyl-2-picrylhydrazyl (DPPH) Free Radical (DPPH Method): The nitrogen centered stable free radical DPPH has often been used to characterize antioxidants. It is reversibly reduced and the odd electron in the DPPH free radical gives a strong absorption maximum at 517nm, which is purple in color. This property makes it suitable for spectrophotometric studies. A radical scavenging antioxidant reacts with DPPH stable free radical and converts into 1,1-diphenyl-2-picrylhydrazine. The resulting decolorization is stoichiometric with respect to the number of electrons captured. The change in the absorbance produced in this reaction has been used to measure antioxidant properties. The solutions of test compounds (100µM) were added to DPPH (100µM) in ethanol. The tubes were kept at an am-

TABLE 4 : Antioxidant activity* of the target compounds (5a-h)

Compound	% Inhibition at 100 µM	
	Nitric oxide method	DPPH method
5a	82.25	84.74
5b	34.33	38.12
5c	91.18	93.65
5d	29.21	27.75
5e	70.23	72.25
5f	24.35	22.25
5g	75.10	74.25
5h	78.24	68.25

*c = 100µM

bient temperature for 25 minutes and the absorbance was measured at 517nm. The difference between the test and the control experiments was taken and expressed as the percentage scavenging of the DPPH radical.

RESULTS AND DISCUSSION

The results of the compounds of preliminary antimicrobial testing are shown in TABLE 1 and 2. The results revealed that the inhibitory activity against Gram-positive bacteria was higher than Gram-negative bacteria. The imidazole derivatives (5b), (5e), and (5h) were displayed least activity. The compounds (5a), (5c), (5d), (5f), and (5g) showed excellent activity against Gram-positive bacteria (inhibitory zone > 25mm) and good activity against Gram negative bacteria (inhibitory zone > 20mm). All the test compounds (5a), (5c), (5d), (5g), and (5h) excellent activity and compounds (5b), (5e) and (5f) exhibited moderate activity when compared to that of standard drug fluconazole at the same concentration as the test drugs against *Aspergillus niger*. Compounds (5a), (5c), (5e), (5f) and (5g) excellent activity and compounds (5b), (5d) and (5h) exhibited moderate activity when compared to that of standard drug fluconazole at the same concentration as the test drugs against *Penicillium chrysogonium* (TABLE 1 & 2). The compounds (5a), (5c), (5e), (5g) and (5h) exhibited high antioxidant property in both Nitric Oxide and DPPH methods at 100 µM concentrations (TABLE 3).

REFERENCES

- [1] H.J.Schaeffer, R.N.Johnson, E.Odin, C.Hansch; J.Med.Chem., **13**, 452 (1970).
- [2] H.J.Schaeffer, E.Odin; J.Med.Chem., **10**, 181 (1966).
- [3] H.J.Kelly, E.F.Soroko; J.Med.Chem., **29**, 1133 (1986).
- [4] J.L.Kelley, M.P.Krochmal, J.A.Linn, E.W.Mc Lean, E.F.Soroko; J.Med.Chem., **31**, 606-612 (1988).
- [5] S.A.Lanfer, D.M.Domeyer, T.R.F.Sciort, W.Albrecht, D.R.J.Hauser; J.Med.Chem., **48**, 710-722 (2005).
- [6] S.Banerjee, S.Dutta, K.S.Chakraborti; J.Indian Chem.Soc., **59(3)**, 417-418 (1982).
- [7] J.L.Kelley, E.W.Mc.Lean, R.M.Ferris, J.L.Howerd; J.Med.Chem., **33**, 1910-1914 (1990).
- [8] J.A.Montgomery, Jr., C.Temple; J.Am.Chem.Soc., **80**, 409 (1958).
- [9] S.M.Greeberg, L.O.Ross, R.K.Robins; J.Org.Chem., **24**, 1314 (1959).
- [10] D.J.Brown, N.W.Jacobsen; J.Chem.Soc., 3776 (1965).
- [11] A.Hari, B.L.Miller; Tetrahedron Lett., **40**, 245-248 (1999).
- [12] L.G.J.Hammarstrom, M.E.Meyer, D.B.Smith, F.X.Talamas; Tetrahedron Lett., **44**, 8361-8363 (2003).
- [13] W.R.Boon, W.G.M.Jones, G.R.Ramage; J.Chem. Soc., 96 (1951).
- [14] D.J.Brown; J.Applied Chem., **72**, 4 (1954).
- [15] C.L.Lee, K.P.Chan, Y.Lam, S.Y.Lee; Tetrahedron Lett., **42**, 1167 (2001).
- [16] R.A.Scheuerman, D.Tumelty; Tetrahedron Lett., **41**, 6531 (2000).
- [17] M.K.Schwarz, D.Tumelty, M.A.Gallo; Tetrahedron Lett., **39**, 8397 (1998).
- [18] R.K.Robins, K.L.Dille, B.E.Christensen; J.Org. Chem., **19**, 930 (1954).
- [19] R.K.Robins, K.L.Dille, C.H.Willets, B.E.Christensen; J.Am.Chem.Soc., **75**, 263 (1953).
- [20] N.J.Leonard, T.R.Henderson; J.Am.Chem.Soc., **97(17)**, 4990-4999 (1997).
- [21] B.K.Chun, S.Olgen, J.H.Hang, M.G.Newton, C.K.Chu; J.Org.Chem., **65**, 685-693 (2000).
- [22] A.Albert, D.J.Brown, H.C.S.Wood; J.Chem.Soc., 3832 (1954).