



## Relationships between genetic variability and life-history features of fishes based on S7 ribosomal protein gene

Tangjie Zhang\*

College of Veterinary medicine, Yangzhou University, Jiangsu Yangzhou 225009, (CHINA)

E-mail: slx@yzu.edu.cn

### ABSTRACT

To research relationship between genetic variation and life-history variables of *Actinopterygii*, as indicated by common length, maximum length, maximum weight and longevity, and environmental variation, as indicated by three different fishes' living environments, we applied analysis of independent regression and phylogenetically-independent contrasts methods to evaluate life-history variables correlations with rps7 neutral genetic diversity. Polymorphism datasets of rps7 gene, belonging to 48 genera, 25 families and 9 orders, of *Actinopterygii*, was obtained from Polymorphix and Popset of GenBank. Life-history variables were obtained from the AnAge database and fishbase. The results showed that neutral genetic diversity of fishes is significantly negatively associated with common length. No strong level of correlation was found between fish's neutral genetic diversity and maximum size, maximum weight or maximum longevity. No correlation was found among neutral genetic diversity and fishes' habits (marine, freshwater and marine-freshwater) either. © 2015 Trade Science Inc. - INDIA

### KEYWORDS

Genetic diversity;  
*Actinopterygii*;  
Rps7;  
Life-history.

### INTRODUCTION

There are two principal types of genetic diversity: adaptive and neutral. Interpreting the genetic diversity in organisms with diverse life and population histories can be difficult since the mechanisms and processes that regulate this diversity are complex and still poorly. The causes of variation could be sample error, variance in gene flow, natural selection, genetic drift and so on. The relative importance of these variation factors in shaping the genetic structure of a population could depend on the life history of the species. For example, to explain

mtDNA substitution rate variations, there are some hypotheses-related to life-history- have been proposed: the generation time, the metabolic rate, and the longevity have been advanced to explain variation in DNA substitution rate<sup>[28]</sup>. In *Actinopterygii*, some studies showed that in marine fishes, the level of genetic diversity is higher and the level of population divergence is lower than in freshwater species, most likely due to the better dispersal capabilities and larger effective population sizes of marine species<sup>[14, 37, 11]</sup>.

Appropriate DNA markers must be selected for the phylogenetic and genetic diversity analyses of

fishes. Mitochondrial DNA (mtDNA) is one of the most widely studied genetic markers in the fields of population genetics and phylogeography. However, a more serious criticism is that mtDNA, which is more prone to hitch-hiking because of the lack of recombination, may not conform to neutral expectations<sup>[4, 13]</sup>. The RPS7 gene, which belongs to a family of genes called RPS (ribosomal proteins), has been the most popular marker<sup>[29, 26, 20]</sup> of nuclear neutral molecular diversity in fishes for its high similarity in taxa<sup>[6]</sup>. In this study, the polymorphism of fish RPS7 gene was therefore used to be analyzed for fish's molecular diversity.

The DNA substitution rate is widely used for the studies of genetic diversity, but accurate estimation of mutation rate is obviously required for accurate molecular dating. Another most fundamental in population genetics is the mutation parameter or genetic diversity  $\theta = 4Ne\mu$  under the assumption that mutations are effectively neutral. Here  $\mu$  denotes the expected number of mutations for an individual DNA sequence per generation and  $Ne$  denotes the effective population size. The estimate of  $\theta$  is based on coalescent theory. Watterson's estimator ( $\theta_w$ ) is commonly used for its simplicity. Under the standard neutral model, mutation rates will definitely positively lead to polymorphism levels. Definitely, the analysis of nuclear genes' polymorphism, which are based on increasing molecular genetic data, are useful to understand the mechanism of molecular diversity because they offer broad and comprehensive coverage.

Molecular genetic data have greatly improved our ability to test hypotheses about organisms' evolution. A large amount of nuclear data has been collected from diverse organism's populations dramatically during past decade or so. In this study, we analyzed genetic diversity by surveying polymorphism of fish's *rps7* gene. Our goal of this work was to ascertain if there is a relationship between single nuclear gene (*rps7*) neutral diversity and life-history variables, such as common length, maximum length, maximum weight and longevity, presented above, and a relationship between fish's living environments and neutral genetic diversity.

## MATERIALS AND METHODS

### Data collection

Polymorphism datasets of *rps7* gene was obtained from Polymorphix<sup>[3]</sup> and Popset of GenBank. Two sequences were taken not to be clustered if there was a mismatch of >50nt with <80% similarity flanking the CLUSTALW program<sup>[35]</sup>. Such a mismatch is interpreted as evidence that the sequences represent duplicate genes in distinct genomic contexts. Sequences were inspected by eye and corrected when required.

Data: 146 species of fish belonging to 48 genera and 25 families and 9 orders. Each group was aligned by eye using CLUSTALW. Details of the species sampled are listed in Appendix S.

### Polymorphism sequence data analyses

Genetic diversity measures - Watterson's estimator ( $\theta_w$ ) were calculated separately from the intron and synonymous sites of the analyzed fragments of same species, expressed in per-site level of diversity, and the no. of mutation and sites both synonymous and nonsynonymous sites were counted simultaneously.

### Life history data

Many researches have been done on life history traits in fishes. Kawasaki (1980) suggested that the grouping of life history traits of fishes differed from the traditional methods developed for terrestrial animals<sup>[19]</sup>. In this study, common length, maximum length, maximum weight and longevity, data were obtained from the AnAge database<sup>[10]</sup> and fishbase (<http://www.fishbase.org/home.htm>).

Body length or size was usually used as a proxy for abundance, given the expected negative relationship between body size and abundance. Indeed, a significant negative relationship between body size and genetic variation was found in mammals based on allozymes<sup>[36]</sup>. We got two types of body length, common length and maximum length. Using common length as a proxy for abundance allowed us to increase the number of species evaluated, as catch data are only available for a limited number of species.

### Statistical tests

## Regular Paper

$\theta_w$  were arc-sine transformed<sup>[34]</sup>. Quantitative life-history variables were log transformed. We first calculated  $\theta_s$  for synonymous sites and  $\theta_i$  for intron sites and weighted average of  $\theta_s$  or /and  $\theta_i$  from rps7 for same species. Analysis of independent regression methods was applied to evaluate one or more life-history correlations with neutral genetic diversity ( $\theta$ ) (synonymous and intron sites).

### Phylogenetic reconstructions

We created a phylogenetic hypothesis for the species included in this study by grafting them onto a higher-level phylogenetic supertree of *Actinopterygii* using PhyloWidget<sup>[16]</sup>. The topology was a composite of information drawn from TreeBASE (<http://www.treebase.org/treebase-web/home.html>). Phylogenetically-independent contrasts (PIC) were conducted using Phylogenetic Comparative Methods of COMPARE, version 4.6b<sup>[23]</sup>.

## RESULTS

### Neutral genetic diversity of fishes based on rps7

A smaller amount of life-history data is available especially for max weight, which documented only 17 species in 146 fish species. We thus focused on the effects on maximum size and common length. The common length, however, could be taken as a proxy of population size<sup>[21, 29]</sup>.

To correlate life-variables to all neutral sites' diversity of sampled S7 ribosomal protein gene, we weighted average  $\theta$  by combining intron with synonymous and performed nonphylogenetic regression

(logistic regressions) analyses and phylogenetically regression of independent contrasts. Both the application of phylogenetically independent contrasts and logistic regression analyses showed that  $\theta_w$  of weighted neutral sites was significantly negatively associated with common length ( $n=67$ ,  $p=0.031$ ,  $n=67$ ,  $p=0.011$ ). No strong level of correlation was found between  $\theta_w$  of weighted neutral sites and maximum size, maximum weight or maximum longevity (TABLE 1).

To investigate the relationship between fish's living environments and neutral genetic diversity, we correlated environments to all neutral sites' diversity of sampled S7 ribosomal protein gene on the basis of three entirely different environments: marine, freshwater and marine-freshwater (TABLE 1). The result showed that no correlation was found between neutral genetic diversity and fish's habits ( $n=143$ ,  $p=0.120$ ).

## DISCUSSION

Early studies have revealed marked differences in the level of genetic differentiation and genetic diversity between populations of marine and freshwater species, with marine species generally exhibiting lower levels of inter-population differentiation and higher genetic diversity<sup>[14, 37, 11]</sup>. This general observation has generally been hypothesized to be a result of higher effective population sizes and/or higher inter-population migration rates in marine, compared to freshwater environments and has implications for the conservation of genetic diversity. Lower effective population sizes and/or lower in-

TABLE 1 : Single variable regressions of weighted neutral (introns and synonymous) sites

Trait	N	Non-phylogenetic			Phylogenetic regression of independent contrasts		
		Slope	R <sup>2</sup>	p-value	Slope	R <sup>2</sup>	p-value
Max size	130	-0.001	0.024	0.078	-0.002	0.036	0.05
Common length	67	-0.001	0.069	0.031*	-0.001	0.087	0.011*
$\theta_w$ Max weight	17	0.000	0.023	0.545	-0.001	0.053	0.865
Max Longevity	36	0.001	0.020	0.408	0.002	0.013	0.565
Environment	143	-0.001	0.007	0.310	-0.002	0.008	0.120

ter-population migration rates in the freshwater environment predict that populations of freshwater species are expected to be more prone to extinction than marine species and thus should be of particular conservation concern.

Gene diversities of neutral molecular markers are most influenced by effective population size ( $N_e$ )<sup>[9, 24]</sup>. Diversities are expected to be high in large, stable populations because the magnitude of random drift is less, leading to the retention of a larger proportion of existing diversity and a greater number of new mutations. Although early studies have also indicated that marine fish species have low levels of population genetic differentiation when compared to freshwater species<sup>[37, 11]</sup>, more studies have shown deep divergences on marine species populations. In some cases, the degree of differentiation is so high that speciation events are proposed<sup>[18, 27, 31, 7, 5]</sup>. Marine organisms disperse much more due to high connectivity between their habitats. Therefore, the degree of genetic differentiation in this environment is attenuated. However, there are some other limits (spatial, directional or temporal) to dispersion that may promote the genetic differentiation in marine organisms. Amongst the different types of limits are physical barriers such as patterns of oceanic circulation, water temperature, gradients of salinity, restrict dispersion of eggs, larval or adults, phylopatry, selection and historic events such as glaciations<sup>[33, 30]</sup>.

The assumptions on the relationship between body weight/length and genetic polymorphism for certain loci, which were confirmed by correlation analysis in small samples based on isozymes, were made<sup>[17, 18, 12]</sup>. Under natural conditions, one indeed could expect a strong negative relationship between the body length and the proportion of polymorphism<sup>[1, 2]</sup>.

In this study, examination of neutral genetic diversity was based on S7 gene DNA polymorphisms. It was shown that neutral genetic diversity of fishes, measured by  $\theta_w$ , Watterson's estimator, is significantly negatively associated with common length. Compared to longer length fishes, shorter fishes

display higher rates of generation change, resulting in accelerated growth of population size and faster accumulation of genetic variability, which was also found in Vertebrates<sup>[12, 25]</sup>. Based on common length, used as a proxy for effective population size, our result was with consistent the concept that genetic diversity showed higher in large populations, although this concept has been challenged<sup>[28, 4]</sup>

Metabolic rate hypotheses proposed that the production of mutagenic free radicals, reactive oxygen species (ROS), increases with increasing rates of respiration; therefore, so does the rate of mutation<sup>[25, 22]</sup>. Taking body weight as a proxy of metabolic rate, the results of this study show that there was no significant correlation between  $\theta_w$  of fish weighted neutral sites and metabolic rate. Neither did we find a significant correlation between neutral fish diversity and longevity after multiple regression analyses, which indicated that common length was the most crucial factor of all variables included in affecting nuclear neutral diversity.

Anadromy is a life-history trait of fishes that refers to migration from a freshwater breeding habitat to a marine feeding habitat and back to freshwater for spawning. Several summaries of genetic diversity in fishes have been published<sup>[14, 37, 11]</sup>. These have shown that there is some evidence the level of genetic diversity and population divergence of anadromous species is intermediate to that of marine and freshwater species. This seems logical given that, for example, the level of between populations migrations may be expected to be higher in rivers connected to the ocean compared to completely isolated water bodies. There, however, has been little statistical support for some of these findings, most probably due to the small number of anadromous populations assessed and to the fact that earlier studies have compared genetic diversity indices based on completely different sets of loci. In this study, we also found there is no correlations between neutral genetic diversity and habits (marine, freshwater and marine-freshwater), even though the anadromous fishes (10 species) sampled were limited.

*Regular Paper*

## Appendix S Species sampled for analyses of rps7 gene polymorphism

Order	Families	Genera	Species			
Aulopiformes	Haemulidae	Haemulon	Haemulon parra			
			Haemulon scudderi			
	Balitoridae	Oxynoemacheilus	Oxynoemacheilus	Oxynoemacheilus bureschi		
				Cobitidae	Cobitis	Cobitis bilineata
						Cobitis elongata
						Cobitis ohridana
						Cobitis taenia
						Cobitis tanaitica
				Dionda	Dionda	Codoma
						Codoma ornata
						Dionda episcopa
				Gobio	Gobio	Dionda ipni
						Dionda melanops
						Dionda nigrotaeniata
Gobio carpathicus						
Cypriniformes	Cyprinidae	Gobio	Gobio caucasicus			
			Gobio gobio			
			Gobio insuyanus			
			Lepidomeda			
			Lepidomeda aliciae			
			Lepidomeda copei			
			Notropis			
			Notropis calientis			
			Pimephales			
			Pimephales notatus			
Richardsonius	Richardsonius	Richardsonius	Pimephales promelas			
			Pimephales tenellus			
			Richardsonius balteatus			
			Richardsonius egregius			
			Rutilus			
			Rutilus frisii			
			Tinca			
			Tinca tinca			
Tribolodon	Tribolodon	Tribolodon	Tribolodon brandtii			
			Tribolodon hakonensis			
			Yuriria			
			Yuriria alta			
Chondrostoma	Chondrostoma	Chondrostoma	Chondrostoma duriense			
			Chondrostoma nasus			
			Chondrostoma polylepis			
			Chondrostoma toxostoma			
			Chondrostoma willkommii			
Cyprinodontiformes	Cyprinodontidae	Poecilia	Poecilia dauli			
			Poecilia gillii			
Gasterosteiformes	Macroramphosidae	Macroramphosus	Macroramphosus scolopax			
	Syngnathidae	Hippocampus	Hippocampus mohnikei			
			Hippocampus reidi			

Order	Families	Genera	Species			
Osmeriformes	Osmeridae	Hypomesus	Hypomesus japonicus			
			Hypomesus nipponensis			
			Hypomesus olidus			
			Hypomesus pretiosus			
			Hypomesus transpacificus			
		Osmerus	Osmerus eperlanus			
			Osmerus mordax			
			Spirinchus lanceolatus			
			Spirinchus starksi			
		Thaleichthys	Thaleichthys pacificus			
			osteoglossiformes	Mormyridae	Campylomormyrus	Campylomormyrus compressirostris
						Campylomormyrus numenius
						Campylomormyrus rhynchophorus
Campylomormyrus tamandua						
Perciformes	Blenniidae	Salaria	Salaria economisidi			
			Salaria fluviatilis			
			Salaria pavo			
	Tripterygiidae	Tripterygion	Tripterygion delaisi			
			Tripterygion melanurus			
			Tripterygion tripteronotus			
	Blenniidae	Coryphoblennius	Coryphoblennius galerita			
	Centrarchidae	Pomoxis	Pomoxis nigromaculatus			
	Chaenopsidae	Acanthemblemaria	Acanthemblemaria macrospilus			
	Cichlidae	Lepidiolamprologus	Lepidiolamprologus kendalli			
			Lepidiolamprologus nkambae			
	Clinidae	Clinus	Clinus cottoides			
			Clinus superciliosus			
		Muraenoclinus	Muraenoclinus dorsalis			
			Gillichthys	Gillichthys mirabilis		
		Mesogobius	Gillichthys seta			
			Mesogobius batrachocephalus			
		Neogobius	Apollonia fluviatilis			
			Apollonia melanostoma			
Gobiidae	Proterorhinus	Proterorhinus semipellucidus				
		Proterorhinus marmoratus				
	Neogobius	Proterorhinus semilunaris				
		Neogobius eurycephalus				
			Neogobius gorlap			
			Neogobius gymnotrachelus			
			Neogobius Kessler			
			Haemulidae	Anisotremus	Anisotremus caesius	
Anisotremus davidsoni						
Anisotremus davidsonii						
Anisotremus dovii						
Anisotremus interruptus						
Anisotremus moricandi						
Anisotremus pacifici						
Anisotremus scapularis						
Anisotremus surinamensis						
Anisotremus taeniatus						
Anisotremus virginicus						

**Regular Paper**

Order	Families	Genera	Species
		Halichoeres	Halichoeres bivittatus
	Labridae	Thalassoma	Thalassoma hardwicke Thalassoma janseni Thalassoma quinquevittatum
		Pagothenia	Pagothenia borchgrevinki
	Nototheniidae	Trematomus	Trematomus bernacchii Trematomus eulepidotus Trematomus hansonii Trematomus loennbergii Trematomus newnesi Trematomus nicolai Trematomus pennellii.fasta Trematomus scotti
		Crystallaria	Crystallaria asprella Crystallaria cincotta
			Etheostoma basilare Etheostoma blennioides Etheostoma burri Etheostoma caeruleum Etheostoma camurum Etheostoma chlorobranchium Etheostoma derivativum Etheostoma fragi Etheostoma longimanum Etheostoma maculatum Etheostoma mariaae Etheostoma microlepidum Etheostoma nigrum Etheostoma obeyense Etheostoma olmstedi Etheostoma perlongum Etheostoma planasaxatile Etheostoma podostemone Etheostoma punctulatum Etheostoma rufilineatum Etheostoma sanguifluum Etheostoma simoterum Etheostoma smithi Etheostoma spectabile Etheostoma susanae Etheostoma uniporum Etheostoma virgatum Etheostoma vitreum
	Percidae	Etheostoma	
		Percina	Percina burtoni Percina caprodes Percina fulvitaenia
	Scaridae	Scarus	Scarus ghobban
	Gobiidae	Zosterisessor	Zosterisessor ophiocephalus
Pleuronectiformes	Pleuronectidae	Hypsopsetta	Hypsopsetta guttulata
			Pseudoplatystoma corruscans Pseudoplatystoma fasciatum Pseudoplatystoma metaense Pseudoplatystoma orinocoense Pseudoplatystoma reticulatum Pseudoplatystoma tigrinum
Siluriformes	Pimelodidae	Pseudoplatystoma	

## ACKNOWLEDGMENTS

This research was supported by the Priority Academic Program Development of Jiangsu Higher Education Institutions and Jiangsu Co-innovation Center for Prevention and Control of Important Animal Infectious Diseases and Zoonoses.

## REFERENCES

- [1] Altukhov, YuP.; Allozyme Heterozygosity Maturation Rate, and Life Span, *Genetika (Moscow)*, **34**(7), 908–919 (1998).
- [2] Altukhov, YuP.; Genome Heterozygosity, Metabolic Intensity, and Life Span, *Dokl.Akad.Nauk*, **369**(5), 704–707 (1999).
- [3] E.Bazin, L.Duret, S.Penel, N.Galtier; Polymorphix, a sequence polymorphism database, *Nucleic Acids Res.*, **33**, D481–484 (2005).
- [4] E.Bazin, S.Glemin, N.Galtier; Population size does not influence mitochondrial genetic diversity in animals, *Science*, **312**, 570–572 (2006).
- [5] J.L.Carlin, D.R.Robertson, B.W.Bowen; Ancient divergences and recent connections in two tropical Atlantic reef fishes *Epinephelus adscensionis* and *Rypticus saponaceus* (Percoidei: Serranidae), *Mar.Biol.*, **143**, 1057–1069 (2003).
- [6] F.Cecconi, C.Crosio, P.Mariottini, G.Cesareni, M.Giorgi, S.Brenner, F.A.Amaldi; Functional Role for Some Fugu Introns Larger Than the Typical Short Ones: The Example of the Gene Coding for Ribosomal Protein S7 and snoRNA U17, *Nucleic Acids Res.*, **24**, 3167–3172 (1996).
- [7] S.F.Chenoweth, J.M.Hughes, R.C.Connolly; Phylogeography of the pipefish, *Urocampus carinirostris*, suggests secondary intergradation of ancient lineages, *Mar.Biol.*, **141**, 541–547 (2002).
- [8] J.Colborn, R.E.Crabtree, J.B.Shaklee, E.Pfeiler, B.W.Bowen; The evolutionary enigma of bonefishes (*Albula* spp.): cryptic species and ancient separations in a globally distributed shorefish, *Evol.* **55**, 807–820 (2001).
- [9] J.F.Crow, M.Kimura; An introduction to population genetic theory, Harper and Row, New York, (1970).
- [10] J.P.De Magalhaes, J.Costa, O.Toussaint; HAGR: the human ageing genomic resources, *Nucleic Acids Res.*, **33**, D537–D543 (2005).
- [11] J.A.DeWoody, J.C.Avise; Microsatellite variation in marine, Freshwater and anadromous fishes compared with other animals, *J.Fish.Bio.*, **56**, 461–473 (2000).
- [12] A.N.Evsvukov, M.V.Oficerov, I.V.Kononov; Analysis of correlation between body length and genetic polymorphism for loci ESTD-1 and IDHP-3 in the atlantic salmon *salmo salar*, *Russ.J.Genet.*, **38**(7), 810–815 (2002).
- [13] W.S.Grant, I.B.Spies, M.F.Canino; Biogeographic evidence for selection on mitochondrial DNA in North Pacific walleye pollock *Theragra chalcogramma*, *J.Heredity.*, **97**, 571–580 (2006).
- [14] U.Gyllensten; The genetic structure of fish: differences in the intraspecific distribution of biochemical genetic variation between marine, Anadromous, and freshwater species, *J.Fish Bio.*, **26**, 691–699 (1985).
- [15] S.He, R.Mayden, X.Wang, W.Wang, K.L.Tang, Y.Chen; Molecular phylogenetics of the family Cyprinidae (Actinopterygii: Cypriniformes) as evidenced by sequence variation in the first intron of S7 ribosomal protein-coding gene: Further evidence from a nuclear gene of the systematic chaos in the family, *Mol.Phylogenet Evol.*, **46**, 818–829 (2008).
- [16] G.Jordan, W.Piel; PhyloWidget: Web based visualizations for the tree of life, *Bioinformatics*, **24**, 1641–1642 (2008).
- [17] W.C.Jordan, A.F.Youngson, J.H.Webb; Genetic variation at the malic enzyme-2 locus and age at maturity in sea-run atlantic salmon (*salmo salar*), *Can.J.Fish Aquat.Sci.*, **47**(9), 1672–1677 (1990).
- [18] W.C.Jordan, A.F.Youngson; Genetic protein variation and natural selection in atlantic salmon (*salmo salar* L.) *Parr.J.Fish Biol* **39**(supply A), 185–192 (1991).
- [19] T.Kawasaki; Fundamental relations among the selections of life history in marine teleosts, *Bull.Jap.Soc.Sci.Fish*, **46**, 289–293 (1980).
- [20] H.López-Fernández, K.O.Winemiller, R.L.Honeycutt; Multilocus phylogeny and rapid radiations in Neotropical cichlid fishes (Perciformes: Cichlidae: Cichlinae), *Mol.Phylogenet.Evol.*, **55**(3), 1070–1086 (2010).
- [21] A.P.Martin, S.Palumbi; Body size, metabolic-rate, Generation time, and the molecular clock, *Proc.Natl.Acad.Sci., USA*, **90**, 4087–4091 (1993).
- [22] A.P.Martin, G.Naylor, S.Palumbi; Rates of mitochondrial DNA evolution in sharks are slow compared with mammals, *Nature*, **357**, 153–155 (1992).
- [23] E.P.Martins; Compare, Version 4.6b, Computer programs for the statistical analysis of comparative data, Distributed by the author at <http://>

## Regular Paper

---

- compare.bio.indiana.edu/ Department of Biology, Indiana University, Bloomington IN, (2004).
- [24] M.R.Mccusker, P.Bentzen, M.R.Mccusker, P.Bentzen; Positive relationships between genetic diversity and abundance in fishes, *Mol.Ecol.*, **19**(22), 4852–4862 (2010).
- [25] S.V.Mezhzhherin; Correlation between genetic variability and body size in vertebrates, *Russ.J.Genet.*, **38**(9), 1060–1065 (2002).
- [26] Z.Musilová, O.Rícan, K.Janko, J.Novák; Molecular phylogeny and biogeography of the Neotropical cichlid fish tribe Cichlasomtini (Teleostei: Cichlidae: Cichlasomatinae), *Mol.Phylogenet.Evol.*, **46**, 659–672 (2008).
- [27] A.Muss, D.R.Robertson, C.A.Stepien, P.Wirtz, B.W.Bowen; Phylogeography of *Ophioblennius*: the role of ocean currents and geography in reef fish evolution, *Evol.*, **55**, 561–572 (2001).
- [28] B.Nabholz, S.Glemin, N.Galtier; The erratic mitochondrial clock: variation of mutation rate, Not population size, Affect mtDNA diversity across birds and mammals, *BMC Evol.Boil.*, **9**, 54 (2009).
- [29] B.Nabholz, S.Glémin, N.Galtier; Strong variations of mitochondrial mutation rate across mammals-the longevity hypothesis, *Mol.Biol.Evol.*, **25**, 120–130 (2008).
- [30] S.R.Palumbi; Genetic divergence, Reproductive isolation, And marine speciation, *Annu.Rev.Ecol.Syst.*, **25**, 547–572 (1994).
- [31] S.Planes, P.J.Doherty, G.Bernardi; Strong genetic divergence among populations of a marine fish with limited dispersal, *Acanthochromis polyacanthus*, With in the great barrier reef and the coral sea, *Evol.*, **55**, 2263–2273 (2001).
- [32] O.Rícan, R.Zardoya, I.Doadrio; Phylogenetic relationships of Middle American cichlids (Cichlidae, Heroini) based on combined evidence from nuclear genes, mtDNA, and morphology, *Mol.Phylogenet.Evol.*, **49**(3), 941–957 (2008).
- [33] M.Sinclair; Marine populations: An essay on population regulation and speciation, University of Washington Press, Seattle, (1988).
- [34] R.R.Sokal, F.Rohlf, Biometry.W.H.Freeman, San Francisco, (1981).
- [35] J.D.Thompson, D.G.Higgins, T.J.Gibson; Clustal W: Improving the sensitivity of progressive multiple sequence alignment through sequence weighting, Position-specific gap penalties and weight matrix choice, *Nucleic.Acids Res.*, **22**: 4673–4680 (1994).
- [36] M.C.Wooten, M.H.Smith; Large mammals are genetically less variable? *Evol.*, **39**, 210–212 (1985).
- [37] R.D.Ward, M.Woodwark, D.O.F.Skibinski; A comparison of genetic diversity levels in marine, freshwater, and anadromous fishes, *J.Fish Bio.*, **44**, 213–232 (1994).