Potentiality of *Bacillus weihenstephanensis* PKD5 Keratinase for Eco-Friendly Dehairing of Skins and Hide

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Abstract

Comparative dehairing efficiency of Na₂S (1.5%), CaO (1.5%) and crude keratinase (15.3 U/ml) of *Bacillus weihenstephanensis* PKD5 was investigated in single and in combination on cow hide, goat and sheep skin. Complete dehairing was achieved by treatment with Na₂S, CaO with Na₂S, Na₂S with crude keratinase and crude keratinase after 2, 2, 2 and 10 h respectively. Among all the treatments, the enzyme treated skin/hide was comparatively more white, smooth and shiny due to release of intact hair with follicle from skin/hide. Though chemical(s) and combination of chemical with enzyme dehair the skin/hide in relatively less time but the enzymatic dehairing improved the leather quality. So, the keratinase is produced using low cost poultry feather which can be employed for dehairing that is sustainable and potential to replace the chemical dehairing in leather industries.

Keywords: *Bacillus weihenstephanensis*; PKD5; Keratinase; Leather; Dehairing

Introduction

Leather industry has been a traditional industrial sector for the production of processed leather. It is a by product of meat industry which is prepared from skin and hide of animal. It was estimated that worldwide 5.5 million tons of wet salted raw hides were processed and $4.6 \times 10^5$ tons of heavy leathers was produced per year [1]. The leather processing involves various operations in a sequential manner from raw skin/hide to processes leather. Classical leather processing involves the major steps like soaking, dehairing, bating and tanning.

Dehairing or unhairing is an essential and unavoidable step to remove hair from skin and hide. After dehairing, the leather processed through many treatments like deliming, bating, degreasing, and pickling in a cascade manner; then it converts into a durable, long-lasting material for various uses including shoes, hats, jackets, skirts, trousers, belts, book binding, leather wallpaper, and as a furniture covering. Conventionally, lime (CaO) and sulphide (Na₂S/NaSH) used concurrently for dehairing for a long time [2]. Lime as an alkali causes swelling of skins/hides by hydrolyzing asparagine and glutamine in

collagen structure that helps to distort fiber bundles to remove hairs [3,4]. Whereas sulphide like Na₂S/NaSH was used for dehairing, as the sulphide ion attack the disulphide bridges between cysteine residues present in keratin structure [5]. The production and quality of leather did not reach to optima due to lack of technology other than the use of lime and sodium sulphide.

The two main ingredients for dehairing was lime which produces a poisonous sludge whereas sulphide is highly toxic with obnoxious odour. They cause pollution in tannery’s wastewater which contaminates community water sources. They may also decline the efficiency of treatment plants to recycle the effluents [6]. Exposure to chemicals in air or in solution tanks in processing plants can be hazardous to workers and the symptoms are skin irritations, respiratory problems and dizziness. In this concern, the enzymatic dehairing approach, specifically protease is a better, greener and sustainable alternative to reduce environmental pollution, harmful health effects on workers and also improve the quality of leather [7-10].

Few reports on dehairing attested that employment of microbial proteases provide better quality leather in contrast with chemical dehairing agent [11-16]. Among the microbial sources, bacterial proteases gain much more appreciation and attention due to easy production through submerged fermentation, relatively higher yield, less production time and easy recovery of product. However commercial implementation of enzymatic dehairing is limited due to certain drawback like stability of enzyme in wide range of pH and temperature, consistent performance as well as production cost.

With the aim to search for a viable and eco-friendly alternative of the harmful chemicals, the present study was aimed to evaluate the dehairing efficiency of crude keratinolytic protease of *Bacillus weihenstephanensis* PKD5 along with Na₂S and CaO (as chemical dehairing agent) in single and in combination.

**Materials and Methods**

**Microorganism and keratinase production**

A potent keratinase producing bacteria *Bacillus weihenstephanensis* PKD5 (GenBank accession no. JN897383) was previously isolated from soil sample of feather dumping area was employed in present study [17].

The strain was grown in 250-ml Erlenmeyer flasks in optimized condition (g/l): NaCl 5.0, MgSO₄ 1.0, K₂HPO₄ 1.0, (NH₄)₂SO₄ 2.0, white chicken feather 10.0, at pH 8.0, 2% inoculum (10⁹ cells/ml), 40°C, 120 rpm for 7 days [17]. Thereafter the fermented broth was centrifuged (at 5590×g for 5 min) and the supernatant was considered as crude enzyme.

**Enzyme assay**

The keratinolytic activity of crude enzyme was assayed with keratin powder (Hi-Media, India) as a substrate with the modified method of Gradisar et al. [18]. One unit of keratinase activity was defined as the amount of enzyme required to liberate 1 μg of tyrosine per minute under the experimental conditions.

**Dehairing assay**

Fresh fleshed skin of goat; sheep and cow hide were collected from local slaughter house, washed sequentially with sodium chloride solution and water to remove impurities then dried in hot air oven at 50°C. The skins were cut into pieces and
incubated with 40 ml solution in 100 ml beaker (dip method) of different agents viz. 1.5% sodium sulphide (Na$_2$S), 1.5% lime (CaO) and crude enzyme (15.3 U/ml) in single and in combination (1.5% CaO+1.5% Na$_2$S, 1.5% lime+15.3 U/ml keratinase and 1.5% Na$_2$S+15.3 U/ml keratinase) at room temperature. Skin/hide treated with distilled water was considered as negative control. Skin/hide were withdrawn at regular interval and examined for dehairing by gently scraping with blunt scalpel and the process is continued up to 10 h. Skin pieces and hair were analyzed through stereomicroscope (Magnus MS24). The experiment was carried out in triplicate. After one batch, the used enzyme (single and in combination) also employed upon fresh skins to see its dehairing efficiency.

Results

Effect of different agent(s) on dehairing of skins/hide was investigated in time dependent manner and the appearance of dehaired skins and hide was shown in FIG. 1. Complete dehairing was noticed after treatment with Na$_2$S (1.5%), lime+Na$_2$S (1.5%) and Na$_2$S+ crude keratinase within 2 h of incubation in all the three types of skin/hide. On the other hand in crude enzyme treatment, complete dehairing was achieved within 10 h (FIG. 1). The visual and stereomicroscopic examination revealed that the proteolytic enzyme removed the hairs with follicles as a result the skins become smooth, white and shiny. On the other hand, the chemically treated skins were hard, dark brown and wrinkled (FIG. 2).

<table>
<thead>
<tr>
<th>Skin</th>
<th>Negative Control</th>
<th>Surface area±SD (cm$^2$)</th>
<th>Weight ±SD (g)</th>
<th>Lime</th>
<th>Na$_2$S</th>
<th>Lime + Na$_2$S</th>
<th>Keratinase</th>
<th>Keratinase + Lime</th>
<th>Keratinase + Na$_2$S</th>
</tr>
</thead>
<tbody>
<tr>
<td>Goat</td>
<td>5.92±1.78</td>
<td>7.83±1.86</td>
<td></td>
<td>10 h</td>
<td>2 h</td>
<td>2 h</td>
<td>10 h</td>
<td>10 h</td>
<td>10 h</td>
</tr>
<tr>
<td>Cow</td>
<td>6.92±1.92</td>
<td>8.12±2.65</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sheep</td>
<td>5.84±1.73</td>
<td>7.66±1.69</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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</tr>
</tbody>
</table>

FIG. 1. Dehairing efficiency of CaO, Na$_2$S and crude keratinase in single and in combination.
FIG. 2. Stereomicroscope picture (10X) showed the dehaired skin of goat (a, d, g), cow (b, e, h, k) and sheep (c, f, i, l).

a-c: Na$_2$S treated, d-f: enzyme+Na$_2$S treated, g-i: lime+Na$_2$S treated, j-l: enzyme treated.

Hair collected from traditional dehairing through the lime-sulfide process was found to be gelatinized and subsequently converted into pulp (FIG. 3 shows Na$_2$S+lime treatment only), whereas hairs collected from enzymatic treatment was intact with follicle (FIG. 4). The used enzyme after one batch could also dehaired all types of skins/hide efficiently within 14 h of incubation (FIG. 5) with mild obnoxious odour.

FIG. 3. Photograph of hair pulp produced by lime + Na$_2$S treatment (a. goat, b. Cow, c. Sheep).
FIG. 4. Stereoscopic microscopic view of hair of goat (a), cow (b) and sheep (c) observed after enzymatic dehairing. Inset pictures are the respective enlarged views showing the hair follicles.

FIG. 5. Photograph showing the enzymatic dehairing of skins and hide after reuse of keratinase enzyme (a. goat, b. cow, c. sheep).

Discussion

In our previous assignment, we explored a potent keratinase producing bacteria *B. weihenstephanensis* PKD5 which capable to produce the enzyme by utilizing low cost chicken feather [17] that will be highly acceptable in industrial sector for large scale production. The enzyme was found stable in wide range of pH (6-9) and temperature (30°C to 60°C) which encourage its application in leather processing [17]. In the present study, the crude keratinase of *B. weihenstephanensis* PKD5 takes about 10 h for dehairing (at 15.3 U/ml enzyme concentration, pH 8.0, room temperature) that is comparatively better than the reported protease of Bacillus subtilis [19], Pseudomonas stutzeri strain K4 [20], *Bacillus safensis* [21], *Paenibacillus woonsonensis* (TKB2) [22] which take 24 h, 20 h, 16 h, 14 h respectively (TABLE 1). Though, the purified enzyme of *Brevibacillus brevis* US575 takes about 10 h for dehairing [23], but the purification process is costly, time consuming and sensitive. Therefore, the application of crude enzyme eliminates the cumbersome purification process. Moreover, the presence of other proteolytic enzymes in crude preparation may enhance or they may act synergistically with keratinase which in turn hasten the dehairing efficiency.
TABLE 1. Comparison of dehairing conditions of *B. weihenstephanensis* PKD5 with reported bacterial keratinases/proteases.

<table>
<thead>
<tr>
<th>Bacterial strain</th>
<th>Enzyme (Purified /Partially purified/ Crude) and its concentration/amount</th>
<th>Dehairing condition</th>
<th>Type of skin/hide</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Paenibacillus woosongensis</em> TKB2</td>
<td>Crude enzyme (3:1 w/v)</td>
<td>14 h, pH 8.9</td>
<td>Goat</td>
<td>[22]</td>
</tr>
<tr>
<td><em>Pseudomonas stutzeri</em> strain K4</td>
<td>Crude enzyme (40 U/ml)</td>
<td>20 h, pH 8.0, 30°C</td>
<td>Goat</td>
<td>[20]</td>
</tr>
<tr>
<td><em>Bacillus subtilis</em></td>
<td>Crude enzyme (59 U/ml)</td>
<td>24 h, pH 8.0, 37°C</td>
<td>Goat</td>
<td>[19]</td>
</tr>
<tr>
<td><em>Bacillus safensis</em></td>
<td>Crude enzyme (35.4 U/ml to 50.4 U/ml)</td>
<td>16 h, 30°C ± 2°C</td>
<td>Goat</td>
<td>[21]</td>
</tr>
<tr>
<td><em>Brevibacillus brevis</em> US575</td>
<td>Purified (2,000 U/ml)</td>
<td>10 h, 37°C</td>
<td>Rabbit, goat, sheep, bovine</td>
<td>[23]</td>
</tr>
<tr>
<td><em>Bacillus cereus</em> strain AT</td>
<td>Crude enzyme (4813 U/g ± 62 U/g)</td>
<td>18 h, pH 9.0, 30°C</td>
<td>Goat</td>
<td>[24]</td>
</tr>
<tr>
<td><em>Pseudomonas aeruginosa</em> MCM B-327</td>
<td>Partially purified (1%, w/w in 20% water)</td>
<td>16-21 h, pH 7.0, 26°C to 30°C</td>
<td>Buffalo</td>
<td>[25]</td>
</tr>
<tr>
<td><em>Exiguobacterium</em> sp. DG1</td>
<td>Crude enzyme</td>
<td>24 h</td>
<td>Sheep</td>
<td>[26]</td>
</tr>
<tr>
<td><em>Bacillus circulans</em></td>
<td>Purified (2 mg/50 ml)</td>
<td>16 h, pH 11.0, 35°C</td>
<td>Goat</td>
<td>[27]</td>
</tr>
<tr>
<td><em>Bacillus weihenstephanensis</em> PKD5</td>
<td>Crude enzyme (15.3 U/ml)</td>
<td>10 h, pH 8.0, room temperature</td>
<td>Goat, cow, sheep</td>
<td>Present study</td>
</tr>
</tbody>
</table>

The chemicals and the cocktail of enzyme with chemical synergistically accelerate the dehairing process though enzymatic dehairing removes intact hair whereas chemical dehairing produces hair-pulp. So, enzyme-based dehairing significantly reduces the chance of pollution, eliminates the use of toxic chemicals as well as improves the leather quality. Moreover, the intact hair (especially sheep) recovered from enzymatic dehairing may use in textile industry.

**Conclusion**

In this study, an eco-friendly enzymatic dehairing process was developed using keratinolytic protease of *B. weihenstephanensis* PKD5 produced by utilizing low cost chicken feather, which is more effective in various classes of skins/hide in addition to decrease pollution load and improve leather quality. Worldwide, search is going on for a stable enzyme which will be more effective on various types of hide and skin. In this concern, large scale enzyme production and subsequent application for dehairing may cope with the scale of normal leather production at industrial level.

**REFERENCES**


