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## Naturally Occurring Bioactive *o*-Heterocycles: A Quest For New Sources



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### ABSTRACT

*Limnophila heterophylla* and *L. rugosa* (Scrophulariaceae) were established as new sources of natural flavonoids, respectively of nevadensin (5,7-dihydroxy-6,8,4'-trimethoxyflavone) and salvigenin (5-hydroxy-6,7,4'-trimethoxyflavone). Their structures were characterized on the basis of spectral studies. © 2006 Trade Science Inc. - INDIA

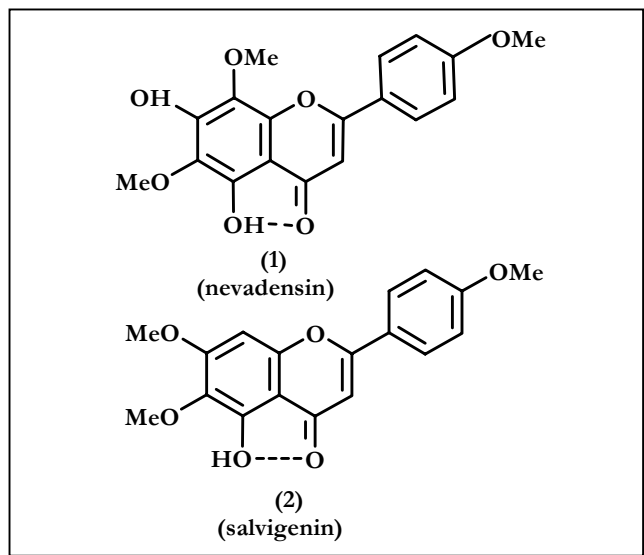
### KEYWORDS

*Limnophila heterophylla*;  
*L. rugosa*;  
Scrophulariaceae;  
Flavonoids; Nevadensin;  
Salvigenin; Spectral studies.

### INTRODUCTION

Among the naturally occurring heterocyclic compounds, *O*-heterocycles are mostly abundant and are of importance and interest to a wide variety of scientists from interdisciplinary fields<sup>[1,2]</sup>. In continuation of our search for natural *O*-heterocycles<sup>[3-7]</sup>, we have found *limnophila heterophylla*<sup>[8]</sup> and *limnophila rugosa* (family: Scrophulariaceae)<sup>[9,10]</sup> as new and rich sources, respectively for nevadensin<sup>[11,12]</sup> and salvigenin<sup>[13]</sup>. Both are the well-known natural bioactive fla-

vonoids. As reported so far, nevadensin possesses anti-mycobacterial and anti-inflammatory activities<sup>[14]</sup>, hypotensive property<sup>[15]</sup>, and moderate cytotoxic and antitubercular activities also<sup>[16]</sup>. Salvigenin is also an important bioactive bioflavonoid<sup>[17]</sup>. *L. heterophylla* and *L. rugosa*, both locally available herbs, are important Indian medicinal plants and find a lot of applications in the traditional system of medicine against various ailments<sup>[4, 9, 18-20]</sup>. The petrol extracts of both the plant materials afforded the flavonoids characterized on the basis of detailed spectral studies.



## RESULTS AND DISCUSSION

The dihydroxytrimethoxy flavone **(1)**,  $C_{18}H_{16}O_7$  ( $[M]^+$  at  $m/z$  344), responded positively towards flavonoid colour reactions and ferric chloride solution, and exhibited characteristic UV absorptions. The infrared absorption bands of **(1)** are also of expected outcome. In the  $^1H$  NMR spectrum of nevadensin, we observed for the first time that the B-ring protons appeared as double doublets (dd) — d 7.89 (2H, dd,  $J= 2.7$  Hz, 11.7 Hz, H-2' & H-6') and d 7.045 (2H, dd,  $J= 3$  Hz, 11.7 Hz, H-3' & H-5') — thereby supporting the proposed B-ring substitution pattern.

The mass spectral fragmented ion-peaks of compound **(1)** clearly suggest that two methoxyl and two hydroxyl functions are attached to the ring-A, while the remaining methoxyl group is linked with the ring-B, and unambiguously it must be placed at C-4' as evidenced from the  $^1H$  NMR spectral analysis. The appearance of intense green colour with ferric chloride imparted by the compound locates one of the hydroxyls at C-5 position<sup>[21]</sup> as also revealed from IR and  $^1H$  NMR spectra. Again, the bathochromic shift of Band I by 20 nm (335→355) in the UV spectrum of **(1)**, in the presence of aluminium chloride that remained unchanged on addition of hydrochloric acid, confirmed the presence of a hydroxyl function at C-5 and one of the methoxyls at C-6 position<sup>[22, 23]</sup>. That the C-6 position is blocked by a methoxyl function, is evidenced by its characteristic mass fragmenta-

TABLE 1:  $^{13}C$  NMR data and HMQC results for nevadensin

C-atom	$\delta_C$ -value	HMQC
2	164.2	-
3	104.2	$\delta_{H-3}$ 6.585
4	183.4	-
5	148.8	-
6	131.5	-
7	149.2	-
8	128.5	-
9	146.2	-
10	105.0	-
1'	124.0	-
2 <sup>1,6</sup> 1	127.8	$\delta_{H-2^{1,6}}$ 7.89
3 <sup>1,5</sup> 1	115.0	$\delta_{H-3^{1,5}}$ 7.045
4 <sup>1</sup>	163.1	-
8-OCH <sub>3</sub>	61.4	$\delta_H$ 4.02
6-OCH <sub>3</sub>	62.3	$\delta_H$ 4.04
4 <sup>1</sup> -OCH <sub>3</sub>	56.0	$\delta_H$ 3.90

tion pattern<sup>[24,25]</sup> as well as the negative response of the compound towards the colour reaction with *o*-dinitrobenzene<sup>[11]</sup>. A negative gossypetone reaction<sup>[26]</sup> suggested the presence of a methoxyl group at C-8 as well, and hence, the remaining hydroxyl group must be located at C-7 position, although there is no characteristic bathochromic shift of Band II in the UV spectrum upon addition of sodium acetate/ethanol  $^{3/4}$  which is the peculiarity of nevadensin itself<sup>[11]</sup>. This structural formulation has been confirmed by  $^{13}C$  NMR and HMQC spectral studies (TABLE 1). The HMQC spectrum exhibited expected heteronuclear cross-peaks, for eight protonated carbons, showing one-bond heteronuclear correlations ( $^1H - ^{13}C$ ) between C<sub>2</sub> and C<sub>6</sub>' protons at d7.89 (2H, dd,  $J= 2.7$  Hz, 11.7 Hz) with 2' and 6' carbons at d127.8; C<sub>3</sub> and C<sub>5</sub>' protons at d7.045 (2H, dd,  $J= 2.7$  Hz, 11.7 Hz) with 3' and 5' carbons at d115.0; C<sub>3</sub>-proton at d 6.585 (1H,s) with C<sub>3</sub>-carbon at d104.2, and the methoxy protons at d3.90, 4.02 and 4.04 of three methoxyl functions respectively with the methoxyl carbons at d56.0 (C<sub>4</sub>'-OCH<sub>3</sub>), 61.4 (C<sub>8</sub>'-OCH<sub>3</sub>) and 62.3 (C<sub>6</sub>'-OCH<sub>3</sub>).

The flavonoid **(2)**,  $C_{18}H_{16}O_6$  ( $[M]^+$  (EIMS)  $m/z$  328) exhibited UV, IR, NMR and mass spectral properties, which are in excellent conformity with

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the reported values for salvigenin (5-hydroxy-6,7,4'-trimethoxyflavone)<sup>[27,28]</sup>.

### EXPERIMENTAL

Melting points are uncorrected. TMS has been used as int. standard in recording NMR spectra. Whole plants of *Limmophila heterophylla* and *L. rugosa* were collected from Santiniketan and their identities were verified by Dr. H R Chowdhury and Dr. S Mondal (Visva-Bharati University). Voucher specimens have been deposited in the Natural Product Laboratory of this University.

#### Extraction and isolation

**Nevadensin (1):** Air-dried defatted powdered whole plants (1.5 kg) of *L. heterophylla* were extracted with petrol (60-80<sup>o</sup>) in a Soxhlet apparatus for 56 hr. The extract was concentrated under reduced pressure and then subjected to column chromatography on silica gel (60-120 mesh, 200 g); the petrol (60-80<sup>o</sup>C): benzene (1:2) eluent afforded nevadensin (5,7-dihydroxy-6,8,4'-trimethoxyflavone) as golden yellow needles (yield 0.8 g), m.p. 184-186<sup>o</sup>C (from ethanol); UV (ethanol):  $\lambda_{\max}$  280, 335 nm; (+AlCl<sub>3</sub>): 280, 310 (sh), 355 nm; IR (KBr)  $\nu$  cm<sup>-1</sup>: 3407, 3100, 2936, 2840, 1663, 1591, 1508, 1060, 1025; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300MHz; TMS):  $\delta$  12.78 (1H, s, C<sub>5</sub>-OH), 7.89 (2H, dd, *J*= 2.7 Hz, 11.7 Hz, H-2' and H-6'), 7.045 (2H, dd, *J*= 3 Hz, 11.7 Hz, H-3' and H-5'), 6.585 (1H, s, C<sub>3</sub>-H), 4.04 (3H, s, C<sub>6</sub>-OCH<sub>3</sub>), 4.02 (3H, s, C<sub>8</sub>-OCH<sub>3</sub>) and 3.90 (3H, s, C<sub>4</sub>-OCH<sub>3</sub>); EIMS (70 ev): *m/z* 344 ([M]<sup>+</sup>), 329 (base peak), 316[M-CO]<sup>+</sup>, 315 [M-CO-H]<sup>+</sup>, 314 [M-2Me]<sup>+</sup>, 312[M-2Me-2H]<sup>+</sup>, 301[M-CO-Me]<sup>+</sup>, 212 and 132 (retro-Diels-Alder fragmented ion peaks of 1), 197 and 132 (retro-Diels-Alder cleavage of mass fragment 329), 169[197-CO]<sup>+</sup>, 168 [169-H]<sup>+</sup>, 153[169-Me]<sup>+</sup>, 141[169-CO]<sup>+</sup>, 135(fragmented ion peak), 126[141-Me]<sup>+</sup>; <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) and HMQC in TABLE 1.

**Salvigenin (2):** Air-dried defatted powdered whole plants (1.5 kg) of *L. rugosa* were extracted with petrol (60-80<sup>o</sup>) in a Soxhlet apparatus for 56 hr. The extract was concentrated under reduced pressure and then subjected to column chromatography on silica gel

(60-120 mesh, 200 g); the petrol (60-80<sup>o</sup>C): benzene (1:3) eluent afforded salvigenin (5-hydroxy-6,7,4'-trimethoxyflavone) as yellow cubes (yield 0.65 g), m.p. 184-88<sup>o</sup>C (from benzene). UV (ethanol):  $\lambda_{\max}$  277, 330 nm; IR (KBr)  $\nu$  cm<sup>-1</sup>: 3420, 3080, 3015, 2920, 2845, 1650, 1600, 1590, 1570, 1360, 1265, 1120; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300MHz; TMS):  $\delta$  7.84 (2H, d, *J*= 8.9 Hz, H-2' and H-6'), 7.01 (2H, d, *J*= 8.9 Hz, H-3' and H-5'), 6.58 (1H, s, C<sub>3</sub>-H), 6.54 (1H, s, C<sub>8</sub>-H), 4.005 (3H, s, C<sub>6</sub>-OCH<sub>3</sub>), 3.976 (3H, s, C<sub>7</sub>-OCH<sub>3</sub>) and 3.925 (3H, s, C<sub>4</sub>-OCH<sub>3</sub>); EIMS (70 ev): *m/z* 328 ([M]<sup>+</sup>, base peak), 327 [M-H]<sup>+</sup>, 313[M-Me]<sup>+</sup>, 312 [M-Me-H]<sup>+</sup>, 300[M-CO]<sup>+</sup>, 299[M-CO-H]<sup>+</sup>, 196 and 132 (retro-Diels-Alder fragmented ion peaks of 2), 168 [196-CO]<sup>+</sup>, 167[168-H]<sup>+</sup>, 153[168-Me]<sup>+</sup>, 135 (fragmented ion peak), 120[135-Me].

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