Volume 10 Issue 4

NPAIJ, 10(4), 2014 [99-101]



*In-vitro* evaluation of the anti-diabetic activity of alcoholic extracts of certain plants belonging to families meliaceae and fabaceae

Mohamed I.S.Abdelhady<sup>1,2\*</sup>, Mahmoud Youns<sup>3,4</sup>

<sup>1</sup>Department of pharmacognosy, Faculty of pharmacy, Umm Al-Qura university, Makkah 21955, (SAUDI ARABIA)
<sup>2</sup>Pharmacognosy department, Faculty of pharmacy, University of helwan, Ain Helwan, Cairo, (EGYPT)
<sup>3</sup>Department of biochemistry and molecular biology, Faculty of pharmacy, University of helwan, Cairo, (EGYPT)
<sup>4</sup>Division of functional genome analysis, German cancer research center (DKFZ), Heidelberg, (GERMANY)
E-mail: mohibrahem@yahoo.com; miabdelhady@uqu.edu.sa

# ABSTRACT

The leaves of 4 plants belong to families *Meliaceae* and *Fabaceae* were exhaustively extracted with hot 80% Ethanol, under reflux. The dry residues of the alcoholic extracts were tested using Sucrase enzyme inhibitory activity test to evaluate their anti-diabetic activity. The alcoholic extracts exhibited significant invitro anti-diabetic activity.

© 2014 Trade Science Inc. - INDIA

#### INTRODUCTION

Herbal medicines have been used since centuries by different cultures worldwide for treatment of diabetes. Diabetes mellitus is a group of metabolic diseases characterized by hyperglycemia resulting from defects in insulin action, insulin secretion, or both<sup>[1]</sup>. Control of postprandial hyperglycemia is critical in the early treatment of diabetes mellitus<sup>[2]</sup> as it could induce non enzymatic glycosylation of various proteins, resulting in the development of chronic complications such as, micro- and macro-vascular diseases<sup>[3]</sup>, and it has also been proposed as an independent risk factor for cardiovascular diseases<sup>[4,5]</sup>. Postprandial hyperglycemia can be controlled by decreasing the absorption of glucose through the inhibition of enzymes responsible for hydrolysis carbohydrate such as sucrase enzyme, in the digestive tract<sup>[6]</sup>. Phytochemicals exhibit their hypoglycemic effect by several mechanisms, such as, inhibition of carbohydrate metabolizing enzymes, manipulation of glucose transporters,  $\alpha$  -cell regeneration, and enhancing the insulin releasing activity<sup>[7]</sup>. The present study deals with the investigation of the in-vitro anti-diabetic activity of several 4 plants belong to families *Meliaceae* (such as *Azadirachta indica*) and *Fabaceae* (such as *Acacia glauca, Acacia Senegal and Acacia nilotica*) through the performance of Sucrase inhibitory activity test.

#### **MATERIALS AND METHODS**

### **Preparation of the extracts**

Powdered, air-dried leaves of *Azadirachta indica*, *Acacia glauca*, *Acacia Senegal and Acacia nilotica* (250 g) were exhaustively extracted with hot 80% Ethanol (4x1 L), under reflux. After filtration the extracts were concentrated under vacuum at 50°C. The dry residues obtained were kept in the refrigerator.

### Assay of sucrase inhibitory activity

A crude enzyme solution of rat intestinal sucrase

# KEYWORDS

Meliaceae; Fabaceae; Anti-diabetic activity.

# Full Paper

enzyme was prepared according to the method of Dahlqvist<sup>[8]</sup>. The effect of samples on sucrase enzyme activity was assayed according to the method of Honda and Hara<sup>[9]</sup>. Enzyme solutions (10  $\mu$ L) were incubated together for 10 min at 37°C, and the volume was made up to 200  $\mu$ L with maleate buffer (pH 6.0) in case of control or up to 200  $\mu$ L with buffer solubilized sample (100  $\mu$ g/mL in maleate buffer with pH 6.0). The enzyme reaction was initiated by adding 100  $\mu$ L of sucrose solution (60 mM). After 30 min, the reaction was terminated by adding 200  $\mu$ L of 3,5-dinitrosalysilic acid reagent and placing the mixture in a boiling water bath for five min. The absorbance of solution was read at 540 nm. The percent inhibitory activities were calculated using the following formula:

% Inhibition = 
$$\frac{\text{Abs control -Abs sample}}{\text{Abs control}} \times 100$$

Where, Abs control is the absorbance of the control reaction (containing all reagents except the test sample), and the Abs sample is the absorbance of the test sample; an untreated enzyme solution was used as the control. All experiments were carried out in triplicate.

## RESULTS

# **Results of the anti-diabetic activity (Sucrase inhibitory activity)**

Acacia nilotica decreases the activity of sucrase enzyme by  $(61.38 \pm 2.46)$  followed by Acacia glauca  $(59.43 \pm 3.42)$ , Acacia Senegal  $(47.54 \pm 1.33)$  and Azadirachta indica (41.34  $\pm$  2.37), as shown in TABLE 1.

TABLE 1: Sucrase enzyme activity

Tested extract	Sucrase I%±SD
Azadirachta indica	$41.34 \pm 2.37$
Acacia glauca	$59.43 \pm 3.42$
Acacia Senegal	$47.54 \pm 1.33$
Acacia nilotica	$61.38\pm2.46$

SD is the standard deviation

#### DISCUSSION

It is a general opinion that medicinal plants inhibit sucrase activity due to the presence of several possible factors and mechanisms, such as polyphenolic concentration<sup>[14]</sup>. In our present investigation, the effect of various extracts of tested plants on carbohydrate hydrolyzing enzyme, namely, rat intestinal sucrase, have been studied using in vitro model systems. The extracts of tested plants significantly inhibited sucrase enzyme activity (Figure 1). The Sucrase inhibitory activity of tested plants supposed to be due to the presence of flavonoid glycosides and/or hydrolysable tannins<sup>[10]</sup>. With a constant rise in the incidence of type II diabetes around the world it appears that more anti-diabetic drugs with complementary mechanisms of action should be developed, in order to achieve durable glycemic control by inhibiting, in a reversible way, the hydrolysis of disaccharides and the ultimate steps of the digestion of dietary polysaccharides, to reduce the rise of postpran-



101

dial blood glucose in diabetics<sup>[11]</sup>. Rat intestinal sucrase occurs as a complex of sucrase and isomaltase, which converts sucrose into glucose<sup>[12]</sup>. So the tested plants (i.e. *Azadirachta indica*, *Acacia nilotica*, *Acacia glauca* and *Acacia Senegal*) may offer a support to control carbohydrate hydrolysis in diabetic disease.

## CONCLUSION

The ethanol extract of the leaves of *Azadirachta indica* (family *Meliaceae*), *Acacia nilotica*, *Acacia glauca* and *Acacia Senegal* (family *Fabaceae*) leaves exhibited significant in-vitro anti-diabetic activity using Sucrase inhibitory activity test. But previous studies observed that those plants which belonging to the family *Fabaceae* (Mimosaceae) contain the toxic alkaloid mimosine<sup>[13,14]</sup>, so we recommend in vitro and in-vivo toxic test to be done to evaluate their safety to be used as complementary drugs to help main medicines in treatments of diabetic patients.

# AKNOWLEDGMENT

The authors would like to thank Institute of Scientific Research and Revival of Islamic Heritage at Umm Al-Qura University (Project ID: 4331014) for the financial support.

### REFERENCES

- [1] C.C.Teixeira, C.A.Rava, P.M.Da Silva, R.Melchior, R.Argenta, F.Anselmi et al.; Absence of antihyperglycemic effect of jambolana in experimental and clinical models. J.Ethnopharmacol, 71, 343-7 (2000).
- [2] Y.J.Shim, H.K.Doo, S.Y.Ahn, Y.S.Kim, J.K.Seong, I.S.Park et al.; Inhibitory effect of aqueous extract from the gall of Rhus chinensis on alpha-glucosidase activity and postprandial blood glucose. J.Etnhopharmacol, 85, 283-7 (2003).

 [3] A.D.Baron; Postprandial hyperglycemia and α-glucosidase inhibitors. Diabetes Res.Clin.Pract., 40, 51-5 (1998).

- J.F.Blickle, E.Andres, J.M.Brogard; Current status of the treatment of type 2 diabetes mellitus: Alphaglucosidase inhibitors. Food Chem., 106, 247-252 (2008).
- [5] A.Ceriello; The emerging role of post-prandial hyperglycaemic spikes in the pathogenesis of diabetic complications.Diabetic Med., 15, 188-193 (1998).
- [6] R.Rhabasa-Lhoret, J.L.Chiasson; International textbook of diabetes mellitus. 3 rd Edition, UK: John Wiley and Sons Ltd., **1**, 901-914 (**2004**).
- [7] A.K.Tiwari, J.M.Rao; Diabetes mellitus and multiple therapeutic approaches of phytochemicals: Present status and future prospects. Curr.Sci., 23, 30-3 (2002).
- [8] A.Dahlqvist; Method for assay of intestinal disaccharides. Anal Biochem 7, 18-25 (1964).
- [9] M.Honda, Y.Hara; Inhibition of rat small intestinal sucrase and α-glucosidase activities by tea polyphenols. Biosci.Biotechnol.Biochem., 57, 123-124 (1993).
- [10] A.Andrade-Cetto, M.Heinnirch; Mexican plants with hypoglycemic effect used in treatment of diabetes. J.Ethnopharmacol, 99, 325-348 (2005).
- [11] Gülçin; Comparison of in vitro antioxidant and antiradical activities of L-tyrosine and L-Dopa Amino Acids., **32**, 431-438 (2007).
- [12] C.Röcken, S.Carl-McGrath; Pathology and pathogenesis of hepatocellular carcinoma.Dig.Dis, 19, 269-278 (2001).
- [13] Muchdar Soedarjo, K.Thomas, Dulal Borthakuri; Mimosine, a Toxin Present in Leguminous Trees (Leucaena spp.), Induces a Mimosine-Degrading Enzyme Activity in Some Rhizobi Strains. Applied and Environemental Microbiology, 4268-4272 (1994).
- [14] A.C.Hammond; *Leucaena toxicosis* and its control in ruminants. Journal of animal Sciences, 73, 1487-1492 (1995).

