ISSN : 0974 - 7435

Volume 9 Issue 5



FULL PAPER BTAIJ, 9(5), 2014 [206-209]

Investigations on gold nanoparticles biosynthesis potential of marine derived fungi

A.K.Vala*

Department of Physics, Maharaja Krishnakumarsinhji Bhavnagar University, Sardar Vallabhbhai Patel Campus, Bhavnagar-364 001, (INDIA) E-mail : anjana_vala@yahoo.co.in

ABSTRACT

The objective of the work was to explore potential of marine-derived fungi for biosynthesis of gold nanoparticles (GNPs). Three marine-derived fungal isolates viz. *Aspergillus candidus, Aspergillus flavus and Aspergillus niger* have been examined for their ability to biosynthesize GNPs. All the test isolates were challenged with different concentrations (0.25, 0.5, 1, 2 and 3mM) of gold (III) chloride under static condition at 27°C for 72h. The isolates could biosynthesize spherical GNPs mostly extracellularly. In certain cases, mode of synthesis extra/intracellular was observed to be dependent on gold ion concentrations. While there is no much information available on exploitation of marine-derived fungi for biosynthesis of metal nanoparticles, present findings suggest marinederived fungi as potential resource for developing cost-effective green technology for GNP synthesis. © 2014 Trade Science Inc. - INDIA

INTRODUCTION

Gold nanoparticles, having potential applications in the areas like catalysis, diagnosis, drug delivery, imaging, photonics and targeted killing of cells^[1-5] are in great demand. While physico-chemical synthesis protocols are expensive, polluting nature and even have restricted use in clinical and pharma applications, biosynthesis of gold nanoparticles offers a promising option to meet the requirement.

A number of microorganisms have been examined for nanoparticle biosynthesis. While researches on nanobiotechnology in future may increasingly depend on marine microbes having ability to grow under ex-

KEYWORDS

Biosynthesis; Extracellular; Gold nanoparticles; Green technology; Marine-derived fungi.

treme conditions, it has been suggested that initiatives should be taken for utilization of these resources in the area of nanobiotechnology^[6] However, marine microbes are comparatively less explored for biosynthesis of gold nanoparticles and only few reports exist^[7,8]. Harnessing fungi for such purposes is more advantageous^[7].

The present study is carried out with a view to examine gold nanoparticles biosynthesis potential of three marine –derived aspergilli. The data showed that all the test fungi could biosynthesize gold nanoparticles mostly extracellularly. In certain cases, mode of synthesis extra/intracellular was observed to be dependent on gold ion concentrations.

207

MATERIALS AND METHODS

Isolation and identification of test fungus

The test fungi were isolated from waters of Bhavnagar coast (Lat. 21° 45' N and Long. 72°. 14' E), Gulf of Khambhat, West Coast of India. The isolates were grown and maintained on potato dextrose agar (PDA) medium^[9] and stored at 4 °C until use. The medium was prepared in aged seawater and distilled water at a ratio of 3:1^[10]. Identification of the test strains was carried out by its macroscopic and microscopic characteristics and confirmed identification as *Aspergillus candidus*, *Aspergillus flavus and Aspergillus niger* was carried out by the Agharkar Research Institute, Pune, India.

Biosynthesis and characterization of gold nanoparticles

One ml inoculum (spore suspension approximately 10⁶/ml) was inoculated in 250 ml Potato Dextrose medium (prepared in 75% 'aged' seawater). The inoculated flasks were incubated at room temperature for 4 days. After incubation period, fungal biomass was separated from the medium by filtration and washed extensively with sterile distilled water.

Approximately 5g fungal biomass of each test isolate was challenged with gold chloride of different concentrations, 0.25mM, 0.5mM, 1mM, 2mM and 3mM. The gold chloride solutions were prepared in sterilized deionized water. The inoculated flasks were incubated at 27°C for 72h under static condition. Negative and positive controls were also run along with the experimental ûasks.

Two ml sample from each flask was withdrawn at predetermined time intervals and the spectra were recorded in the range of 380 to 800nm at a resolution of 1 nm using UV–visible spectrophotometer (Elico BL-198). The experiments were carried out in triplicates^[7].

Transmission electron microscopic (TEM) images of biosynthesized gold nanoparticles (mounted on carbon-coated copper grids) were obtained on JEOL (Model GEM 200).

RESULTS

All the test flasks containing biomass of Aspergillus candidus and gold chloride solution showed gradual colour change (pink-purple) in the reaction mixture within 18h, except the test flask containing 1mM gold chloride. In the flask containing 1mM gold chloride, biomass exhibited purple-violet coloration. Figure 1 shows spectrophotometric data of the biosynthesized GNPs. Figure 2 (a) shows TEM image of the GNPs biosynthesized by *A. candidus*.

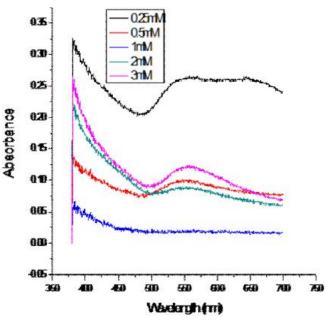


Figure 1 : Uv spectra of GNPs biosynthesized by *Aspergillus* candidus at different gold chloride concentrations

The test fungus *A*. *flavus* could also synthesize GNPs, here also, the biosynthesis was extracellular within 24h (data not shown).

In case of *A. niger*, gradual change in coloration of the reaction mixure was observed within 18h. Uv spectra of biosynthesized GNPs by *A. niger* are shown in Figure 3 while Figure 2(b) shows TEM micrograph of the GNPs. The test fungi produced GNPs in the size range of 9-37nm.

DISCUSSION

Coloration of the reaction mixture indicates extracellular biosynthesis of GNPs. When exposed to 1mM gold chloride, coloration of *A. candidus* biomass was observed indicating intracellular mode of GNP biosynthesis. Hence, all the three test fungi in general exhibited extracellular biosynthesis and *A. candidus* showed a different behaviour (intracellular biosynthesis) in certain case. Concentration dependent change in mode of

BioTechnology An Indian Journal

Full Paper C

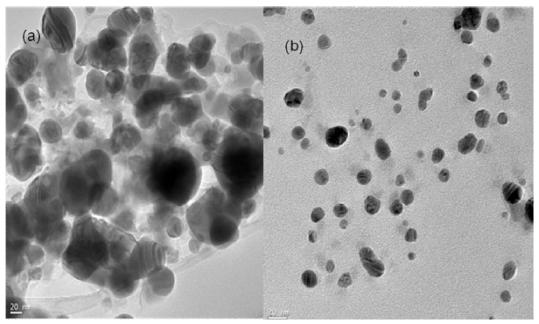


Figure 2 : TEM images of GNPs biosynthesized by (a) Aspergillus candidus and (b) Aspergillus niger

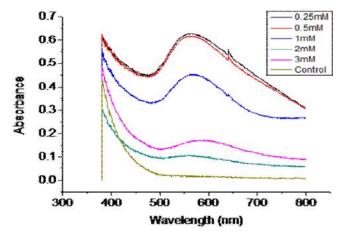


Figure 3 : Uv spectra of GNPs biosynthesized by A. niger at different gold chloride concentrations

biosynthesis has been observed by Vala^[7]. Ahmad et al.^[11] reported intra- and extracellular synthesis of gold nanoparticles by fungus Trichothecium. Sheikhlou et al ^[12] observed simultaneous intra-extracellular biosynthesis of gold nanoparticles by *Rhizopus oryzae*. UV– Vis spectrophotometry is one of the most extensively used techniques for characterization of gold nanoparticles, The observed surface plasmon resonance band (SPR) in Figures 1 and 3 is characteristic of GNPs, confirming the biosynthesis in all cases. Transmission electron microscopy offers a powerful tool for determination of shape and size of nanoparticles^[13]. The TEM data (Figure 2(a,b)) revealed spherical shape of the particles. Spherical GNP production by marine-derived

Rhizopus oryzae has been reported by Vala^[7]. Shivshankar et al.^[14] used geranium leaves (*Pelargo-nium graveolens*) and its endophytic fungus *Colletotrichum* sp. and reported that gold nanoparticles synthesized using the fungus were essentially spherical in shape while the particles grown using the leaves exhibited a variety of shapes.

The present study demonstrates the marine-derived fungi as potential resource for developing cost-effective green technology for biosynthesis of gold nanoparticles.

ACKNOWLEDGEMENT

Council of Scientific and Industrial Research (CSIR), New Delhi, is gratefully acknowledged for financial support under Scientists' Pool scheme (Pool No.8426-A).

REFERENCES

- M.Han, X.Gao, J.Z.Su, S.Nie; Quantum-dot-tagged microbeads for multiplexed optical coding of biomolecules.Nat.Biotechnol., 19, 631-635 (2001).
- [2] X.Huang, I.El-Sayed, W.Qian, M.A.El-Sayed; Cancer cell imaging and photothermal therapy in the near-infrared region by using gold nanorods.J.Am.Chem.Soc., 128, 2115-2120

FULL PAPER

209

(2006).

- [3] C.A.Mirkin, R.L.Letsinger, R.C.Mucic, J.J.Storhoff; A DNA-based method for rationally assembling nanoparticles into macroscopic materials.Nature, 382, 607-609 (1996).
- [4] A.K.Salem, P.C.Searson, K.W.Leong; Multifunctional nanorods for gene delivery.Nat.Mater, 2, 668-671 (2003).
- [5] K.Zhang, L.Hao, S.J.Hurst, C.A.Mirkin; Antibodylinked spherical nucleic acids for cellular targeting, J.Am.Chem.Soc., 134, 16488-16491 (2012).
- [6] D.Chandramohan; Marine Microbiology: Challenges and Future Directions.In: Marine Microbiology: Facets & Opportunities, N.Ramaiah (Ed.), National Institute of Oceanography, Goa, 7 (2004).
- [7] A.K.Vala; Intra and extracellular biosynthesis of gold nanoparticles by a marine-derived fungus Rhizopus oryzae.Synth.React.Inorg.Metal-Org.Nano-Metal Chem.(Accepted for publication), (2013).
- [8] N.Sharma; A.K.Pinnaka, M.Raje, F.N.U.Ashish, M.S.Bhattacharyya, A.Roy Choudhury; Exploitation of marine bacteria for production of gold nanoparticles.Microbial Cell Factor, 11, 86 (2012).

- [9] Anonymous; Plant Pathologist's Pocketbook.Commonwealth Mycological Society, KEW, Surrey, England, (1968).
- [10] C.Schlieper, (Ed.); Research Methods in Marine Biology.Sidgwick and Jackson Ltd, London, (1972).
- [11] A.Ahmad, S.Senapati, M.I.Khan, R.Kumar, M.Sastry; Extra-/intracellular biosynthesis of gold nanoparticles by an alkalotolerant fungus, Trichothecium sp.J.Biomed.Nanotechnol, 1, 47–53 (2005).
- [12] Z.Sheikhlou, M.Salouti, Z.Farahmandkia, S.Mahmazi, A.Einlou, Intra-extracellular biosynthesis of gold nanoparticles by fungus Rhizopus Oryza.ZUMS.J., 20(78), 47-56 (2012).
- [13] T.M.Mayhew, C.Muhlfeld, D.Vanhecke, M.Ochs; A review of recent methods for efficiently quantifying immunogold and other nanoparticles using TEM sections through cells, tissues and organs.Ann.Anat.Anatomischer Anzeiger, 191(2), 153–170 (2009).
- [14] S.Shivshankar, A.Ahmad, R.Pasricha, M.Sastry; Bioreduction of chloroaurate ions by Geranium leaves and its endophytic fungus yields gold nanoparticles of different shapes.J.Mater.Chem., 13, 1822–1826 (2003).

BioTechnology An Indian Journal