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Development of a Validated Stability Indicating RP-HPLC Method for Simultaneous Determination of Benazepril, Benazeprilat, and Amlodipine in Presence of Their Degradation Products and Application to Pharmaceutical Formulation and Human Plasma

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Abstract

A sensitive, reproducible, and rapid stability indicating RP-HPLC method was developed and subsequently validated for simultaneous determination of both benazepril (BENZ), benazeprilat (BENZT), and amlodipine (AML) in presence of their basic hydrolysis degradation products in bulk powder, pharmaceutical formulation, and application to human plasma using moexipril (MOX) as an internal standard (IS). The method uses Inertsil C18 column (250 mm \times 4.6 mm, 5 μ) and the mobile phase used was consisting of acetonitrile-potassium hydrogen phosphate buffer pH 7 adjusted by 0.1 N NaOH (40/60 v/v) with a flow rate of 1.0 mL/min and detection at 242 nm. A detailed validation of the methods was performed following the ICH guidelines and the standard curves were found to be linear in the range of 0.5 μ g/mL to 100 μ g/mL, 0.5 μ g/mL to 100 μ g/mL and 5 μ g/mL to 100 μ g/mL for BENZ, BENZT, and AML, respectively. Statistical comparison was done between the proposed method and the reported one where no significant difference was found between the two methods.

Keywords: Benazepril; Benazeprilat; Amlodipine; Stability indicating; RP-HPLC

Introduction

Benazepril, (3S)-3-[(1S)-1-ethoxycarbonyl-3 phenylpropylamino]-2,3,4,5-tetrahydro-2-oxo-1H-1-benzazepin-1-yl] acetic acid hydrochloride is an antihypertensive drug, which belongs to the group of angiotensin convertase inhibitors. It acts on the reninangiotensin-aldosterone system by inhibition of the conversion of the inactive angiotensin I to the highly potent vasoconstrictor angiotensin II. It also reduces the degradation of bradykinin. Benazepril HCl is applied in pharmacotherapy as a first choice drug for treatment of arterial hypertension, ischemic heart disease, hypertrophy of the left heart ventricle and post infarction heart dysfunction [1]. *In vivo*, benazepril is hydrolyzed to a pharmacologically active metabolite, the diacid benazeprilat, (3-[(1-

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carbonyl-3-phenyl-(1S)-propyl)-amino]-2,3,4,-5-tetrahydro-2-oxo-1-(3S)-benazepine-1-acetic acid). Amlodipine is a potent dihydropyridine calcium antagonist [2,3] useful in the management of angina pectoris and hypertension [4].

It is reported that benazepril was identified by HPLC [5-8], spectrophotometry [9-15], and with benazeprilat by liquid chromatography-mass spectrometry [16,17], and gas chromatography-mass spectrometry [18]. Also it is reported that amlodipine was determined by HPLC [19,20] spectrophotometry [21,22]. Benazepril with Amlodipine also were determined by HPLC [23], spectrophotometry [24], and liquid chromatography-mass spectrometry [25]. Loadless®, a drug used in the Egyptian market, which is a combination of BENZ and AML is used for the treatment of hypertension. Literature survey revealed that BENZ is approved in United States Pharmacopeia [26], while AML is approved in British Pharmacopeia [27]. Only one of the reported methods [6] can be comparable to the present one, but the run time was about 20 min. and the sensitivity was intermediate, also there is no quantitation of the active metabolite BENZT. Our goal in this manuscript is to shorten the run time and increase the method sensitivity to be applicable to determination of BENZ, BENZT, and AML in biological fluids as human plasma. Moreover, there is no structure elucidation of the degradation products in the reported method [6], where our present study introduces structure elucidation of the basic degradation products.

Experimental

Material and reagents

BENZ, AML, and MOX (IS) were kindly supplied from National Organization for Drug Control and Research (NODCAR, Cairo, Egypt). The purity of the standards was certified to be higher than 99%. Structures of the compounds are shown in FIG. 1.

Loadless® commercial capsules, labeled to contain 5 mg AML and 20 mg BENZ batch number 520034, 520047, 520080 was obtained from local market.

- Methanol: HPLC-grade (Sigma-Aldrich).
- Acetonitrile: HPLC-grade (Sigma-Aldrich).
- Double distilled water was used.
- Other reagents were of analytical-grade.

FIG. 1. Chemical structures of a) benazepril, b) benazeprilat, and c) amlodipine.

Instrument

Analysis was performed on a chromatographic system Jasco LC-Net II/ADC (Japan) equipped with UV detector (UV-2070 plus), isocratic pump (PU-2080 plus) and 4-line degasser (DG-2080-54). A chromatographic separation was achieved by Inertsil C-18, 250 mm \times 4.6 mm, 5 μ analytical column. Data acquisition were made with Chrom NAV software.

Mass spectrometer for structure elucidation of the degradation product

This is done by MS/MS detection in positive ion mode using a MDS Sciex (Foster City, CA, USA) API-3200 MS/MS triple quadrupole mass spectrometer, equipped with a Turbo ion spray interface at 350°C. The common parameters, nebulizer gas (GS1), heater gas (GS2) and collision activated dissociation gas (CAD), were set at 30, 40, and 5 psi, respectively. The compound parameters, declustering potential (DP), collision energy (CE) and collision exit potential (CXP) were set at 56, 31, and 12 V respectively. The analytical data were processed using Analyst software (version 1.4.2).

Procedure

Preparation of the basic degradation products of the studied drugs

BENZ and AML were subjected to basic degradation. 100 mg of each powder was weighed, transferred separately to two conical flasks. Then, 0.1 N NaOH was added separately on the two volumetric flasks. Then the flasks were refluxed for 30 min for basic degradation. After the reflux, the conical flasks were neutralized against 0.1 N HCl, and then completed to 100 mL by distilled water. The complete degradation was confirmed using HPLC and mass spectrometry.

Preparation of the acidic degradation product of BENZ

BENZ was weighed for 100 mg and transferred to a conical flask. Then, 0.1 M HCl was added, and the flask was refluxed at 90°C for 3 h. After the reflux, the content was neutralized against 0.1 M NaOH, transferred quantitatively to 100 mL volumetric flask, and then completed to 100 mL by distilled water.

Chromatographic conditions

Separation of the two analytes along with the degradation products and the IS was accomplished using Inertsil C18 column (250 mm \times 4.6 mm, 5 μ column) as a stationary phase. The mobile phase used was acetonitrile-potassium hydrogen phosphate buffer pH=7 adjusted by 0.1 N NaOH (40/60 v/v) was used as a mobile phase. The flow rate was 1 mL/min and the detection wavelength was 242 nm.

Preparation of standard stock and working solutions

Primary stock solutions of BENZ, AML, and MOX internal standard (IS) (all at 1.0 mg/mL) were separately prepared by dissolving 100 mg of each standard powder in the least amount of methanol and completed to the volume by distilled water, where BENZT was prepared by degradation (as explained earlier). Primary stock solutions were diluted with the mobile phase for the RP-HPLC to prepare standard working solutions of BENZ, BENZT, AML, and MOX (100 µg/mL). All solutions were stored at 4°C, and equilibrated to room temperature before use.

Linearity and calibration standards of the pure bulk powder

Accurately measured aliquots of BENZ, BENZT, and AML were transferred from their working standard solution (0.1 mg/mL) into three series of 10 ml volumetric flasks and complete to volume with the mobile phase (acetonitrile-potassium hydrogen phosphate buffer (40/60 v/v) pH=7 adjusted by 0.1 N NaOH). The calibration samples consist of six concentrations of BENZ (0.5 μ g/mL to 100 μ g/mL), BENZT (0.5 μ g/mL to 100 μ g/mL), and AML (5 μ g/mL to 100 μ g/mL). The samples were injected separately along with MOX (IS) into the Inertsil C18 column under a flow rate of 1 mL/min. The relative peak area of each analyte was recorded against its concentration, the linearity curves were constructed and the regression equations computed.

Validation

Accuracy: The accuracy of the results was checked by applying the proposed methods for determination of three replicates of different concentrations of the analytes. The concentrations were obtained from the corresponding regression equations, from which the percentage recoveries suggested good accuracy of the proposed methods.

Precision

Repeatability: Three concentrations of the analytes were analyzed three times intra-daily using the proposed methods under the same experimental conditions. The relative standard deviations were calculated.

Reproducibility (intermediate precision): The previous procedures were repeated inter-daily on three different days for the analysis of the three chosen concentrations. The relative standard deviations were calculated.

Laboratory prepared mixtures (selectivity): Solutions containing different ratios of the analytes were prepared by transferring accurately measured aliquots from their standard working solutions into a series of 10 ml volumetric flasks and the volume was completed to the mark with mobile phase. The final concentration ranges were 5 μg/mL to 20 μg/mL for BENZ, 10 μg/mL to 25 μg/mL for BENZT, and 5 μg/mL to 25 μg/mL for AML. The chromatograms of these different laboratory prepared mixtures were recorded and the procedure under linearity was then followed. Concentrations of the analytes in the prepared samples were calculated from the corresponding computed regression equations.

Application to pharmaceutical formulation

To determine the content of BENZ and AML in commercial capsules (Loadless®) (each capsule labeled to contain 20 mg BENZ and 5 mg AML), 20 capsules were weighed and finely powdered. A portion of powder equivalent to one capsule was weighed accurately and transferred to a 100 ml beaker. Methanol (50 ml) was added, stirred using a magnetic stirrer for 15 min and filtered through 0.5 µm Whatman filter paper into a 100 ml volumetric flask. The residue was washed three times each with 10 ml of methanol and the solution was completed to the mark with the same solvent. From the above prepared solution, further dilutions were prepared in the obtained linearity ranges using mobile phase. The general procedure described under linearity was followed to determine the concentration of both drugs in the prepared dosage form solution. The analysis was done in triplicates. Concentrations of BENZ and AML in the prepared samples were calculated from the corresponding computed regression equations.

Application to spiked human plasma sample

Aliquot volumes from the standard working solutions of BENZ, BENZT, and AML were added on 500 μ L human plasma, then 30 μ L IS was added. Then, 3 mL of acetonitrile was added, the samples were mixed on a vortex for 1 min, followed by centrifugation for 10 min at 10,000 rpm. The organic phase solution was transferred to a clean tube and evaporated to dryness under a gentle stream of nitrogen gas at 40°C. The residue was reconstituted with 100 μ L mobile phase, and 20 μ L was injected. Then, the same procedure was repeated but spiking was done after the extraction step to calculate the recovery percentage.

Results and Discussion

This manuscript describes the use of a RP-HPLC method to quantify BENZ, its active metabolite BENZT, and AML in the pharmaceutical dosage form. Also the paper describes the application of the proposed method to determine the three analytes in spiked human plasma without interference form endogenous plasma constituents. All the determinations were done in a

short run time and with high sensitivity. The novelty in our work is the analysis and quantitation of BENZ, AML, and the active degradation product of BENZ which is BENZT. BENZT is the active moiety that exerts the antihypertensive effect in the body, so its quantitation is very important. Also, full structure elucidation of the analytes and their degradation products was done using tandem mass spectrometry.

Method development

Different organic modifiers proportions along with different buffers with different pH were tried. Our goal is to obtain optimum resolution, symmetric peak shape, reasonable run time, and best sensitivity TABLE 1. Methanol and acetonitrile were tried, methanol caused longer run time where the run time extended to more than 20 min. Phosphate buffer with different pH's (3, 5, 7) were tried. The optimum resolution and peak shape obtained with acetonitrile-potassium hydrogen phosphate buffer pH=7 adjusted by 0.1 N NaOH (40/60 v/v) as a mobile phase. The flow rate for better resolution and rapid separation was adjusted to 1 mL/min. Also, two types of stationary phases C8 and C18 were investigated for the optimum resolution of the analytes' peaks, however the more hydrophobic Inertsil C18 was found to be more superior in separating analytes' peaks in a reasonable run time. All the tried conditions are summarized in TABLE 1. Full separation of the three peaks of BENZ, BENZT, and AML was obtained, where the retention time of the analytes was 6.3 min, 2.4 min, and 7.8 min, respectively, where the retention time for MOX (IS) was 5.3 min as shown in FIG. 2. System suitability parameters are shown in TABLE 2.

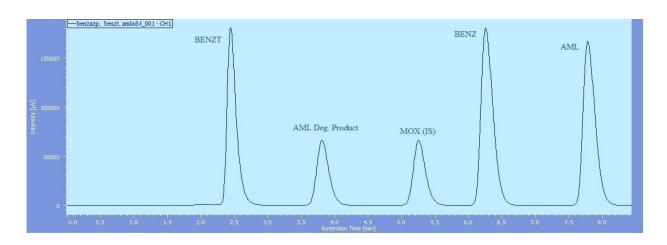


FIG. 2. HPLC chromatogram showing the complete resolution of BENZ, BENZT, and AML (80 μ g/mL each) with MOX (IS) and AML degradation product.

Stability of BENZ and AML

BENZ with a molecular weight (424.5) is an ester pro-drug which is hydrolyzed *in vivo* by esterase enzyme to the pharmacologically active biacid BENZT with a molecular weight (396). So to obtain this active form, hydrolysis of BENZ was tried by exposing to 0.1 N NaOH and refluxing for 30 min for basic degradation. Also complete degradation of AML was done by the same method. AML has a molecular weight of (408.8) where its basic degradation has a molecular weight (366). The complete degradation was confirmed by HPLC (with the disappearance of BENZ and AML peaks at retention times 6.3 min and 7.8 min, respectively. and appearance of the BENZT and basic degradation product of AML peaks at retention times 2.4 min and 3.8 min, respectively (FIG. 3a and 3b), and mass spectrometry (with the appearance of m/z for BENZT and degradation product of AML at 397 and 367.2, respectively, and disappearance of m/z for BENZ and AML at

425.2 and 409.2, respectively) as shown in FIG. 4a and 4b. The structures of the degradation products were suggested according to HPLC and MS data as shown in FIG. 5a and 5b.

TABLE 1. Different conditions tried for obtaining good separation and peak shape.

Organic modifier %	Result
50% Methanol	Long run time more than 15 min
40% Methanol	Long run time more than 20 min
50% Acetonitrile	Bad resolution
40% Acetonitrile	Good resolution
pН	Result
3	Bad resolution for degradation and Amlodipine peaks
5	Bad peak shape
7	Ideal resolution and peak shape

TABLE 2. System suitability parameters of the proposed RP-HPLC method.

Parameter	Benazepril	Benzeprilat	Amlodipine
Retention time	6.3	2.4	7.8
Resolution*	3.5	4.2	-
Tailing factor	1.2	1.2	1.3
Column Capacity	4.73	1.18	6.09
Column efficiency (No. of theoretical plates)	6879	1955	6348
НЕТР	0.004	0.013	0.004

^{*}Resolution is calculated relative to the next peak.

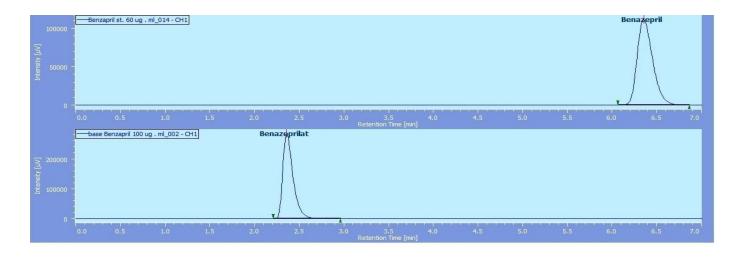


FIG. 3a. HPLC chromatograms of BENZ (60 μ g/mL) and BENZT (100 μ g/mL) confirming the complete degradation of BENZ by basic and acidic hydrolysis.

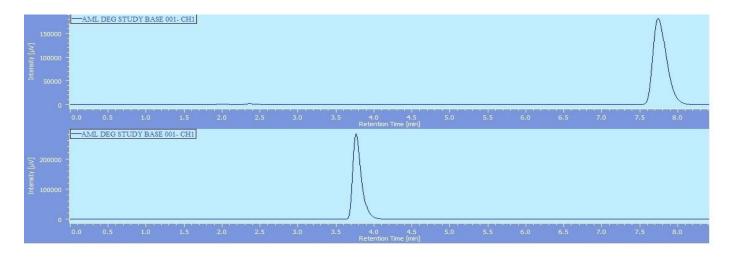


FIG. 3b. HPLC chromatograms of AML (80 μ g/mL) and AML basic degradation product, confirming the complete degradation of AML by basic hydrolysis.

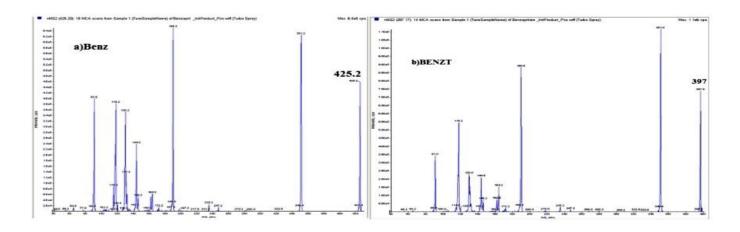


FIG. 4a. MS spectra of i) BENZ and ii) BENZT confirming the complete degradation by basic hydrolysis.

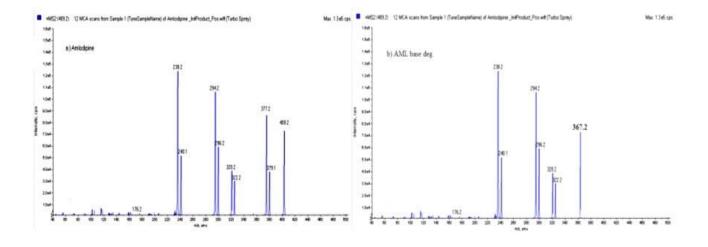


FIG. 4b. MS spectra of i) AML and ii) AML basic degradation product confirming the complete degradation by basic hydrolysis.

FIG. 5a. Suggested degradation pathway of BENZ by acidic and basic hydrolysis according to HPLC and MS data.

Amlodipine M. Wt =
$$408.8$$

OH-

 H_3C
 NH_2
 NA^+

Amlodipine M. Wt = 408.8

Amlodipine Alkaline Deg.

M. Wt = 366

FIG. 5b. Suggested degradation pathway of AML by basic hydrolysis according to HPLC and MS data.

Validation

Validation of the proposed methods was assessed according to ICH guidelines [28].

Linearity and Range: The linearity and range of the method were evaluated by analyzing six concentrations of BENZ, BENZT, and AML. Each concentration was repeated three times. The assay was performed according to the experimental conditions previously mentioned. The linear regression equations are summarized in TABLE 3.

Accuracy: The accuracy of the results was checked by applying the proposed methods for determination of different samples of BENZ, BENZT, and AML. The concentrations were obtained from the corresponding regression equations, from which the percentage recoveries suggested good accuracy of the proposed methods. Results are shown in TABLE 3.

Precision

Repeatability: Three concentrations of BENZ, BENZT, and AML were analyzed three times intra-daily using the proposed methods. The relative standard deviations were calculated TABLE 3.

Reproducibility (intermediate precision): The previous procedures were repeated inter-daily on three different days for the analysis of BENZ, BENZT, and AML. The relative standard deviations were calculated (TABLE 3).

Selectivity: Selectivity of the methods was achieved by the analysis of different laboratory prepared mixtures of BENZ, BENZT, and AML within the linearity range. Satisfactory results were obtained as shown in TABLE 4.

Application of the method in assay of capsules

The proposed method was applied for the determination of BENZ and AML in their combined pharmaceutical formulation (Loadless® capsules). The validity of the methods was assessed by applying the standard addition technique (TABLE 5). It shows that the developed methods are accurate and specific for determination of the cited drugs in presence of dosage form excipients.

Application to spiked human plasma sample

The chromatographic method was applied for the determination of BENZ, BENZT, and AML in biological fluids. Drug free human plasma was obtained from healthy volunteer, and then Spiked with BENZ, BENZT, and AML. The method showed no interference from endogenous plasma constituents as shown in FIG. 6. The recovery data are presented in TABLE 6.

TABLE 3. Validation parameters of the proposed chromatographic methods.

Parameter	Benazepril	Benazeprilat	Amlodipine	
Range µg/mL	0.5-100	0.5-100	5-100	
Regression Equation	y=0.0454x+0.0222	y=0.0423x+0.0142	y=0.0572x+0.0577	
Correlation coefficient (r)	0.9998	0.9998	0.9998	
Accuracya	99.54 ± 1.045	101.25 ± 0.854	100.73 ± 1.356	
Repeatability ^b	98.26 ± 1.211	100.25 ± 1.341	99.14 ± 1.234	
RSD%	1.232	1.338	1.245	
Intermediate precision ^c	99.12 ± 1.231	99.21 ± 1.421	101.21 ± 1.544	
RSD%	1.242	1.432	1.526	

^aThree concentrations of each analyte (10 μg/mL, 15 μg/mL and 25 μg/mL), repeated three times for each concentration b Intra-day (n=3), average of three concentrations of the analytes (10 μg/mL,15 μg/mL and 25 μg/mL) repeated 3 times within the same day; c Inter-day (n=3), average of three concentrations of the analytes (10 μg/mL, 15 μg/mL and 25 μg/mL) repeated 3 times in three consecutive days.

TABLE 4. Determination of the three analytes in their laboratory prepared mixtures by the proposed methods.

Concentration (µg/mL)			(Recovery % ± SD)		
Benazepril	Benazeprilat Amlodipine		Benazepril	Benazeprilat	Amlodipine
20	10	5	99.25 ± 0.251	99.47 ± 0.421	101.25 ± 0.585
15	15	10	99.87 ± 0.341	98.41 ± 0.1214	99.54 ± 1.471
10	20	15	98.65 ± 0.428	99.25 ± 0.751	99.15 ± 0.778
5	25	20	98.32 ± 0.497	99.41 ± 0.674	100.45 ± 0.541

All Calculations were done in triplicates.

TABLE 5. Determination of benazepril and amlodipine in Loadless® capsules by the proposed method and application of the standard addition technique.

Loadless®	Claimed	Standard addition			Recovery (Mean \pm SD)	
Product	(μg/mL)	Added	Found	Recovery	Proposed	Standard
		(µg/mL)	(µg/mL)	(%)	method	addition
		5	5.05	101.00		99.78 ± 1.086
Benazepril	40	10	9.89	98.90	100.45 ± 0.248	
		15	14.92	99.47		
		5	4.96	99.20		
Amlodipine	10	10	9.86	98.60	99.86 ± 0.485	99.02 ± 0.367
		15	14.89	99.27		

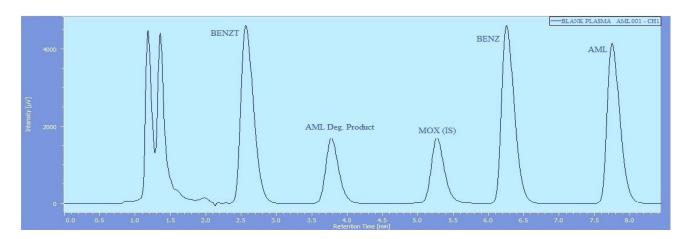


FIG. 6. HPLC chromatogram of spiked human plasma sample showing no interference with the analytes (5 μ g/mL each).

TABLE 6. Determination of the analyte in spiked human plasma by the proposed RP-HPLC method.

Spiked Concentration μg/mL		Recovery % ± SD*			
Benazepril	Benazepril Benazeprilat Amlodipine		Benazepril	Benazeprilat	Amlodipine
1	1	10	85.47 ± 3.234	88.34 ± 2.715	80.26 ± 3.471
2	2	20	84.22 ± 4.724	88.82 ± 3.421	81.35 ± 3.826
5	5	30	84.63 ± 4.871	87.44 ± 2.514	80.64 ± 5.286

^{*}The mean percentage recovery of 3-separate determinations

Statistical analysis

Results obtained by the proposed method for the determination of pure samples of BENZ, BENZT and AML are statistically compared to those obtained by the reported methods. The calculated t and F values were found to be less than their corresponding theoretical ones confirming good accuracy and excellent precision TABLE 7. Separation was achieved on Zorbax SB C18, 5 μ , 250 mm \times 4.6 mm i.d. column using phosphate buffer and acetonitrile in the proportion of 65:35 (v/v) with apparent pH adjusted to 7.0 at a flow rate of 1ml/min, and UV detection at 240 nm using a photodiode array detector.

Stability

BENZ, BENZT, and AML working solutions showed no changes up to two weeks when stored at room temperature.

TABLE 7. Statistical comparison for the results obtained by the proposed methods and the reference methods [6] for the determination of amlodipine and benazepril in pure powder form.

Parameter	Benazepr	il	Amlodipine		
1 arameter	Reference Method	RP-HPLC	Reference Method	RP-HPLC	
Mean	100.41	99.54	100.12	100.73	
SD	0.482	1.045	0.924	1.356	
N	6	6	6	6	
Variance	0.232	1.092	0.853	1.838	
Student t (2.23) ^a		1.325		1.751	
F test (5.05) ^a		4.706		2.154	

^aThe values in parenthesis are the corresponding theoritical values of t and F at p=0.05.

Conclusion

In summary, we developed and validated a highly sensitive, specific, reproducible and high-throughput stability indicating chromatographic methods for simultaneous quantification of BENZ, BENZT, and AML in bulk powder and pharmaceutical formulation. Also the chromatographic method was applied on spiked human plasma. According to the validation parameters, we concluded that the developed method could be useful for quality control laboratories and bioequivalent studies with desired precision and accuracy.

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