



ANTIOXIDANT PROPERTIES OF *POLYGALA JAVANA* DC WHOLE PLANT (POLYGALACEAE) IN ALLOXAN INDUCED DIABETIC RATS

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ABSTRACT

Administration of ethanol extract of *Polygala javana* whole plant (100 mg/Kg and 200 mg/Kg body weight) to alloxan induced diabetic rats for 14 days reduced the elevated level of lipid peroxidation (LPO). The treatment also resulted in significant increase in reduced glutathione (GSH), glutathione peroxidase (GPx), superoxide dismutase (SOD) and catalase (CAT) in serum, liver and kidney. The results confirm the antioxidant activity of *P. javana* whole plant and suggest that because of its antioxidant effects its administration may be useful in controlling the diabetic complications in experimental diabetic rats.

Key words: Antioxidant, *Polygala javana*, Oxidative stress.

INTRODUCTION

Diabetes is usually associated with an increased production of the molecules of reaction oxygen species (ROS) and / or impaired antioxidant defense systems, which result in oxidative damage leading to ROS mediated diabetic pathogenesis. Disturbances of the antioxidant defense system in diabetes involve enhancement of lipid peroxidation, alteration in antioxidant enzymes and impaired glutathione metabolism¹.

Oxidative stress is the imbalance between the generation of ROS and the body defense mechanisms. Environmental pollutants, toxic habits, inadequate nutrition, excess solar radiation, large exposure to toxic substances, drug metabolism, and a high physical or physical stress are the most common exogenous factors originating ROS in the human body². Oxidative stress has also been implicated in the pathogenesis of diabetes, liver damage, nephrotoxicity, inflammation, cancer, cardiovascular disorders, neurologic disorders as well as in the process of aging³.

Recently, many investigations indicated that supplementation with nature antioxidants may allviate the oxidative damage in diabetes^{4,5}. In particular, it is recognized that consumption of natural antioxidants of fruits, vegetable and grains are important for the prevention of chronic illnesses in diabetes patients^{6,7}. A considerable interest has grown in finding both hypoglycemic and antioxidative properties of natural antioxidants for the treatment of diabetes⁸. Plants often contain substantial amounts of antioxidants, flavonoids and tannins⁹. Implication of oxidative stress in the pathogenesis of diabetes is regulated not only by oxygen free-radical generation but also due to non-enzymatic protein glycosylation and alteration in antioxidant enzymes¹⁰.

Polygala was traditionally used by Americans to treat snake bites¹¹ and as an expectorant to treat cough and bronchitis. *Polygala* is considered as a powerful tonic herb¹² that can help to develop the mind and aid in creative thinking. However, in spite of traditional use, pharmacology of its whole part has not yet been explored scientifically. To our knowledge no report on the *in vivo* antioxidant activity of effect of *Polygala javana* whole plant on alloxan induced diabetic rats. This study was therefore undertaken to evaluate the effect of ethanol extract of whole plant of *Polygala javana* on *in vivo* antioxidant activity in alloxan induced diabetic rats.

EXPERIMENTAL

Materials and methods

Plant material

The whole plant of *Polygala javana* were freshly collected from the well grown healthy plants inhabiting the natural region of Scott Christian College, Nagercoil, Kanyakumari district, Tamil Nadu. The plant were identified and authenticated in Botanical Survey of India, Southern Circle, Coimbatore, Tamil Nadu, India. A voucher specimen was deposited in Ethnopharmacology Unit, Research Department of Botany, V. O. Chidambaram College, Tuticorin, Tamil Nadu.

Preparation of plant extract for phytochemical screening and antidiabetic studies

The *P. javana* whole plant were shade dried at room temperature and the dried whole plant were powdered in a Wiley mill. Hundred grams of powdered *P. javana* whole plant was packed in a Soxhlet apparatus and extracted with ethanol. The extracts were subjected to qualitative test for the identification of various phytochemical constituents as per the standard procedures^{13,14}. The ethanol extracts were concentrated in a rotary evaporator. The concentrated ethanol extract were used for antidiabetic studies.

Animals

Normal healthy male Wistar albino rats (180-240 g) were housed under standard environmental conditions at temperature ($25 \pm 2^\circ\text{C}$) and light and dark (12 : 12 h). Rats were fed with standard pellet diet (Goldmohur brand, MS Hindustan lever Ltd., Mumbai, India) and water *ad libitum*.

Acute Toxicity Study

Acute oral toxicity study was performed as per OECD – 423 guidelines (acute toxic class method), albino rats (n = 6) of either sex selected by random sampling were used for acute toxicity study¹⁵. The animals were kept fasting for overnight and provided only with water, after which the extracts were administered orally at 5 mg/Kg body weight by gastric incubations and observed for 14 days. If mortality was observed in two out of three animals, then the dose administered was assigned as toxic dose. If mortality was observed in one animal, then the same dose was repeated again to confirm the toxic dose. If mortality was not observed, the procedure was repeated for higher doses such as 50, 100, and 2000 mg/Kg body weight.

Induction of diabetes in experimental animal

Rats were induced diabetes by the administration of simple intraperitoneal dose of alloxan monohydrate (150 mg/Kg)¹⁶. Two days after alloxan injection, rats screened for diabetes having glycosuria and hypoglycemia with blood glucose level of 200-260 mg/100 mL were taken for the study. All animals were allowed free access to water and pellet diet and maintained at room temperature in plastic cages.

Experimental design

In the present investigation, a total of 30 rats (24 diabetic surviving rats and 6 normal rats) were taken and divided into five groups of 6 rats each.

Group I : Normal untreated rats

Group II : Diabetic control rats

Group III : Diabetic rats given ethanol extract of *P. javana* whole plant (100 mg/Kg body weight)

Group IV : Diabetic rats given ethanol extract of *P. javana* whole plant (200 mg/Kg body weight)

Group V : Diabetic rats given standard drug glibenclamide (600 µg/Kg body weight).

The animals were sacrificed at the end of experimental period of 14 days by decapitation. Blood was collected, sera separated by centrifugation at 3000 g for 10 minutes and serum was stored at -4°C until analyses completed. The liver and kidney tissues were excised, rinsed in ice cold saline, cut into small pieces and homogenized with homogenizer in Tris-HCl buffer (pH 7.4). The homogenate was centrifuged at 10,000 rpm for 10 min. Supernatant was used for enzyme assays for the estimation of non enzymatic and enzymatic antioxidants such as lipid peroxidation (LPO)¹⁷; superoxide dismutase (SOD)¹⁸ catalase (CAT)¹⁹, glutathione peroxidase (GPx)²⁰ and reduced glutathione (GSH)²¹.

RESULTS AND DISCUSSION

The phytochemical screening of ethanol extract of *P. javana* whole plant revealed the presence of alkaloid, catechin, coumarin, flavonoid, tannin, saponin, steroid, phenol, glycoside, terpenoid and xanthoprotein. Acute toxicity study revealed the non-toxic nature of the ethanol extract of *P. javana* whole plant. The results (Tables 1, 2 and 3) showed increased lipid peroxidation (LPO) in serum, liver and kidney of alloxan induced diabetic rats. Earlier studies have reported that there was an increased lipid peroxidation in liver, kidney and brain of diabetic rats^{22,23}. This may be, because the tissues contain relatively high concentration of early peroxidizable fatty acids. In the present study, an increase in the levels of LPO was found and there levels were significantly reduced after the supplementation of the ethanol extract of *P. javana* and glibenclamide (Tables 1, 2 and 3). This indicates that plant extract inhibit oxidative damage due to the antiperoxidative effect of ingredients present in ethanol extract of *P. javana*. This should be correlated with previous study reported that *Cassia auriculata* flower, *Syzigium cumini*, *Tinospora cardifolia*, *Scoparia dulcis* and *Nigella sativa*²⁴⁻²⁸ has antiperoxidative and antihyperlipidaemic effect of diabetic animals. Apart from the regulation of carbohydrate metabolism, insulin also plays an important role in the lipid metabolism. Insulin is a potent inhibitor of lipolysis, since it inhibits the activity of hormone sensitive lipase in adipose tissue and suppresses the release of free fatty acids²⁹.

The levels of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx) and reduced glutathione (GSH) (Tables 1, 2 and 3) were significantly reduced in serum, liver and kidney of alloxan induced diabetic rats. These adverse changes were reversed to near normal values in ethanol extract of *P. javana* whole plant treated. It is well known that CAT, SOD and GPx play an important role as protective enzymes against free radical formation of tissues³⁰. SOD has been postulated as one of the most important enzymes in the enzymatic antioxidant defense system which catalyses the dismutation of superoxide radicals to produce H₂O₂ and molecular oxygen³¹. Hence diminishing the toxic effects caused by their radical. The observed decrease in SOD activity could result from inactivation by H₂O₂ or by glycation of enzymes³². The superoxide anion has been known to inactivate CAT, which involved in the detoxification of hydrogen peroxide³³. Thus, the increase in SOD activity may indirectly play an important role in the activity of catalase.

Table 1: Effect of *Polygala javana*, extracts on serum LPO, GPx, GSH, SOD and CAT in the normal, diabetic and drug treated rats

Groups	Parameters				
	LPO (nanomol/ mg protein)	GPx (u/mg protein)	GSH (u/mg protein)	SOD (u/mg protein)	CAT (u/mg protein)
I	1.63 ± 0.08	621.14 ± 34.19	31.96 ± 1.36	428.64 ± 31.44	73.94 ± 2.48
II	3.09 ± 0.03**	298.24 ± 26.33***	20.33 ± 1.03*	278.08 ± 23.11***	60.14 ± 1.94*
III	2.68 ± 0.14	498.26 ± 17.21	21.48 ± 1.04	321.08 ± 16.51	63.14 ± 1.31
IV	1.93 ± 0.02	518.19 ± 24.08	26.16 ± 1.23	378.99 ± 26.57	69.84 ± 1.63
V	1.54 ± 0.11	593.94 ± 23.08	29.29 ± 1.08	391.56 ± 19.36	78.23 ± 1.47

Each value is SEM ± 6 individual observations

* $P < 0.05$; ** $P < 0.01$, *** $P < 0.001$ Compared normal control vs -Diabetic rats

Table 2: Effect of *Polygala javana*, extracts on liver LPO, GPx, GSH, SOD and CAT in the normal, diabetic and drug treated rats

Groups	Parameters				
	LPO (nanomol/mg protein)	GPx (u/mg protein)	GSH (u/mg protein)	SOD (u/mg protein)	CAT (u/mg protein)
I	0.094 ± 0.013	8.14 ± 0.12	49.11 ± 1.37	5.27 ± 0.93	81.96 ± 2.04
II	0.173 ± 0.021**	3.91 ± 0.09*	12.94 ± 1.14*	2.09 ± 0.05**	62.22 ± 1.86**
III	0.156 ± 0.018	4.21 ± 0.15	28.26 ± 1.14	2.84 ± 0.09	68.26 ± 1.31
IV	0.131 ± 0.017	5.27 ± 0.11	39.46 ± 1.26	3.19 ± 0.27	78.53 ± 1.33
V	0.081 ± 0.001	6.93 ± 0.16	48.53 ± 1.76	4.97 ± 0.13	83.66 ± 1.22

Each Value is SEM ± 6 individual observations

* $P < 0.05$; ** $P < 0.01$ Compared normal control vs -Diabetic rats

Table 3: Effect of *Polygala javana*, extracts on Kidney LPO, GPx, GSH, SOD and CAT in the normal, diabetic and drug treated rats

Groups	Parameters				
	LPO (nanomol/mg protein)	GPx (u/mg protein)	GSH (u/mg protein)	SOD (u/mg protein)	CAT (u/mg protein)
I	0.078 ± 0.004	5.26 ± 0.19	31.56 ± 1.24	15.13 ± 1.03	43.11 ± 1.74
II	1.843 ± 0.014*	2.13 ± 0.12**	12.92 ± 1.07**	7.96 ± 0.36*	13.96 ± 1.03**
III	1.436 ± 0.009	3.30 ± 0.11	18.49 ± 1.30	9.54 ± 0.31	26.48 ± 1.12
IV	1.101 ± 0.021	3.81 ± 0.18	25.76 ± 1.34	13.33 ± 0.74	36.31 ± 1.27
V	0.086 ± 0.004	5.64 ± 0.24	28.11 ± 1.09	18.39 ± 0.36	40.56 ± 1.11

Each Value is SEM ± 6 individual observations

* $P < 0.05$; ** $P < 0.01$ Compared normal control vs -Diabetic rats

Catalase (CAT) is a heme protein, which catalyses the reduction of hydrogen peroxides and protects the tissues from highly reactive hydroxyl radicals³⁴. The decrease in CAT activity could result from inactivation by glycation of enzyme³⁵. Reduced activity of SOD and CAT in the serum, liver and kidney have been observed during diabetes and this may result in a number of deleterious effects due to the accumulation of superoxide radicals and hydrogen peroxides³⁶. The reductions of hepatic SOD and CAT activities in alloxan induced diabetic rats when compared with normal rats were reported³⁷. Whereas, the extract treated groups showed a significant increase in the hepatic SOD and CAT activities of the diabetic rats. This means, that the extracts can reduce the potential glycation of enzymes or they may reduce reactive oxygen free radicals and improve the activities of antioxidant enzymes.

GSH is a major non-protein thiol in living organisms, which plays a central role in co-ordinating the body's antioxidant defense processes. Perturbation of GSH status of a biological system can lead to serious consequences. GPx catalyses the reaction of hydroperoxides with reduced glutathione to form glutathione disulphate (GSSG) and the reduction product of the hydroperoxide. In the present study, decline in the activities of these enzymes in alloxan induced rats and attainment of normalcy in *P. javana* whole plant extract treated rats indicate that oxidative stress elicited by alloxan was significantly reduced by this extract .

The present study reveals that the *P. javana* whole plant extract had antioxidant activity. The bioactive components, responsible for the observed activities are not precisely known but it may be one or more of the phytochemical constituents established to be present in the whole plant extracts. In the present study, phytochemical screening reported that the presence of phenolics and flavonoids in extracts, which might be the constituents responsible for the antioxidant activities. Further identification and isolation of these compounds may be fruitful.

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