Antidiabetic, antihyperlipidemic and histopathological analysis of Cleome droserifolia methanolic extract on alloxan induced diabetic rats

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ABSTRACT

Objective: To investigate antidiabetic, hypolipidemic histopathological analysis of Cleome droserifolia methanolic extract (CDE) extract in alloxan induced diabetic rat by administering oral dose (0.31 g /kg body weight / day). Methods: Optimal cutoff level of each of the four plasma glucose values of oral glucose tolerance test in alloxan diabetic rat was done. Other parameters as liver profile, renal profile and total lipid levels were determined in normal and alloxan induced diabetic rats after oral administration of the extract for 30 days. Histopathological changes in diabetic rat organs (pancreas, liver and kidney were also observed after extract treatment. Results: Daily oral administration CDE (0.31 g /kg body weight) and glimebride (10 microg/g body wt) showed beneficial effects on blood glucose level (P<0.001) as well as improving kidney, liver functions and hyperlipedemia due to diabetes. The extract treatment also showed enhanced serum insulin level and body weight of diabetic rats as compared to diabetic control group. Furthermore, the extract has a favorable effect on the histopathological changes of the pancreas, liver and kidney in alloxan induced diabetes. Conclusion: Cleome droserifolia posses antidiabetic property as well as improve body weight, liver profile, renal profile and total lipid levels. CDE has also favorable effect to inhibit the histopathological changes of the pancreas and kidney in alloxan induced diabetes.

INTRODUCTION

Diabetes mellitus (DM) is a very commonly occurring metabolic disorder characterized by hyperglycemia and altered metabolism of carbohydrates, lipids and proteins. DM occurs due to absolute or relative deficiency of insulin or insulin resistance.[12].

This metabolic disorder affects approximately 4% of the population worldwide and is expected to be increased by 5.4% in 2025. Because DM is associated with oxidative stress, it alters the cellular microenvironment in many different types of tissues causing myriad untoward effects, collectively referred to as 'diabetic complications'. Two cellular processes affected by dia-
betes are inflammation and apoptosis[30].

Glimepiride is a medium-to-long acting sulfonylurea anti-diabetic drug, sometimes classified as the first third-generation sulfonylurea and sometimes classified as second-generation. Glimepiride acts as an insulin secretagogue. It lowers blood sugar by stimulating the release of insulin by pancreatic beta cells and by inducing increased activity of intracellular insulin receptors[33].

Synthetic drugs usually come with considerable side effects, such as hypoglycemia, drug-resistance, dropsy, and weight gain. In recent years due to the adverse effects of synthetic hypoglycemic drugs, interests in alternate therapeutic approach have become very popular. Nowadays, herbal drugs are gaining popularity in the treatment of diabetes and its complications due to their efficacy, low incidence of side effects and low cost.

The medicinal plants provide a useful source of oral hypoglycemic compounds for the development of new pharmaceutical leads. Some of the plants which are being used for the treatment of diabetes have received scientific or medicinal scrutiny and even the WHO expert committee on diabetes recommends that this area warrant further attention[32].

*Cleome droserifolia* (Forssk.) Del., is a plant of the Cleomaceae family. Methanolic extract of leaves and stems for *Cleome droserifolia* (CDE) is rich in Bioactive compounds as flavanoids, flavonol glycosides, alkaloids, tannins and Steroids as shown in Figure (2.8). So far as plant phenolics constitute one of the major groups of compounds acting as primary antioxidants or free radical terminators, Flavonoids as one of the most diverse and widespread group of natural compounds are probably the most important natural phenolics as they possess radical scavenging properties.

MATERIALS AND METHODS

**Glimepiride**

The drug was purchased from (Delta Pharma for Pharmaceuticals, Egypt). Oral dose: Oral administration of (10 microg/g body wt) daily in accordance with[16].

**Plant material**

Mature whole *Cleome droserifolia* were collected from Eastern desert, Red Sea region, Sinai, Egypt and authenticated as *Cleome droserifolia* by Dr. H. Ezzat, Department of Pharmacognacy, Faculty of pharmacy, Minia university.

**Extract preparation**

The freshly collected leaves and stem part of *Cleome droserifolia* were washed with distilled water and air-dried under the control conditions and powdered. The powdered plant material was percolated with petroleum ether to remove fatty substances; the marc was further exhaustively extracted with of 80% methanol for 3 days. The extract was filtered, concentrated on rotavapour and then freeze-dried under high vacuum (1.3 Pa) and at temperature of - 40± 2°C. The extract will be dissolved in 0.5 g Carboxy methyl cellulose (0.5w/v) for oral administration.

**Chemicals**

Alloxan was purchased from Loba chemie Pvt. Ltd. Mumbai, India. Total cholesterol (TC), serum high-density lipoprotein (HDL), serum Creatinine (SC), serum urea (SU), alanine transaminase (ALT), serum aspartate transaminase (AST) and triglyceride (TG) standard kits were obtained from Erba diagnostics Mannheim Gambh, Germany. Blood glucose level was measured using Elegance glucose meter (CT-X10) of Convergent Technologies, Germany. All reagents used in study were analytical grade.

**Animals**

40 White male albino rats weighting about 190±10 g were used as experimental animals in the present investigation. The animals were housed in standard polypropylene cages and maintained under controlled room temperature (22±2 æ°C) and humidity (55±5%) with 12 h light and 12 h dark cycle and were fed a standard diet of known composition, and water ad libitum. The chow was purchased from El-Gomhoria Company, Cairo, Egypt. They were housed for two weeks for accommodation. Our work was carried out in accordance with the guidelines of El Minia University for animal use. These animals were used for induction of Diabetes mellitus.

**Induction of diabetes**

Rats were made diabetic by a single intraperitoneal injection of alloxan monohydrate (Loba Chemie,
TABLE 1: Effect of CDE on optimal cutoff level of each of the four plasma glucose values of oral glucose tolerance test in alloxan induced diabetic rats (A-D)(n=10).

<table>
<thead>
<tr>
<th>Group</th>
<th>Fasting glucose level</th>
<th>1. h glucose level</th>
<th>2. h glucose level</th>
<th>3. h glucose level</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal (N)</td>
<td>69.12±2.3</td>
<td>112.3±9.8</td>
<td>89.3±7.2</td>
<td>74.9±6.8</td>
</tr>
<tr>
<td>Diabetic Control (DC)</td>
<td>142±12.3 ***a</td>
<td>465±41.8 ***a</td>
<td>435±30.25 ***a</td>
<td>412±38.5 ***a</td>
</tr>
<tr>
<td>Diabetic + Glimepride (G)</td>
<td>176±16.2 **a</td>
<td>484±40.28 **a</td>
<td>401±29.8 **a</td>
<td>314±27.1 **a</td>
</tr>
<tr>
<td>Diabetic + CDE (CDE)</td>
<td>125±9.4 **b</td>
<td>435±36.44 **a</td>
<td>215±17.49 **b</td>
<td>139±7.4 **b</td>
</tr>
</tbody>
</table>

* Values significantly different compared to normal P***<0.001; * Values are expressed as means ± SE. Means not sharing common letter are significantly different (p<0.05) based on one--way ANOVA with Tukey’s post-hoc test.

Bombay; 150 mg/kg i.p.) in sterile saline. Twelve days after Alloxan injection, rats with blood glucose level of >200 mg/dL were separated and used for the study. Blood glucose levels were measured using blood glucose test strips with elegance glucometer (Frankenberg, Germany) at weekly intervals till the end of study (i.e. 3 weeks). Blood glucose estimation was done on 0, 1, 2 and 3 hours after administration of extract orally at the 30th day of starting experiment.

**Experimental design**

Overnight fasted rats were divided into four groups and for each group 10 animals and treated orally once a day for 30 days as follows:

- Group I: Normal healthy control: given only vehicle (Tween)
- Group II: Diabetic control: given only vehicle (Tween 80, 5% 80, 1% v/v) v/v)
- Group III: Diabetic rats given Glimepride (10 microg/g body wt.)
- Group IV: Diabetic rats given CDE 0.31 g/kg b body weight /day

**Biochemical parameters**

Optimal cutoff level of each of the four plasma glucose values of oral glucose tolerance test in alloxan diabetic was measured with elegance glucometer (Frankenberg, Germany) at hour intervals i.e. 0, 1, 2 and 3 hours after daily administration of extract orally. After blood glucose estimation on day 30, whole blood was collected by cardiac puncture under mild ether anesthesia from rats. Serum cholesterol, triglycerides, creatinine, urea, and HDL cholesterol levels were also evaluated in normal and alloxan induces diabetic rats. Serum alanine transaminase (ALT) and serum aspartate transaminase (AST) were measured by autoanalyser

**TABLE 2: Effect of CDE on serum insulin level in alloxan induced diabetic rats (A-D)(n=10)**

<table>
<thead>
<tr>
<th>Group</th>
<th>Serum insulin levels (ng/ml) After 30 days of treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal (N)</td>
<td>1.18±0.07</td>
</tr>
<tr>
<td>Diabetic control (DC)</td>
<td>0.48±0.07***a</td>
</tr>
<tr>
<td>Diabetic + Glimepride (+G)</td>
<td>0.62±0.04 **a</td>
</tr>
<tr>
<td>Diabetic + CDE (CDE)</td>
<td>0.90±0.04 **b</td>
</tr>
</tbody>
</table>

* Values significantly different compared to normal P***<0.001; * Values are expressed as means ± SE. Means not sharing common letter are significantly different (p<0.05) based on one--way ANOVA with Tukey’s post-hoc test.

(Erra Chem 7, Mannheim, Germany) using Erba diagnostic kits. Serum insulin levels were determined using insulin ELISA kit.

**Statistical analysis**

Statistical analysis was carried out using Graph Pad Instat software (version 3, ISS-Rome, Italy). Groups of data were compared with ANOVA, followed by Tukey-Kramer (TK) multiple comparisons post-test. Values of P < 0.05 were regarded as significant. Data were expressed as mean ± standard error (SEM).

**RESULTS**

**Antidiabetic activity**

Single dose alloxan monohydrate (150 mg/kg) significantly (P<0.01) increases the blood glucose as shown in TABLE 1. After the daily oral administration with CDE, for 30 days, significant decreased (P<0.01) in the blood glucose levels was observed in the diabetic rats (TABLE 1. The reduced insulin level in diabetic rats was also significantly improved by treatment of CDE. At the end of experiment (30th day) blood glucose level was (139±7.4) mg/dL of the groups treated
Effect on body weight of rats

In diabetic rats, continuous reduction in body weight was observed as shown in TABLE 2. CDE treatment significantly (P<0.05) improved the body weight of diabetic rats.

Effect on lipid profile

In diabetic rats, there was a significant increase of serum total cholesterol, triglycerides (TABLE 3) and significant decrease in HDL cholesterol in compared to that of normal control. The standard drugs as well as CDE used in the experimental study significantly decreased (P<0.05) the levels of cholesterol and triglycerides whereas HDL cholesterol significantly increased (TABLE 4).

Effect on liver functions

The effect of CDE on liver functions is represented in the TABLE 5. ALT, AST were significantly elevated in alloxan induced diabetes. The rats treated with CDE showed significant (P<0.01) reduction in the elevated levels of liver enzymes (transaminases) in a dose dependent manner as shown in TABLE 5.

Effect on kidney functions

Kidney function markers like urea and creatinine were elevated in the alloxan induced diabetic rats when compared with the normal rats. CDE reduced both the levels in dose dependent manner (TABLE 6).

Histology of pancreas

Histology of pancreas (Figure 1) showed normal acini, and normal cellular in the islets of langerhans in the pancreas of normal control (1.a). In diabetic animals treated extensive damage to islets of langerhans and reduced dimensions of islets were observed in diabetic rats (1.b). On other hand, CDE treatment showing few vacuolizations and appear highly divided β-cells in the islets of Langerhans (1.d).

Histology of liver

Photomicrographs of liver (Figure 2) showed normal

### TABLE 3: Effect of CDE on body weight, triglycerides (T.G) and total cholesterol (T.C) in alloxan induced diabetic rats.

<table>
<thead>
<tr>
<th>Group</th>
<th>Body weight gain</th>
<th>T.G</th>
<th>T.C</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>gm</td>
<td>mg/dl</td>
<td>mg/dl</td>
</tr>
<tr>
<td>Normal (N)</td>
<td>37±3.21</td>
<td>74.2±6.8</td>
<td>87.2±8.4</td>
</tr>
<tr>
<td>Diabetic Control (DC)</td>
<td>-27±3.4</td>
<td>178.2±10.4</td>
<td>163.2±7.9</td>
</tr>
<tr>
<td>Diabetic + Glimepride (G)</td>
<td>-21±2.6</td>
<td>120.3±8.6</td>
<td>102.3±7.4</td>
</tr>
<tr>
<td>Diabetic+ CDE (CDE)</td>
<td>-9±1.2</td>
<td>110.3±7.1</td>
<td>99.26±5.6</td>
</tr>
</tbody>
</table>

* Values significantly different compared to normal P***<0.001; * Values are expressed as means ± SE. Means not sharing common letter are significantly different (p<0.05) based on one–way ANOVA with Tukey’s post–hoc test.

### TABLE 4: Effect of CDE on HDL cholesterol and LDL cholesterol in alloxan induced diabetic rats

<table>
<thead>
<tr>
<th>Group</th>
<th>HDL.C</th>
<th>LDL.C</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>mg/dl</td>
<td>mg/dl</td>
</tr>
<tr>
<td>Normal (N)</td>
<td>63.2±5.7</td>
<td>13.5±7.3</td>
</tr>
<tr>
<td>Diabetic Control (DC)</td>
<td>39.3±4.5</td>
<td>53.2±4.8</td>
</tr>
<tr>
<td>Diabetic + Glimepride (G)</td>
<td>45.2±3.4</td>
<td>35.1±3.3</td>
</tr>
<tr>
<td>Diabetic+ CDE (CDE)</td>
<td>46.2±3.2</td>
<td>30.2±2.9</td>
</tr>
</tbody>
</table>

* Values significantly different compared to normal P***<0.001; * Values are expressed as means ± SE. Means not sharing common letter are significantly different (p<0.05) based on one–way ANOVA with Tukey’s post–hoc test.

### TABLE 5: Effect of CDE on liver parameters in normal and diabetic rats

<table>
<thead>
<tr>
<th>Group</th>
<th>sALT</th>
<th>sAST</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>U/l</td>
<td>U/l</td>
</tr>
<tr>
<td>Normal (N)</td>
<td>19.51±0.84</td>
<td>23.17±0.75</td>
</tr>
<tr>
<td>Diabetic Control (DC)</td>
<td>62.39±0.87**a</td>
<td>84.29±0.94***a</td>
</tr>
<tr>
<td>Diabetic + Glimepride (G)</td>
<td>59.18±0.91a</td>
<td>70.06±0.89a</td>
</tr>
<tr>
<td>Diabetic+ CDE (CDE)</td>
<td>44.27±0.38b</td>
<td>47.18±0.54b</td>
</tr>
</tbody>
</table>

* Values significantly different compared to normal P***<0.001; * Values are expressed as means ± SE. Means not sharing common letter are significantly different (p<0.05) based on one–way ANOVA with Tukey’s post–hoc test.
TABLE 6: Effect of CDE on kidney parameters in normal and diabetic rats (n=10)

<table>
<thead>
<tr>
<th>Group</th>
<th>Urea (mg/dl)</th>
<th>Creatinine (mg/dl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal (N)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Diabetic Control (DC)</td>
<td>66.14±2.14***a</td>
<td>1.31±0.01***a</td>
</tr>
<tr>
<td>Diabetic + Glimepride (G)</td>
<td>29.26±2.65b</td>
<td>0.98±0.03 b</td>
</tr>
<tr>
<td>Diabetic+ CDE (CDE)</td>
<td>31.2±2.11b</td>
<td>0.87±0.03 c</td>
</tr>
</tbody>
</table>

* Values significantly different compared to normal P***≤0.001; * Values are expressed as means ± SE. Means not sharing common letter are significantly different (p<0.05) based on one–way ANOVA with Tukey’s post–hoc test.

Figure 1: Effect of CDE on pancreas of alloxan induced diabetic rats

Figure 2: Effect of CDE on liver of alloxan induced diabetic rats
mal hepatic cells with well preserved cytoplasm, nucleus, nucleolus and central vein (2.a). In case of group II diabetic rats, the normal lobular structure was preserved. The central vein was prominent and prominently congested. Focal areas of hemorrhage were also seen. Fatty change was evident.

The portal tracts appeared normal (2.b). In group IV [diabetic rats + CDE mg/kg], showing kupffer cells activation (2.d).

**Histology of kidney**

Histology of kidney (Figure 3) in normal animals showed normal structure (3.a). In diabetic rats, mild thickening of the basement membrane of the arterioles of glomeruli along with mild change of density of mesangial mesangium were observed. No other significant changes were seen (3.b). After CDE 400 mg/kg treatment, these changes were improved towards normal condition (3.d).

**DISCUSSION**

Glucose is the key physiological regulator of insulin secretion; indeed, short-term exposure of β-cells to increasing glucose concentrations induces proliferation in a concentration-dependent manner. In addition to its effect on β-cell turnover, hyperglycemia also impairs β-cell secretory function. This glucotoxic effect is evident before apoptosis leads to a significant decrease in β-cell mass\(^{[10]}\).

The present investigation indicated that a single dose of alloxan (150 mg/kg) intraperitoneally to adult male albino rats (210-220g) was suitable to induce histological changes of the islets of Langerhans characterized appearance, hypoinsulinemia and hyperglycemic state. The present dose as well as the observed histopathological and biochemical manifestations agree with the literature of\(^{[13]}\).

A gradual loss of β-cells due to apoptosis significantly hinders insulin production and inhibits cell viability. During apoptosis, cells shrink; chromatin condenses; DNA is cleaved into pieces at inter nucleosomal regions. A proactive way to increase β-cell viability is to decrease apoptosis level in order to retain the cell population and increase insulin production.

Oral administration of glimepiride causes significant decrease in levels of blood glucose in accordance with who revealed that glimepiride exerts its insulin-releasing effect mainly by inhibiting ATP-sensitive potassium channels. In the pancreatic β-cell this action induces depolarization of the cell membrane, allowing an influx of calcium in the cell. This in turn induces insulin release into the blood.

Pfützner et al\(^{[25]}\) revealed that the increase in the number of β-cells in the islets of Langerhan’s in glimepiride-treated diabetic rats in comparison to alloxan induced diabetic rats can be attributed to the fact that glimepiride affect the activation of the redox sensitive transcription factor NF (Kappa) β in alloxan in-
duced diabetic rats. Although the mechanism of β-cell neoformation is not clear but there is strong evidence that islet stem cells may exist in the pancreatic duct and that these ductal epithelial cells may be switched into a proliferative/regenerative phase leading to neodeoblastosis (neogenesis of islets).

Antidiabetic effect of glimepiride was further examined by histological observations made on the pancreatic tissue of glimepiride treated rats that show vacuolation and necrosis of some β-cells of islets of Langerhans.

Oral administration of CDE causes significant decrease in levels of blood glucose in accordance with [22]. [19] revealed that CDE has a hypoglycemic effect through potentiation of peripheral, hepatic insulin sensitivity and diminishing intestinal glucose absorption.

On other hand, [4] explained that flavonoids, the major active constituent in CDE are potential antidiabetic agents because they exert multiple actions that are both hypoglycemic (insulinomimetic action) and antihyperglycemic (insulin secretagogue). Also, flavonoid-enriched extract from efficiently inhibited α-glucosidase activity and may inhibit the non-Na+ dependent facilitated diffusion of monosaccharides in intestinal epithelial cells. Consequently, the parallel concentration Na+ dependent transport ATPase for monosaccharides gains efficiency [9].

Antidiabetic effect of CDE was further evidenced by histological observations made on the pancreatic tissue of CDE treated rats that show few vacuolization and highly divided β-cells in the islets of Langerhans.

The characteristic features of diabetic dyslipidemia are a high plasma triglyceride concentration, low HDL-cholesterol concentration and increased concentration of small dense LDL-cholesterol particles. The lipid changes associated with diabetes mellitus are attributed to increased flux of free fatty acids into the liver secondary to insulin deficiency/resistance. This results in excess fatty acid accumulation in the liver, which is converted to triglycerides. The impaired ability of insulin to inhibit free fatty-acid release leads to elevated hepatic VLDL-cholesterol production. The increased VLDL-cholesterol and triglyceride levels decrease the level of HDL-cholesterol and increase the concentration of small dense LDL-cholesterol particles by activation of lipoprotein lipase and lecithin acyl-cholesterol transferase [20].

In our study, elevated levels of serum TC, TG, LDL and VLDL-cholesterol and decreased HDL-cholesterol concentration in alloxan-induced diabetic rats are in accordance with [34]. On other hand, Induction of diabetes by alloxan resulted in loss of body weight in the diabetic control rats in accordance with that may due to catabolic effect on protein metabolism by retarding protein synthesis and stimulating protein degradation.

Oral administration of glimepiride causes significant decrease in the serum levels of triglycerides, total cholesterol and LDL-cholesterol in contrast to significant elevation in HDL-cholesterol and body weight in accordance with [11].

Motoyama et al. [19] revealed that glimepiride improved HDL-c level via improvement of plasma adiponectin level as adiponectin could increase HDL-c levels directly via increased lipoprotein lipase and decreased hepatic lipase activity. On other hand, the antilipidemic action of glimepiride may reside in their ability to stimulate insulin secretion and action.

Oral administration of CDE causes significant decrease in the serum levels of triglycerides, total cholesterol and LDL-cholesterol in contrast to significant elevation in HDL-cholesterol and body weight in accordance with [23].

Flavonoids in CDE inhibit the activity of cAMP-dependent protein phosphokinase, the consequence is that the cAMP concentration increases and that phosphorylation of the Hydroxy methyl glutaryl-CoA reductase, but endogenous cholesterol production is diminished. In addition, the flavonoids can interact with the enzyme protein phosphatase, which liberates the aliphatic phosphoesters from Hydroxy methyl glutaryl-CoA-CoA HMG-CoA reductase, thus restoring the activity of this. Thus, flavonoids inhibit HMG-CoA reductase by a dual mechanism [29].

The liver plays an important role in the maintenance of blood glucose levels by regulating its metabolism. Alloxan causes significant increase in activity of sAST and sALT, in accordance with that is directly related to changes in metabolism in which the enzymes are involved. The increased activities of transaminases, which are active in the absence of insulin due to the availability...
of amino acids in the blood of DM and are also responsible for the increased gluconeogenesis and ketogenesis\(^{[8]}\).

The present investigation indicated that, a single dose of alloxan (150 mg/kg) intraperitoneally to adult male albino rats (210-220g) was suitable to induce histological changes in the liver of alloxan induced diabetic rats with characterized appearance, enlarged and swollen hepatocytes\(^{[31]}\).

Oral administration of CDE causes significant decrease in activity of sAST, s ALT, hepatic Glycogen phosphorylase, hepatic G6P in accordance with\(^{[1]}\) due to estrogen-like plant-derived molecules as potent antioxidants.

Also, explained that Flavonoids, major active constituent in CDE possess common effects, namely, induction of apoptosis involving the release of cytochrome c from mitochondria, activation of caspases and down-regulation or up-regulation of Bcl-2 family members, but also induction of cell cycle arrest and inhibition of survival/proliferation signals.

Hepatic protective effect CDE of was further evidenced by histological observations made on the hepatic tissue of CDE treated rats that showing kupffer cells activation.

Alloxan led to a significant increase in serum urea and serum creatinine. A similar effect was recorded by\(^{[7]}\).\(^{28}\)revealed that enhanced protein catabolism and accelerated amino acid deamination for gluconeogenesis is probably an acceptable postulate to interpret the elevated levels of urea and creatinine. Furthermore, Alloxan increased the productions of reactive oxygen species, enhanced lipid peroxidation and protein carbonylation in association with decreased intracellular antioxidant defense in the kidney tissue.

Cao et al\(^{[8]}\) suggests that development of diabetic renal dysfunction may due to activation of endoplasmic reticulum stress that can mediate progressive endothelial damage through growth and migration of vascular smooth muscle and inflammatory cells, alteration of extracellular matrix, apoptosis of endothelial cells, over-expression of inflammatory cytokines.

Oral administration of glimepiride causes significant decrease in level of serum urea and creatinine in accordance with\(^{[27]}\).

In the present study the improvement in blood urea, serum creatinine and subsequent amelioration of histomorphological changes in kidneys of glimepiride treated rats can be attributed to the recovery of renal function which is explained by the regenerative capability of the renal tubules as good metabolic control is beneficial in slowing the progression of renal dysfunction in diabetes\(^{[18]}\).

Glimepiride could ameliorate the glomerular and tubular lesions that characterize diabetic renal dysfunction and subsequently recover renal morphology and function. Reno-protective effect glimepiride of was further evidenced by histological observations made on the renal tissue of glimepiride treated rats that revealed normal structure of renal parenchyma.

Renal dysfunction is a slowly progressive process that is postulated to be accelerated by intervening diseases, such as diabetes, due in part to the addition of excessive stress and inflammation\(^{[17]}\).

Oral administration of CDE causes significant decrease in level of seum urea and creatinine in accordance with\(^{[2]}\).

The mechanism by which the CDE prevents renal oxidative stress may include an increasing rate of GSH or by induction of its synthesis or by a scavenger effect. Instead of the toxic reactive metabolites binding to glutathione and consume, they will be captured by the flavonoids\(^{[24]}\).

Reno-protective effect of CDE was further evidenced by histological observations made on the renal tissue of CDE treated rats that revealed showing no histological changes.

**CONFLICT OF INTEREST STATEMENT**

We declare that we have no conflict of interest.

**ACKNOWLEDGEMENTS**

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