



# ADSORPTION STUDY OF CARBOFURAN ONTO MESOPOROUS ACTIVATED CARBON PREPARED FROM PALM OIL FRONDS

J. M. SALMAN<sup>\*</sup>, W. A. MAHMOUD, F. M. ABID<sup>a</sup> and  
A. A. MUHAMMED<sup>b</sup>

Ministry of Industry & Minerals, Quality Control Department, BAGHDAD, IRAQ

<sup>a</sup>College of Science for Woman, Baghdad University, BAGHDAD, IRAQ

<sup>b</sup>Ministry of Science & Technology, BAGHDAD, IRAQ

## ABSTRACT

The adsorption of insecticide, carbofuran from aqueous solution onto activated carbon derived from palm oil fronds (PFAC) was investigated through batch study. The effects of both initial concentration and pH of the carbofuran over the range of 25 to 250 mg/L and 2 to 12, respectively on the adsorption of PFAC were studied in batch experiments. Equilibrium data were fitted to the Langmuir, the Freundlich and the Temkin isotherm models. The results obtained from application of these models show that the best fits were achieved with the Langmuir model and a maximum monolayer adsorption capacity of 164 mg/g was obtained at 30°C.

The regeneration efficiency of spent activated carbon was studied and it was found to be 90.0-96.4%. The results indicated that PFAC has good capability as adsorbent for the removal of carbofuran from aqueous solutions.

**Key words:** Carbofuran, Adsorption, Desorption, Activated carbon, Palm oil fronds.

## INTRODUCTION

Pesticides are chemicals, which are commonly used in agriculture to protect crops from pest organisms including insects, plants, fungi, rodents and nematodes. They may appear as pollutants in water sources and threat to human health because of their toxicity, carcinogenicity and mutagenicity. In developed countries, drinking water quality has strict regulations regarding pesticides<sup>1</sup>.

---

\* Author for correspondence; E-mail: jasim\_63@yahoo.com, Mo.: 009647902508032

Presently, on a worldwide basis, intoxications attributed to pesticides have been estimated to be as high as 3 million cases of acute and severe poisoning annually, with many unreported cases and with some 220000 deaths<sup>2</sup>. This situation calls for urgent attention with acceptable solution for the removal of pesticides from water sources. This is because pesticides will continue to be used effectively for pest controls and the responsibility rest on us to find ways of avoiding many of the pesticides poisonings and contaminations that exist today.

Carbofuran is a broad spectrum systemic acaricides, insecticide and nematocide included in the general group of the carbamate derivative pesticides<sup>3</sup>. It is widely used for the control of soil dwelling and foliar feeding insects including wireworms, white grubs, weevils, stem borers, aphids and several other insects<sup>4</sup>. Carbofuran is known to be more persistent than other carbamate or organophosphate insecticides<sup>5</sup>. Carbofuran is degraded in water by hydrolysis, microbial decomposition and photolysis. In the soil, it is degraded by hydrolysis, microbial action, and to a lesser extent, photodecomposition<sup>6</sup>. The maximum acceptable concentration for carbofuran in drinking water is 0.09 mg/L<sup>7</sup>. Determining the level of carbofuran in soil or water has become increasingly important in recent years because of the widespread use of these compounds, which is due to their wide-ranging biological activity and relatively low persistence compared to organochlorine pesticides<sup>8</sup>.

There are several methods either independent or in conjunction that have been used for the removal of pesticides from water, such as chemical oxidation with ozone<sup>9</sup>, photocatalytic method<sup>10</sup>, combined ozone and UV irradiation<sup>11</sup>, ozonation<sup>12</sup>, membrane filtration<sup>13</sup> and adsorption<sup>14</sup>. Adsorption is one of the most frequently applied methods because of its efficiency, capacity and applicability on a large scale. The most commonly used adsorbent in adsorption processes is activated carbon due to its efficiency and economic feasibility. Utilization of activated carbon can be in the form of powder, granular and fiber or cloth. Recently, growing research interest in the production of carbon-based activated carbon has been focused on agricultural by-products. Low cost adsorbents derived from agricultural wastes have demonstrated outstanding capabilities for the removal of pollutants from wastewater. Therefore, low cost agricultural waste adsorbents can be viable alternatives to activated carbon for the treatment of contaminated wastewater. The use of cheap and eco-friendly adsorbents have been studied as an alternative substitution to activated carbon for the removal of dyes from wastewater<sup>15</sup>.

## **EXPERIMENTAL**

### **Materials**

Carbofuran supplied by Sigma–Aldrich was used as the adsorbate in this study. Deionized water was used to prepare all the solutions.

### **Preparation and characterization of activated carbon**

Palm oil fronds (POF) used as precursors for preparation activated carbon in this study, was cut into pieces, dried over night under 75°C to remove moisture until the weight was constant. The dried sample was then crushed using a grinder and thereafter screened to particle size range of 2-4 mm. The screened POF were then carbonized in a stainless steel, vertical tubular reactor, placed in a tube furnace. The temperature of the furnace was ramped from room temperature to 700°C at heating rate of 10°C/min and held for 2 h under nitrogen (99.995%) flowing at the rate of 150 cm<sup>3</sup>/min. The char produced from the carbonization process was subsequently impregnated with KOH pellets (KOH/Char = 2.75 by weight). The impregnated char was thermally treated under nitrogen to a final temperature of 850°C. Once the final temperature was reached, the nitrogen gas flow was switched over to CO<sub>2</sub> and held under that condition for 1 h. The PFAC produced was then cooled to room temperature under nitrogen flow (150 cm<sup>3</sup>/min) and thereafter washed with 0.1 M HCl and hot distilled water to bring the pH of the washing filtrate to about 7.

The surface area, pore size distribution, and the pore volume of the developed PFAC were analyzed using Micromeritics (Model ASAP 2020, USA – surface area and porosity analyzer) employing nitrogen adsorption isotherm at 77 K. In order to determine the surface morphology of the PFAC, the sample was examined using Scanning Electron Microscope (SEM) – model: Leo Supra 35 VP Field Emission SEM.

### **Effect of carbofuran initial concentration and solution pH**

In order to study the effect of carbofuran initial concentration and contact time on the adsorption uptake, 200 mL of carbofuran solutions with initial concentrations of 25–250 mg/L were prepared in a series of 250 mL Erlenmeyer flasks, and 0.30 g of the PFAC was added into each flask covered with glass stopper. The flasks were then placed in an isothermal water-bath shaker at 30°C, with agitation speed of 120 rpm. At specific time intervals, samples were withdrawn for analysis, using a double beam UV-vis spectrophotometer (Shimadzu UV-1700, Japan) at 273 nm, until equilibrium point was reached. The effect of solution pH on the carbofuran adsorption on PFAC was also examined by varying the initial pH of the solutions between 2 and 12 (using 0.1 M HCl

and/or 0.1 M NaOH as buffer solutions). In a typical run, the carbofuran initial concentration was fixed at 100 mg/L, with activated carbon dosage of 0.20 g/200 mL and solution temperature of 30°C.

### Equilibrium data fitting

Three isotherm models were used to test fit the experimental data, the Langmuir isotherm<sup>16</sup> the Freundlich isotherm<sup>17</sup> and the Timken isotherm<sup>18</sup>. The linear form of the Langmuir model is –

$$C_e/q_e = C_e/q_m + 1/K_a q_m \quad \dots(1)$$

Where  $C_e$  is the equilibrium concentration (mg/L);  $q_e$  the amount carbofuran adsorbed at equilibrium (mg/g);  $q_m$  the adsorption for complete monolayer (mg/g);  $K_a$  is the sorption equilibrium constant (L/mg).

The linear form of Freundlich isotherm is -

$$\ln q_e = \ln K_F + (1/n) \ln C_e \quad \dots(2)$$

The constants  $K_F$  and  $1/n$  of the Freundlich model are the constants indicative of the relative adsorption capacity of the adsorbent and the intensity of the adsorption, respectively.

The Timken isotherm has been used in the form as follows:

$$q_e = B \ln A + B \ln C_e \quad \dots(3)$$

Where  $B = RT/b$ ,  $b$  is the Temkin constant related to heat of sorption (J/mol);  $A$  is the Temkin isotherm constant (L/g),  $R$  the gas constant (8.314 J/mol K) and  $T$  the absolute temperature (K).

### Regeneration of activated carbon

The feasibility of regenerating the spent activated carbon was evaluated using ethanol desorption technique<sup>19</sup>. Batch equilibrium tests were performed on the fresh activated carbon prepared, where 100 mL of carbofuran solution with initial concentration of 200 mg/L were placed in a 250 mL Erlenmeyer flasks. 0.30 g of the fresh PFAC was added into the flask and placed in an isothermal water bath shaker at 30°C, with agitation speed of 120 rpm, for 48 h until complete equilibrium was attained. The spent activated carbon was

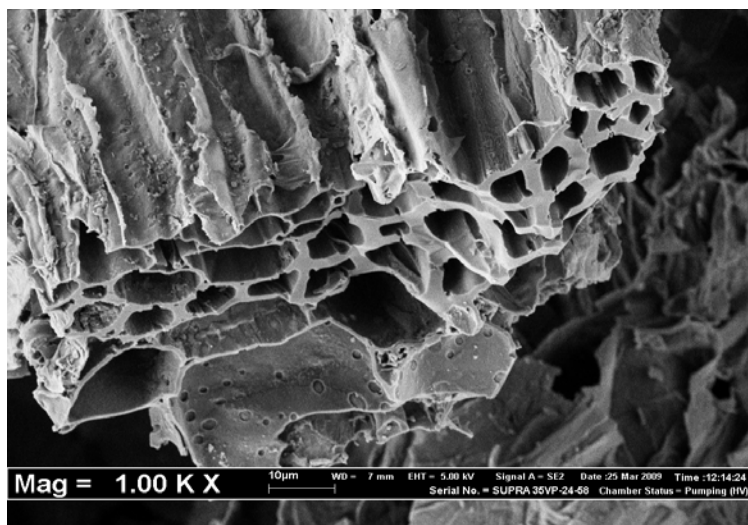
then separated from the solution and dried at 105°C in an oven. It was thereafter mixed with 100 mL of 95 vol. % ethanol in an Erlenmeyer flask for the desorption of the adsorbed carbofuran. The flask was kept in the isothermal water-bath shaker at the same temperature for the same time duration as the adsorption tests. Desorption percentage was calculated from equation 4:

$$\text{Desorption \%} = (C_{de} / C_{ad}) \times 100 \quad \dots(4)$$

## RESULTS AND DISCUSSION

### The morphological and textural characteristics of the developed PFAC

Fig. 1 shows the scanning electron microscopy (SEM) image of the produced PFAC. The PFAC depicts a surface containing a well-developed pores expected of a good absorbent, in which the carbonaceous matters and salts that could have blocked the pores as seen in the precursor had been leached off by the activation process, showing the efficacy of the thermo-chemical activation method adopted in this investigation. This observation is well corroborated by the Brunauer-Emmett-Teller (BET) surface analysis (1237.13 m<sup>2</sup>/g, 0.66723 cm<sup>3</sup>/g and 2.157 nm, values for surface area, pore volume and pore diameter respectively).

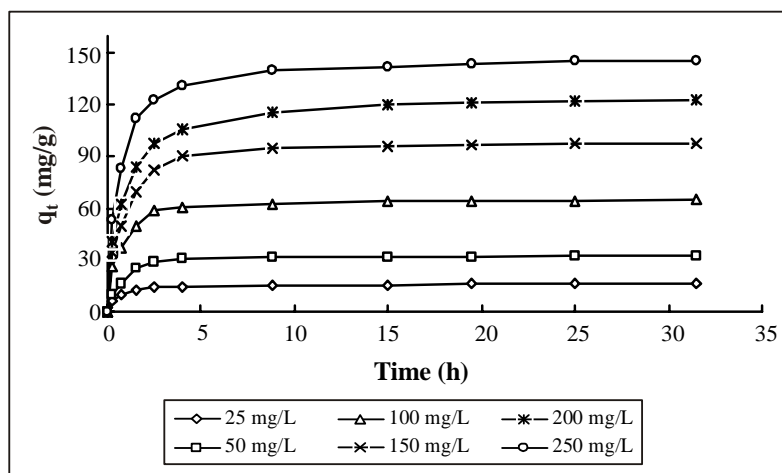


**Fig. 1: Surface morphology of the PFAC (magnifications: 1000×)**

### Effect of initial concentration and agitation time on carbofuran adsorption

The effect of carbofuran initial concentration and agitation time on the carbofuran adsorption onto the PFAC is shown in Fig. 2. It is clear that the amount of carbofuran adsorbed,  $q_t$ , increased onto PFAC surface. The adsorption uptake at equilibrium was found to increase with an increase in the initial insecticide concentration as appear in Fig. 2, which shows that longer contact times were required to reach equilibrium by the carbofuran solutions of higher initial concentrations.

The contact time needed for carbofuran solutions with initial concentration of 25-150 mg/L to reach equilibrium on PFAC was around 2-4 h; at the mean time the contact times for higher initial concentrations (200-250 mg/L) equilibrium times of 8-10 h were required.



**Fig. 2: Effect of carbofuran initial concentration and agitation time on carbofuran adsorption capacity**

Adsorption of carbofuran was fast due to the high affinity of the interacting groups on the surface of the activated carbon. The high adsorption rate at the beginning of adsorption was due to the adsorption of carbofuran to the exterior surface of the adsorbent. The high adsorption uptake of activated carbons prepared in this work were due to the presence of functional groups such as hydroxyl, carbonyl which dissociate and hence the electrostatic attraction between the activated carbon surface and insecticide. Similar trend has been reported for carbofuran adsorption onto carbon slurry<sup>19</sup>. The rate of uptake is rapid in the beginning and the rate of adsorption was found to depend on the initial concentration of insecticide.

### Effect of solution pH on carbofuran adsorption

The effect of pH on carbofuran adsorption was studied by varying the pH from 2 to 12 using 200 mL of a 100 mg/L fixed initial concentration of carbofuran at 30°C. The equilibrium adsorption of carbofuran was found to decrease slightly when the initial pH of the aqueous solution was increased from 2 to 12 (Figure not shown). This may be due to the presence of excess H<sup>+</sup> ions which accelerates the removal of the carbofuran with the anion OH<sup>-</sup> in the aqueous solution. It is also possible that the surface properties of the activated carbon have been altered as a result of the pH of the solution. Thus, the surface charge would depend on the solution pH and the surface characteristics of the carbon<sup>20</sup>.

### Adsorption isotherm

The equilibrium data for carbofuran adsorption on PFAC were modeled with three linearized expressions of the Langmuir, the Freundlich and the Temkin isotherm models (Figures not shown). Table 1 summarizes all the constants and correlation coefficients,  $R^2$  of these three isotherm models at 30°C. The Langmuir model yielded the best fit with  $R^2$  which were higher than 0.993. The monolayer adsorption capacity according to Langmuir model was 163.9 mg/g.

The comparison of adsorption capacity of carbofuran onto PFAC with other activated carbon found in literature carbon slurry (80 mg/g)<sup>21,22</sup>, which indicates that the PFAC is a more effective adsorbent for carbofuran adsorption.

**Table 1: Langmuir, Freundlich and Temkin isotherm model parameters and correlation coefficients for adsorption of carbofuran on PFAC at 30°C**

Isotherm models	Parameters		
Langmuir	$q_m$ (mg/g)	$b$ (L/mg)	$R^2$
	163.9	0.236	0.993
Freundlich	$K_F$ (mg/g (L/mg) <sup>1/n</sup> )	1/n	$R^2$
	31.31	0.521	0.887
Temkin	$A$ (L/g)	$B$	$R^2$
	1.13	32.51	0.958

## Regeneration of activated carbon

PFAC that adsorbed carbofuran was regenerated by ethanol. The desorption of carbofuran from spent PFAC was repeated for four cycles for adsorption and four cycles for desorption using the same activated carbon for the starting cycle. The regeneration efficiency was found to be 90 – 96.4%. This result indicates that the prepared activated carbon has a good regeneration and reusability characteristics for the adsorption of carbofuran and can be used as an alternative to the presently available commercial activated carbons.

## CONCLUSION

This work examined the feasibility of activated carbon prepared from oil palm fronds for the adsorption of carbofuran insecticide from aqueous solutions over a wide range of concentrations. It was found that the PFAC was very effective for this purpose. Equilibrium data were fitted to the Langmuir, Freundlich and Temkin isotherms and the equilibrium data were best described by the Langmuir isotherm model, with the maximum monolayer adsorption capacity of 163.9 mg/g. Ethanol desorption technique was efficient in regenerating the spent activated carbon and this provides a good ground for the reusability of the PFAC in subsequent adsorption runs.

## REFERENCES

1. E. Ayrañci and N. Hoda, *Chemosph.*, **57**, 755- 762 (2004).
2. T. Kumazawa and O. Suzuki, *J. Chromatog.*, **747**, 241-254 (2000).
3. F. J. Benitez, J. L. Acero and F. J. Real, *J. Hazard. Mater.*, **89**, 51-65 (2002).
4. S. P. Kale, N. B. K. Nurthy and K. Raghu, *Chemosph.*, **44**, 893-895 (2001).
5. J. M. Brasel, A. C. Collier and A. Pritsos, *Toxico. Appl. Pharm.*, **219**, 241-246 (2007).
6. World Organization WHO, [Online], (2004), Available from World Wide Web.
7. Sask Water L, [Online] (2008), Available from World Wide Web: <http://www.saskh2o.ca/>
8. M. Gavrilescu, *Eng. Life Sci.*, **5**, 497-526 (2005).
9. R. Broséus, S. Vincent, K. Aboulfadl, A. Daneshvar, S. Sauvé, B. Barbeau and M. Prévost, *Water Res.*, **43**, 4707-4717 (2009).



10. Y. Lu, D. Wang, C. Ma and H. Yang, *Build. Environ.*, **45**, 615-621 (2010).
11. L. Yeasmin S. A. and B. D. Wagner, *J. Photochem. Photobio. Chem.*, **204**, 217-223 (2009).
12. R. Rajeswari and S. Kanmani, *Desalination*, **242**, 277-285. 217-223 (2009).
13. J. M. Salman, V. O. Njoku and B. H. Hameed, *Chem. Eng. J.*, **173**, 361-368 (2011).
14. L. Liu, Z. Zheng and F. Yang, *Chem. Eng. J.*, (2009), Article in Press.
15. V. R. Midathana and V. S. Moholkar, *Indus. Eng. Chem. Res.*, **48**, 7368-7377 (2009).
16. A. Demirbas, *J. Hazard. Mater.*, **167**, 1-9 (2009).
17. I. Langmuir, *J. Am. Chem. Soc.*, **40**, 1361-1403 (1918).
18. H. Freundlich, *Z. Phys. Chem.*, **57**, 384-470 (1906).
19. M. J. Temkin, V. Pyzhev, *Acta Physiochim.*, **12**, 217-222 (1940).
20. W. Tanthapanichakoon, P. Ariyadejwanich, P. Japthong, K. Nakagawa, S. R. Mukai and H. Tamon, *Water Res.*, **39**, 1347-1353 (2005).
21. V. K. Gupta, I. Ali and V. K. Saini, *J. Coll. Interface Sci.*, **299**, 556-563 (2006).
22. M. Sathishkumar, A. R. Binupriya, D. Kavitha, R. Selvakumar, R. Jayabalan, J. G. Choi and S. E. Yun, *Chem. Eng. J.*, **147**, 265-271 (2009).
23. V. K. Gupta, C. K. Jain, I. Ali, S. Chandra and S. Agarw, *Water Res.*, **36**, 2483-2490 (2002).

*Revised : 21.02.2012*

*Accepted : 22.02.2012*